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# Micropatterning MoS<sub>2</sub>/Polyamide Electrospun Nanofibrous Membranes Using Femtosecond Laser Pulses

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**Abstract:** The capability of modifying and patterning the surface of polymer and composite materials is of high significance for various biomedical and electronics applications. For example, the use of femtosecond (fs) laser ablation for micropatterning electrospun nanofiber scaffolds can be successfully employed to fabricate complex polymeric biomedical devices, including scaffolds. Here we investigated fs-laser ablation as a flexible and convenient method for micropatterning polyamide (PA6) electrospun nanofibers that were modified with molybdenum disulfide (MoS<sub>2</sub>). We studied the influence of the laser pulse energy and scanning speed on the topography of electrospun composite nanofibers, as well as the irradiated areas via scanning electron microscopy and spectroscopic techniques. The results showed that using the optimal fs-laser parameters, micropores were formed on the electrospun nanofibrous membranes with size scale control, while the nature of the nanofibers was preserved. MoS<sub>2</sub>-modified PA6 nanofibrous membranes showed good photoluminescence properties, even after fs-laser microstructuring. The results presented here demonstrated potential application in optoelectronic devices. In addition, the application of this technique has a great deal of potential in the biomedical field, such as in tissue engineering.

**Keywords:** electrospinning; MoS<sub>2</sub>; composite nanofibers; femtosecond laser; micromachining

## 1. Introduction

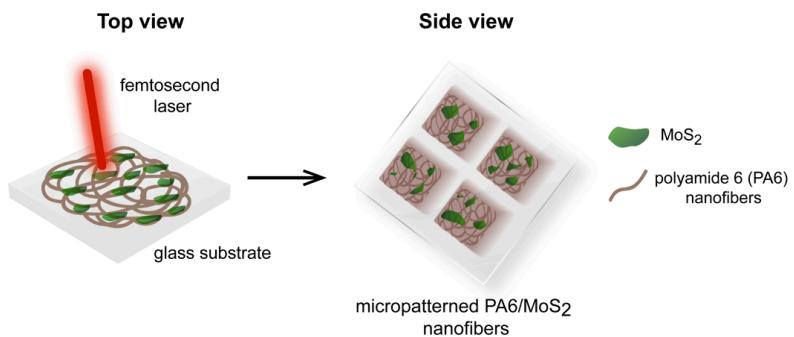
Over the past years, two-dimensional transition metal dichalcogenide (TMD) nanomaterials, a novel inorganic analogous of graphene, have drawn increasing attention due to their potential application in different areas [1–3]. In order to optimize their performance, or even potentially generate new functions, considerable efforts have been made to design functional hybrid TMD nanostructures [4]. To this end, significant work has been pursued to integrate some of the unique properties of TMDs into polymeric materials, aiming at varied biomedical applications [1,2,5–9]. For example, a very recent study demonstrated that incorporating TMDs into electrospun nanofibers was a suitable approach for building biomimetic cellular scaffolds [10]. The final composite nanofibers showed some key characteristics of TMDs, such as a large surface area and good mechanical

strength [8,11], while the fibrous features of the electrospun scaffold—which mimic the extracellular matrix—improved cellular adhesion and proliferation [12–15].

Within the biomedical context, researchers have used different methodologies in attempts to design nano/microscale patterns on prefabricated nanofibrous scaffolds to improve cell attachment, proliferation, spreading, and tissue development on composite nanofibers [15–17]. Femtosecond laser (fs-laser) micromachining is a well-known, precise fabrication technique that induces negligible thermal stress or collateral damage onto target materials, using very short time scales in the laser/material interaction [18–21]. The ultrashort pulse duration of fs-laser minimizes heat diffusion into the surrounding materials, and thus provides advantages over other processing techniques for electrospun nanofibers micromachining [22–24]. The processing of nanofibrous scaffolds by fs-laser is a flexible and rapid way to modify and control the surface topography by producing specific microenvironments that can influence cellular growth patterns [25,26]. In addition, the process can be carried out in a non-sterile environment, which can reduce the cost of the final product, thereby making the device fabrication process more economical [27]. For instance, Jun and coworkers fabricated hierarchically-patterned poly(L-lactic acid) nanofibrous scaffolds using a combination of electrospinning and femtosecond laser technique. They demonstrated that the laser-ablated microscale grooves on the surface of the nanofibrous membranes influenced the behavior of cardiomyoblasts—including adhesive morphology, proliferation, migration, and differentiation—when they were simultaneously stimulated by multiple topographical cues, including scale, oriented direction, and spatial arrangements [26].

Therefore, the use of fs-laser micromachining is an attractive approach for many applications. In fabricating biomedical devices, the aforementioned approach provides a convenient and economical solution because of its ability to produce programmable shapes on surfaces in a single processing step, while easily maintaining the sterility of the material [28]. Ultrafast lasers are also considered promising tools to rapidly process and create complex structures on 2D materials [29,30]. However, to the best of our knowledge, the use of fs-laser micromachining in TMD-composite electrospun nanofibers has not yet been reported. Molybdenum disulfide ( $\text{MoS}_2$ ), a typical semiconductor of the family of TMDs [5], is an indirect bandgap semiconductor in bulk form. As its thickness decreases, it becomes a direct bandgap semiconductor, which enhances photoluminescence [31].

In this regard, the present work aimed to evaluate and characterize the effectiveness of using fs-laser for micromachining the surface of electrospun nanofibers modified with  $\text{MoS}_2$ . We explored the potential of fs-laser ablation to create desired microscale topographical features on polyamide (PA6) nanofibrous scaffolds that were prepared using an electrospinning apparatus; they were then further modified with  $\text{MoS}_2$ , as illustrated in Scheme 1. Using optical microscopy, we then studied the influence of pulse energy and scanning speed on the produced features to determine the optimum irradiation parameters and the energy threshold for material removal. Later, we created patterns on the sample surfaces in two different ways—by functionalizing the PA6 nanofibers with  $\text{MoS}_2$  before and after the microfabrication in order to evaluate the photoluminescence of these structures. We believe that the microstructured  $\text{MoS}_2$  composite nanofibrous membranes provide new insight into the preparation of well-defined nanomaterials and offer significant potential in biomedical applications, such as antibacterial treatment [32], membrane-based separation [33], and photocatalysis [34].



**Scheme 1.** Schematic representation of molybdenum disulfide ( $\text{MoS}_2$ )/polyamide electrospun nanofibrous membranes, micropatterned using femtosecond (fs) laser ablation.

## 2. Materials and Methods

### 2.1. Materials

Polyamide 6 (PA6,  $M_w = 20,000 \text{ g mol}^{-1}$ ) and molybdenum disulfide ( $\text{MoS}_2$ ) were purchased from Sigma-Aldrich. The formic acid was purchased from Synth Chemical (São Paulo, Brazil). All aqueous solutions were prepared with double-distilled water, and the chemicals were used without further purification.

### 2.2. Preparation of PA6- $\text{MoS}_2$ Nanofibers

PA6 was dissolved in formic acid (95%) under vigorous stirring for 2 h at room temperature to prepare a 10% (*w/v*) polymer solution. The electrospun nanofibers were obtained with an electrospinning apparatus using a feed rate of  $0.01 \text{ mL h}^{-1}$  and an electric voltage of 20 kV. A working distance of 10 cm was kept between the syringe and the metallic collector. The inner diameter of the steel needle was 0.7 mm. Nanofibers were directly electrospun onto glass substrates. After the PA6 nanofibers fabrication, they were surface-functionalized by drop-casting an  $\text{MoS}_2$  aqueous dispersion ( $1.0 \text{ mg mL}^{-1}$ ) onto them.

### 2.3. Femtosecond Laser Micromachining and Characterization

The electrospun nanofibers membranes were micromachined using a femtosecond laser oscillator (Femtosource XL) delivering 50-fs pulses centered at around 800 nm, with a repetition rate of 5 MHz. The micromachining was carried out by focusing the femtosecond laser pulses onto the sample, with the aid of a microscope objective (0.65 NA—beam spot diameter at focus of  $\sim 3 \mu\text{m}$ ). The sample's position was controlled by an x-y-z translational stage, moving with constant speed to allow the microfabrication of arbitrary geometries onto the nanofibrous membrane. The entire fabrication process was monitored with a charge-coupled device (CCD) camera, in alignment with the microscope objective. The pulse energy and scanning speed were varied to determine the optimal irradiation parameters.

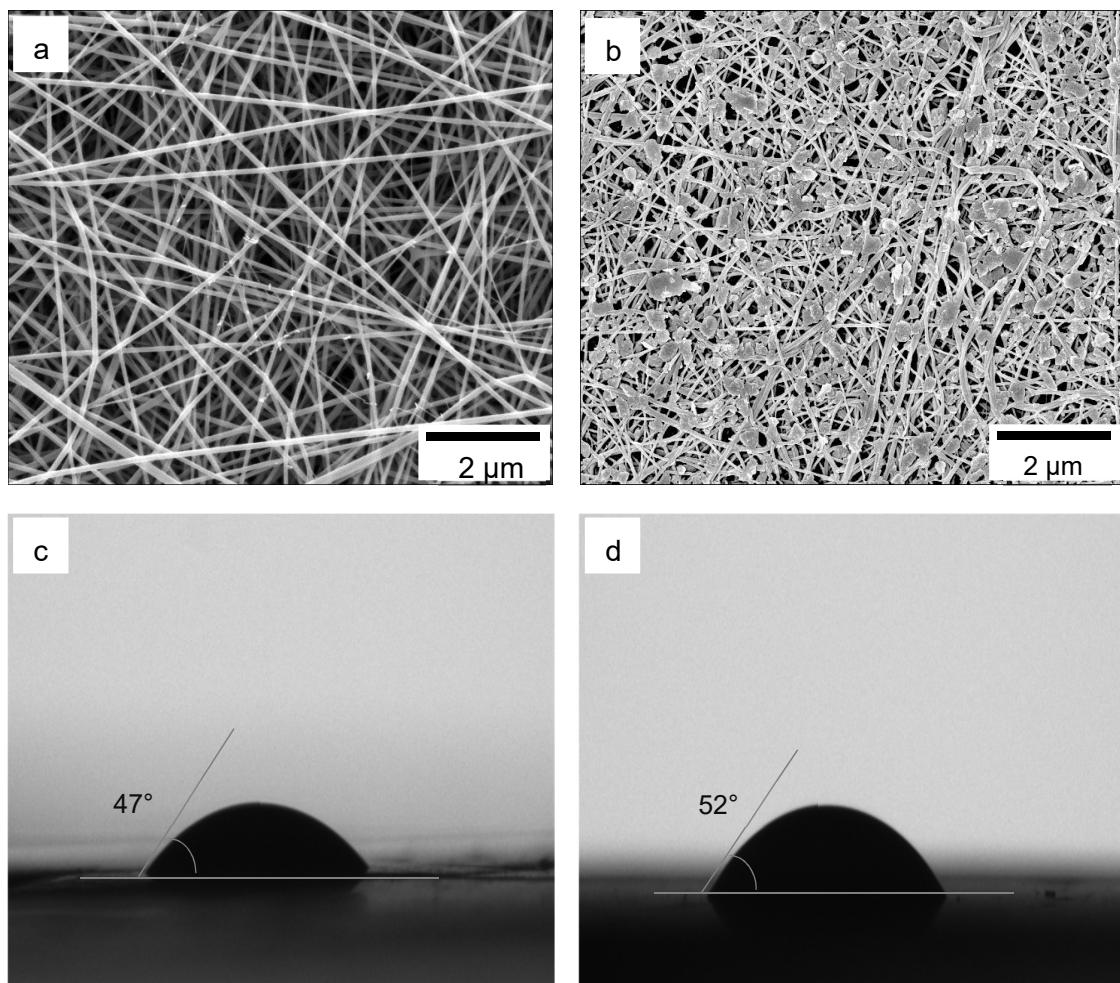
To confirm the presence of  $\text{MoS}_2$ , Raman spectroscopy was applied using a LabRAM micro-Raman system, with a solid-state laser operating at 633 nm as the excitation source. To avoid thermal effects, a 1 mW laser power was employed. To verify the nature of the  $\text{MoS}_2$  surface, contact angle measurements were performed using a KSV CAM 200 goniometer. To check the absorption properties, UV-Vis absorption spectroscopy was performed in the  $\text{MoS}_2$  aqueous dispersion using a Shimadzu UV-1800<sup>®</sup> spectrophotometer. The micromachined samples were then characterized by atomic force microscopy (AFM) using a Nanosurf's easyScan 2<sup>®</sup> to determine the depth of ablated lines. The samples were also analyzed by optical microscopy (Zeiss LSM 700), scanning electron microscopy (SEM) using FEI's Inspect F50 (Eindhoven, The Netherlands), and energy-dispersive X-ray (EDX) spectroscopy. The EDX spectrum was acquired using a field emission gun (FEG)-SEM (JEOL, JSM-6510). Finally,

the Zeiss LSM 700 was used to obtain confocal fluorescence microscopy using a  $40\times$  objective and excitation at 639 nm with a  $1\text{ }\mu\text{m}$  of z-step.

### 3. Results and Discussion

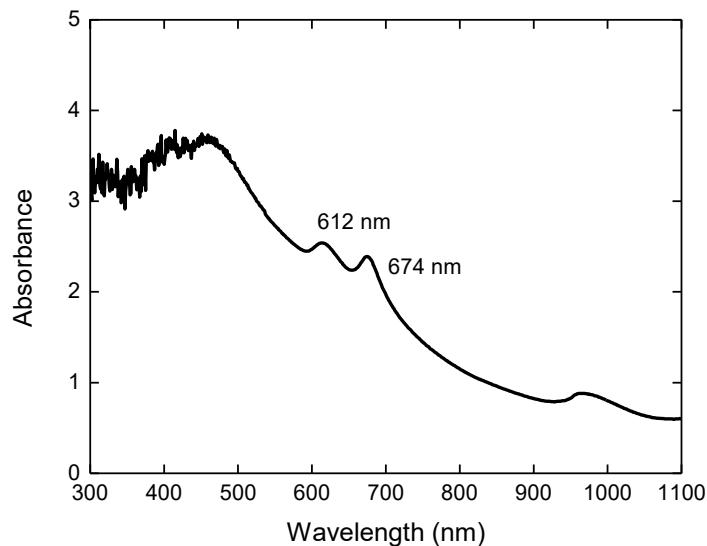
Figure 1a shows SEM images of the PA6 nanofibers prepared by electrospinning. The obtained nanofibers exhibit a smooth surface and regular cylindrical morphology, which arises from the fast evaporation of the solvent during the electrospinning process, leading to the formation of a solid skin that shrinks and collapses upon the evaporation of the remaining solvent [35]. The average fibers diameter was determined to be  $75 \pm 10\text{ nm}$ . As confirmed by the SEM image in Figure 1b, which displays the presence of  $\text{MoS}_2$  flakes, the drop-casting method efficiently functionalized the PA6 nanofibers.

Figure 1c,d show the water contact angles of nanofibrous membranes composed of net PA6 and PA6 functionalized with  $\text{MoS}_2$ , respectively, with both angles being within a partial wetting regime ( $<90^\circ$ ):  $(47 \pm 1)^\circ$  for PA6 and  $(52 \pm 1)^\circ$  for PA6- $\text{MoS}_2$  nanofibers. For both materials, the contact angle decreased rapidly over time, and within 15 s, the samples showed full wetting behavior. This transition is consistent with water absorption into the fiber network [36]. These results are also in agreement with the literature, once the  $\text{MoS}_2$  also presents hydrophilic behavior [37–39].



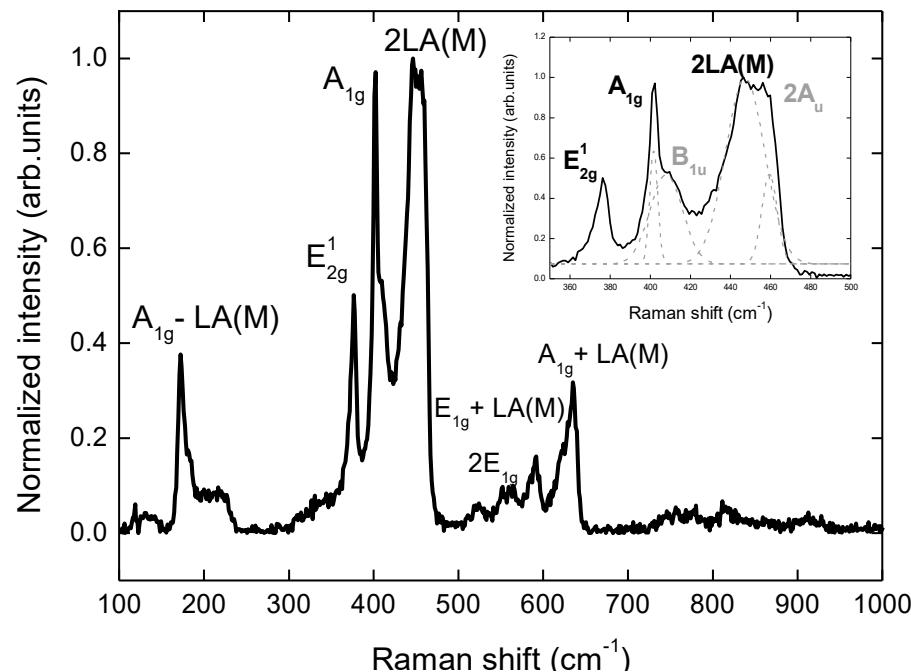
**Figure 1.** Scanning electron microscopy (SEM) images and photographs of water contact angle measurement of (a,c) polyamide (PA6) and (b,d) PA6- $\text{MoS}_2$  nanofibrous membranes.

The typical UV-Vis absorption spectrum of  $\text{MoS}_2$  dispersion in water can be observed in Figure 2. The two absorption bands, centered at 612 and 674 nm, corresponds to exciton B and A, respectively [40,41], and are characteristic of  $\text{MoS}_2$ .



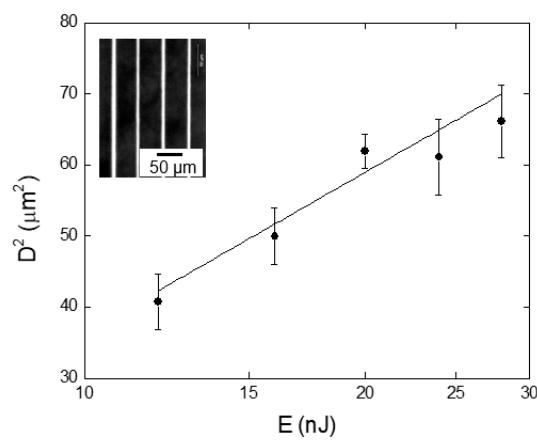
**Figure 2.** UV-Vis absorption spectrum of  $\text{MoS}_2$  dispersion in water.

Figure 3 shows the room temperature Raman spectrum of PA6 nanofibers functionalized with  $\text{MoS}_2$ . The sample was excited with laser light at 633 nm (1.96 eV), which closely matches the energy of the direct bandgap of  $\text{MoS}_2$  at room temperature [42]. Four first-order Raman active modes dominate the resonant Raman spectrum of  $\text{MoS}_2$ , in which  $\text{E}_{1g}$ ,  $\text{E}^1_{2g}$ ,  $\text{A}_{1g}$  and  $\text{E}^2_{2g}$  (not seen in Figure 3) are a result of vibrational modes within the  $\text{MoS}_2$  layer [43]. The  $\text{E}^1_{2g}$  and  $\text{A}_{1g}$  modes of the exfoliated  $\text{MoS}_2$  nanosheets were centered at  $379 \text{ cm}^{-1}$  and  $404 \text{ cm}^{-1}$ , respectively [44]. In low wavenumbers, at  $175 \text{ cm}^{-1}$ , the observed peak was assigned to the two-phonon difference combination mode ( $\text{A}_{1g} - \text{LA}(\text{M})$ ) [45]. At wavenumbers between  $450$  and  $650 \text{ cm}^{-1}$ , there were observed modes at  $450 \text{ cm}^{-1}$ ,  $571 \text{ cm}^{-1}$ ,  $590 \text{ cm}^{-1}$ , and  $642 \text{ cm}^{-1}$ ; these were assigned as  $2\text{LA}(\text{M})$ ,  $2\text{E}_{1g}$ ,  $\text{E}_{1g} + \text{LA}(\text{M})$ , and  $\text{A}_{1g} + \text{LA}(\text{M})$ , respectively, as shown in Figure 3. These modes were mostly of the second order, non-zone center phonons (M-point) or their combination [46]. By performing a Gaussian decomposition, as shown in the inset of Figure 3, the first-order optical phonon peak  $\text{A}_{2u}$  at  $459 \text{ cm}^{-1}$  was observed, and at  $457 \text{ cm}^{-1}$  the  $\text{B}_{1u}$  formed a pair with  $\text{A}_{1g}$  due to the resonant effect [42]. From the data presented in Figure 3, the relative intensity of the  $\text{E}^1_{2g}$  mode with respect to the  $\text{A}_{1g}$  mode was obtained as 0.58. Also, no broad band feature appeared in the spectrum for wavenumbers higher than  $700 \text{ cm}^{-1}$ , due to the luminescence present when the number of layers decreased [46], indicating the presence of multilayer  $\text{MoS}_2$  flakes in our sample, as expected [47].



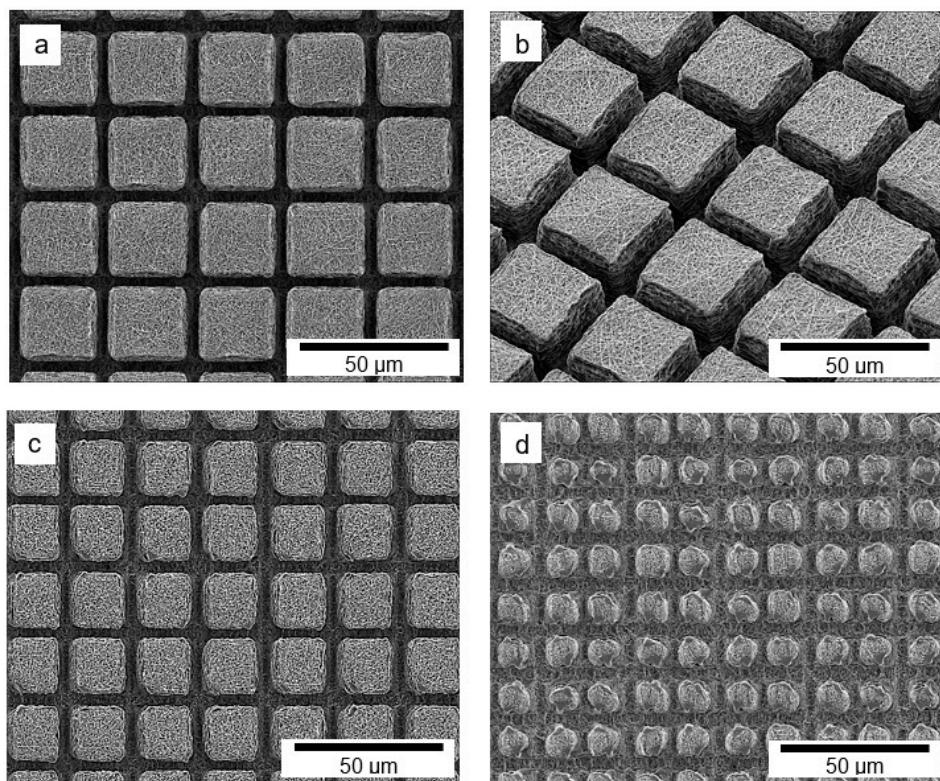
**Figure 3.** Resonant Raman spectra of  $\text{MoS}_2$  modified-PA6 nanofibrous excited using a 633 nm laser.

A set of experiments were performed to study the influence of the pulse energy and scanning speed on the fs-laser micromachining. Using a 0.65 NA microscope objective, lines at different pulse energies and scanning speeds were produced on the nanofibrous membranes and then analyzed by optical microscopy. A scanning speed of  $50 \mu\text{m/s}$  and pulse energies varying from  $12 \text{ nJ}$  to  $28 \text{ nJ}$  were used to micromachine groups of lines. As an illustration, the inset in Figure 4 shows an optical microscopy image of lines micromachined on PA6 nanofibrous membranes using fs-laser pulses with  $25 \text{ nJ}$ . The average line width ranged from  $6.8 \mu\text{m}$  to  $8.1 \mu\text{m}$ , as the pulse energy varied from  $12$  to  $28 \text{ nJ}$ . Figure 4 depicts the squared line width as a function of the pulse energy (log-scale). The solid line in this figure represents the fitting, according to the model described in Reference [18,48], from which we determined the threshold energy for material removal to be  $3.5 \text{ nJ}$ .



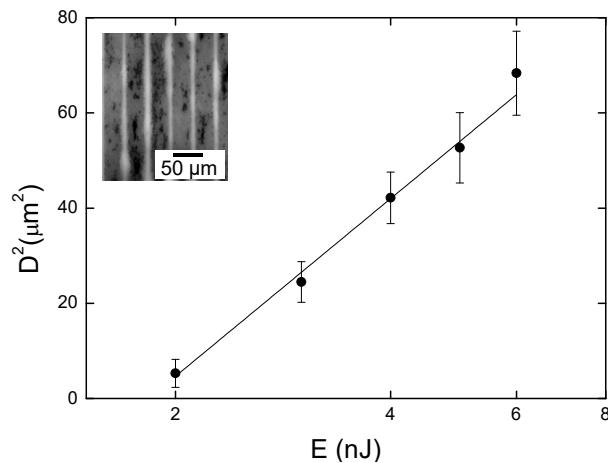
**Figure 4.** Squared line width as a function of the pulse energy for  $50 \mu\text{m/s}$  of translation speed. The inset shows an optical microscopy image of fs-laser micromachined lines using  $25 \text{ nJ}$ , in which the white lines represent the removed (laser-ablated) material and the black regions correspond to the remaining material.

Patterns were created onto the sample surface using optimized laser ablation parameters (20 nJ and 50  $\mu\text{m}/\text{s}$ ). An array of micro-square pillars was produced, with periodicities of 40, 30, and 20  $\mu\text{m}$ , 4  $\mu\text{m}$  high (which value was obtained by atomic force microscopy measure), and a spacing of 7.8  $\mu\text{m}$  between them. The SEM image of the micro-square pillars with 40  $\mu\text{m}$ , created by fs-laser ablation, is shown in Figure 5a (top view) and 5b (tilt angle of 45°), revealing the high resolution achieved by the fs-laser processing. It is important to mention that a decrease in periodicity led to a decrease in resolution with the appearance of many irregularities, as shown in Figure 5c,d for the periods of 30 and 20  $\mu\text{m}$ , respectively.



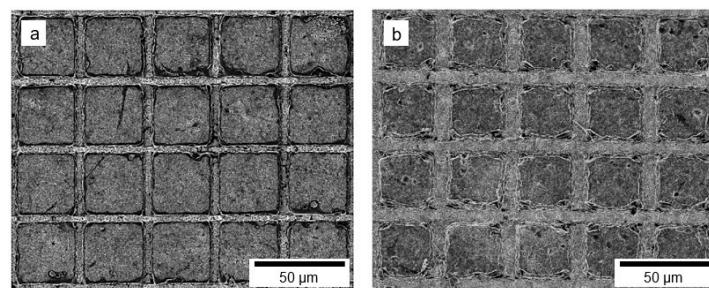
**Figure 5.** SEM images of micromachined pillars with a periodicity of 40  $\mu\text{m}$  (a) in the top view and (b) at a tilted angle of 45°, (c) 30  $\mu\text{m}$  and (d) 20  $\mu\text{m}$  obtained in the top view.

A new set of experiments was performed to study the influence of the pulse energy and scanning speed on the micromachined lines for the MoS<sub>2</sub>-modified PA6 nanofibrous membranes. Using the same experimental setup, lines at different pulse energies and scanning speeds were produced on the nanofibrous membranes and analyzed by optical microscopy. A scanning speed of 50  $\mu\text{m}/\text{s}$  and pulse energies varying from 2 nJ to 6 nJ were used to micromachine groups of lines (500  $\mu\text{m}$  long and separated by 40  $\mu\text{m}$ ) on the sample surface. The same behavior was observed for PA6–MoS<sub>2</sub> nanofibrous membranes; as the pulse energy (E) is increased, the width of the lines (D) also increased. At 50  $\mu\text{m}/\text{s}$ , the average widths ranged from 2.3  $\mu\text{m}$  to 8.2  $\mu\text{m}$  as the pulse energy varied from 2 to 6 nJ, respectively, producing lines with higher resolution. We also determined, by fitting the results in Figure 6 according to references [18,48], that the threshold energy for material removal was 1.8 nJ. This value is smaller than the one previously observed for the pure PA6 sample, likely due to the absorption of MoS<sub>2</sub> at the wavelength of the laser pulse used for micromachining. The inset in Figure 6 shows an optical microscopy image of lines micromachined on PA6–MoS<sub>2</sub> nanofibrous membranes using fs-laser pulses and energy of 5 nJ.



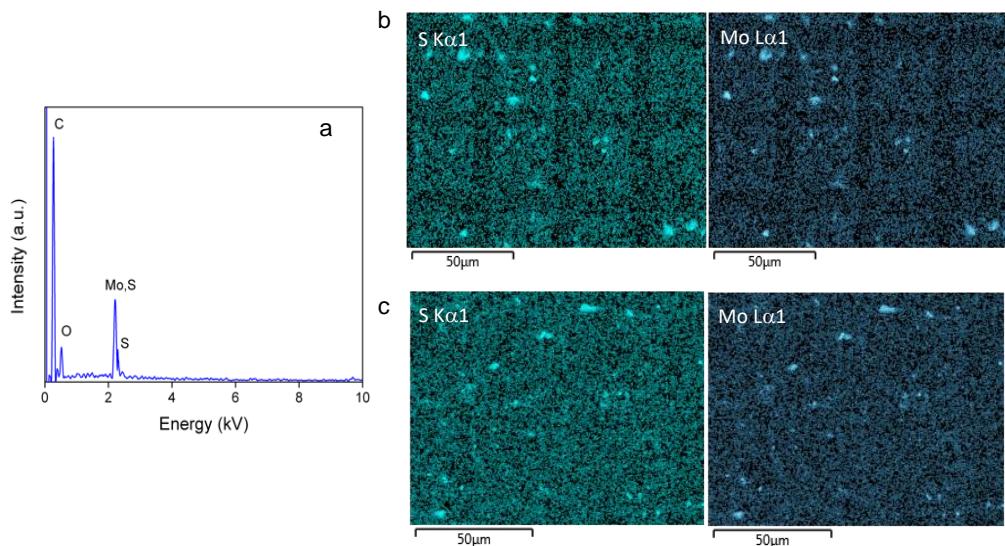
**Figure 6.** Squared line width as a function of the pulse energy for 50  $\mu\text{m}/\text{s}$  of translation speed. The inset shows an optical microscopic image of fs-laser micromachined lines using 5 nJ, in which the white lines represent the removed (laser-ablated) material and the gray regions correspond to the remaining material.

Therefore, two methods can be employed for the production of the patterns on the nanofibrous membranes: the first is by modifying the PA6 nanofibers with  $\text{MoS}_2$  before laser microfabrication (DBM), while the second one consists in modifying the nanofibers with  $\text{MoS}_2$  after microfabrication (DAM). Thus, for both approaches (DBM and DAM), matrices of micro-square pillars with a periodicity of 40  $\mu\text{m}$ , 4  $\mu\text{m}$  high, and a spacing of 6.3 and 7.8  $\mu\text{m}$  between the pillars were produced. The SEM images of the micro-square pillars with 40  $\mu\text{m}$  created by fs-laser ablation using DBM and DAM are shown in Figure 7a,b, respectively. From these images, one can see an increased resolution relative to the width of the lines, yielding to regular structures on the nanofibrous membranes, when the modification with  $\text{MoS}_2$  was performed before the microfabrication (DBM).



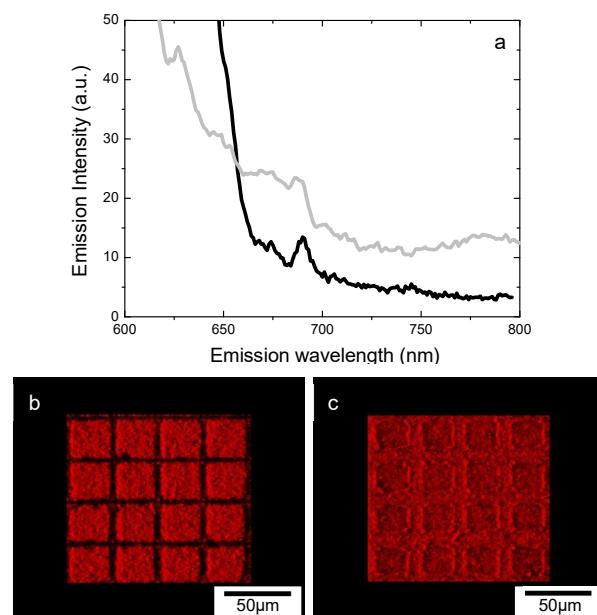
**Figure 7.** SEM images of micromachined pillars with a periodicity of 40  $\mu\text{m}$  obtained by modifying the PA6 nanofibers with  $\text{MoS}_2$  (a) before laser microfabrication (DBM) and (b) after laser microfabrication (DAM).

Typical EDX spectra and chemical element composition analysis of the surface of the microstructured pillars using both approaches (DBM and DAM) are shown in Figure 8. The chemical composition analysis reveals that the sample stoichiometry after fs-laser irradiation is the same as the original, with the S/Mo atomic ratio being 2.03, in agreement with the stoichiometry of  $\text{MoS}_2$ . Figure 8b,c present the mapping of backscatter images of the main elements present in samples micromachined using DBM and DAM microstructures, respectively. The mapping of the S and Mo elements for the DBM sample (see Figure 7b) indicated a uniform distribution of such elements. It is possible to observe a smaller amount of material in the laser line, as expected by the ablation of the material. In Figure 8c (DAM), we observed a homogeneous distribution of Mo and S on the sample surface, as expected, once the modification with  $\text{MoS}_2$  was performed on the entire surface of the material after micromachining.



**Figure 8.** (a) The energy-dispersive X-ray (EDX) spectrum, and elemental mapping of the nanofibers modified with MoS<sub>2</sub> (b) before laser microfabrication and (c) after laser microfabrication.

Figure 9a shows the photoluminescence emission spectrum of the MoS<sub>2</sub>-modified PA6 nanofibrous membranes at two different excitation wavelengths, 575 nm (gray line) and 639 nm (black line), both measured at room temperature. The observed luminescence emissions, called A and B excitons, arose from the direct excitonic transitions at the K point of the Brillouin zone [49–51]. The emission peaks at 690 nm and 627 nm correspond to the energy of 1.79 eV and 1.97 eV, respectively, which is quite consistent with the absorption displayed in Figure 2. Figure 9b,c show confocal fluorescence microscope images obtained using laser excitation at 639 nm for DBM and DAM microstructured surfaces with a periodicity of 40  $\mu$ m, respectively.



**Figure 9.** (a) Photoluminescence emission spectrum of the PA6–MoS<sub>2</sub> composite nanofibers at two different excitation wavelengths: the gray line at 575 nm and the black line at 639 nm. Confocal fluorescence microscope images obtained using laser excitation at 639 nm of micromachined pillars with a periodicity of 40  $\mu$ m for nanofibrous membranes modified with MoS<sub>2</sub> (b) before and (c) after laser microfabrication.

#### 4. Conclusions

MoS<sub>2</sub>-modified PA6 nanofibrous membranes have been successfully fabricated by electrospinning and characterized in terms of physical-chemical properties. The produced nanofibrous membranes were micropatterned using femtosecond laser pulses, and it was observed that the threshold energy for material removal dropped from 3.5 nJ to 1.8 nJ when the nanofibers were modified with MoS<sub>2</sub>. The electron microscopy images revealed that by controlling the laser pulse energy and scanning speed, micromachined pillars with a periodicity in the micrometer scale could be successfully fabricated. Additionally, photoluminescence emission revealed the typical fluorescence bands of MoS<sub>2</sub>, while confocal fluorescence images suggested that MoS<sub>2</sub> is distributed throughout the nanofibrous membrane surface. Our results indicate that the ability to modify the surface of the nanofibrous membrane with luminescent MoS<sub>2</sub> and patterning them with fs laser holds high potential for biomedical and electronics applications.

**Author Contributions:** C.R.M. and D.S.C. conceived this work; L.A.M. and R.S. produced the nanofibrous membranes and their modifications with MoS<sub>2</sub>. K.T.P. and L.A.M. performed the sample characterizations. K.T.P. carried out the laser micromaching, optical measurements and analyzed the results. C.R.M. and D.S.C. supervised the experiments. All authors discussed the results and substantially contributed to the manuscript writing.

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**Conflicts of Interest:** The authors declare no conflict of interest.

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