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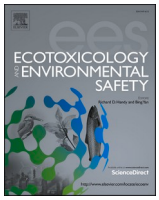

Corrigendum

Corrigendum to "Radiometric approaches with carbon-14-labeled molecules for determining herbicide fate in plant systems" [Ecotoxicol. Environ. Saf. 284 (2024) 117003]

Gustavo Vinícios Munhoz-Garcia [a,*,1], Vanessa Takeshita [a,*,1], Camila de Werk Pinácio [b], Brian Cintra Cardoso [b], Bruno Dalla Vecchia [b], Daniel Nalin [a], Ana Laura Camachos de Oliveira [b], Leandro Fernando Felix [b], Valdemar Luiz Tornisielo [a]

[a] Center of Nuclear Energy in Agriculture, University of São Paulo, Av. Centenário 303, Piracicaba, SP 13400-970, Brazil
[b] Superior School of Agriculture "Luiz de Queiroz", University of São Paulo, Av. Pádua Dias, 11, Piracicaba, SP 13418-900, Brazil

First correction

The quality of the figures in the final manuscript was reduced in a way that reduced the quality and readability of the manuscript. We provide higher quality figures in this document. We apologize for the inconvenience, but the journal does not allow a correction directly in the published paper.

Figures in higher quality

Second correction

The authors regret the following text for the supplementary information was previously published in a wrong format.

The authors would like to apologise for any inconvenience caused.

Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at doi:10.1016/j.ecoenv.2024.117172

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* Corresponding authors.
E-mail addresses: gvmgarcia@usp.br (G.V. Munhoz-Garcia), vanessatakeshita@usp.br (V. Takeshita).
[1] These authors contributed equally to the work.

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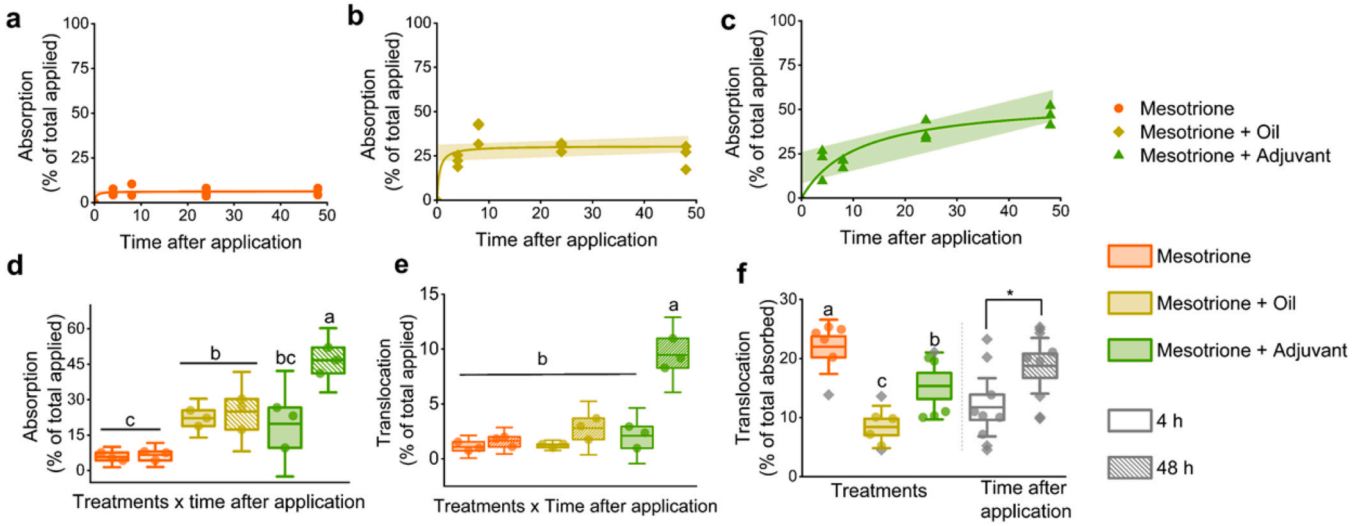

Fig. 1. Absorption (a–d) and translocation (e–f) of ^{14}C-mesotrione in *E. heterophylla* plants. In a, b, and c the line represents the absorption rate as a function of time, adjusted to the Michaelis-Menten model ($p < 0.05$), and the shaded band around the line represents the 95 % prediction band. In e, the lines represent a linear regression. In d and f, the box represents the mean ± SE and the bars represent 95 % of the confidence interval. Symbols represent the data and may cover the bars. Outliers are represented by gray symbols. Treatments followed by the same lowercase letter are not significantly different according to Tukey's test ($p < 0.05$). *Significant ($p < 0.05$).

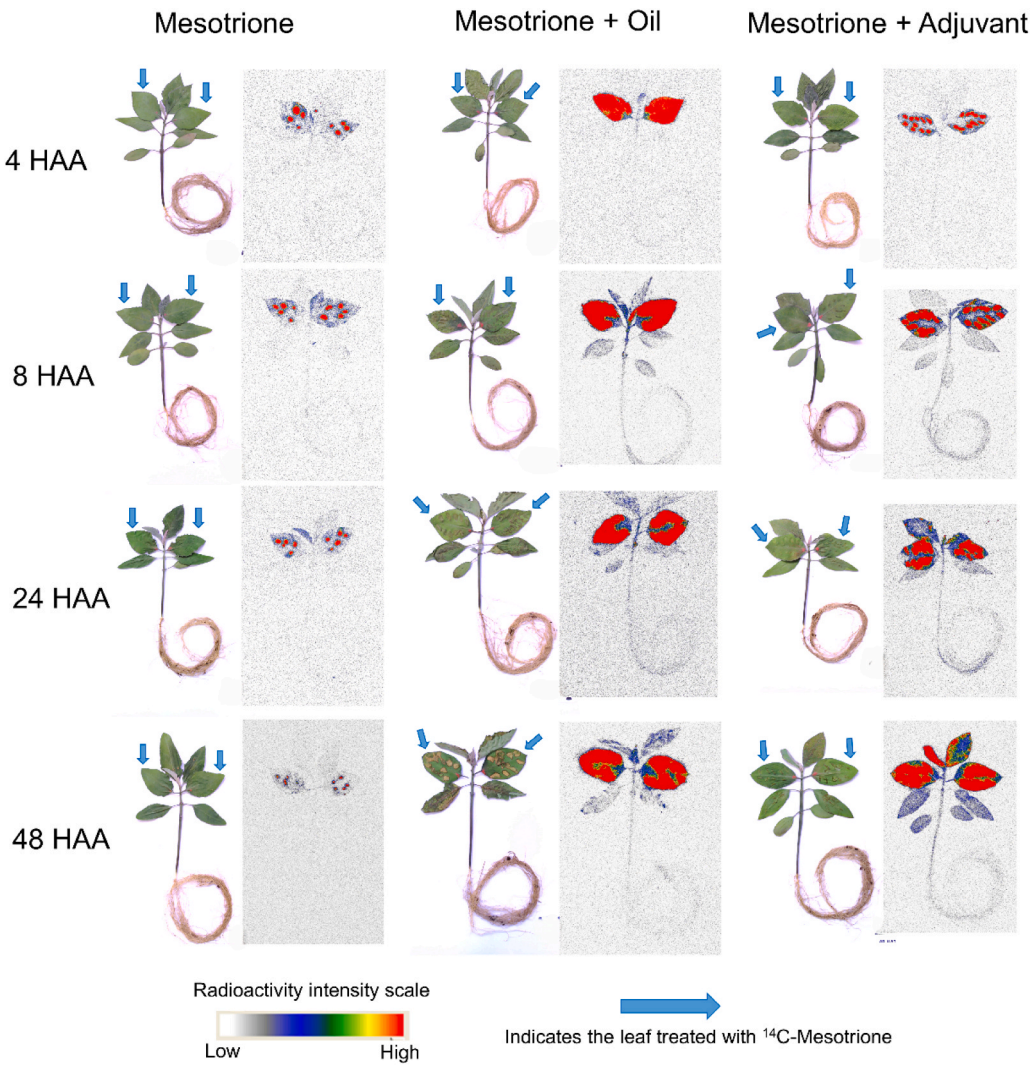

Fig. 2. Images (left) and autoradiography (right) of *E. heterophylla* plants submitted to ^{14}C-mesotrione application, alone or in combination with oil and adjuvant, as a function of time. The blue arrows indicate leaves treated with ^{14}C-mesotrione, the red color indicates a higher intensity of radioactivity, and the blue color indicates a lower intensity. HAA - hours after application.

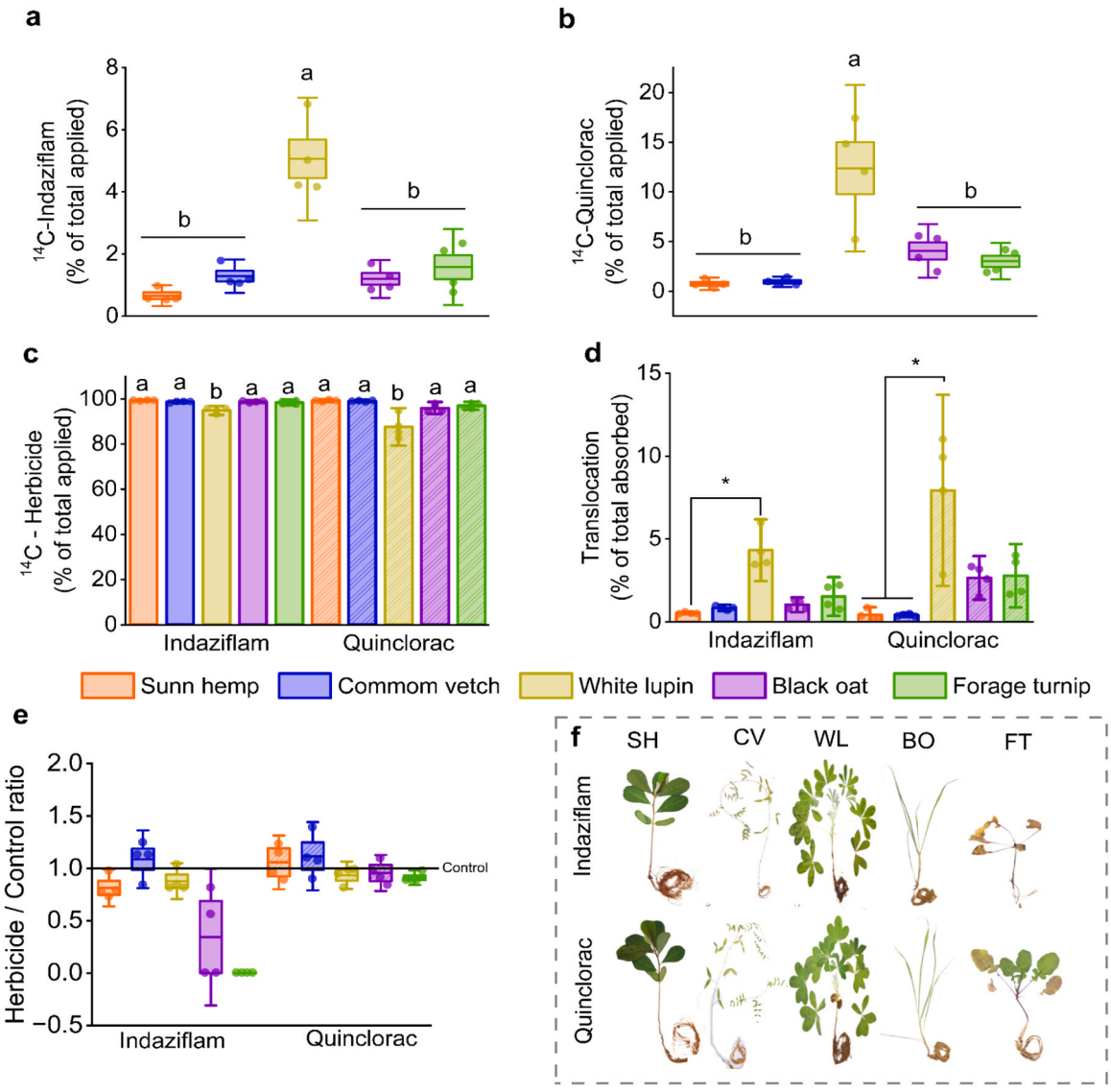

Fig. 3. Uptake of ^{14}C-indaziflam (a) and ^{14}C-quinclorac (b) from soil; herbicide remaining in the soil after cover crop growing (c); ^{14}C-herbicide translocation by cover crops (d); herbicide effect on plant height compared to the control (e) and cover crop images (f). The symbols represent the data, and the bars represent 95 % of the confidence interval. Symbols can cover the bars. In a, b, and e the box represents the mean ± SE. Boxes or bars followed by the same lowercase letter are not significantly different according to Tukey's test ($p < 0.05$). Asterisks indicate a significant difference according to Dunn's test ($p < 0.05$). SH - sunn hemp, CV - common vetch, WL - white lupin, BO - black oat, and FT - forage turnip.

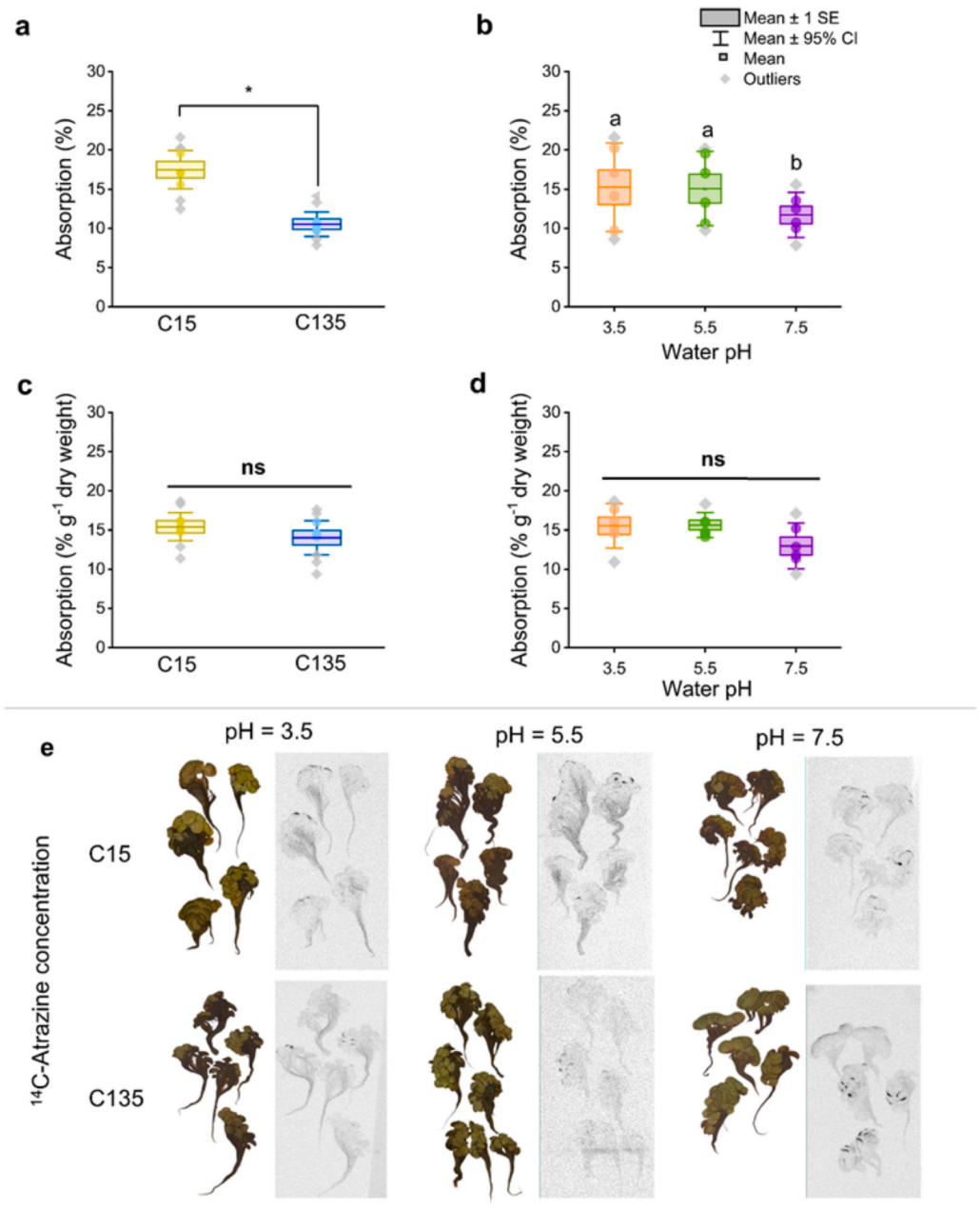

Fig. 4. Absorption of ^{14}C-atrazine from water under different concentrations (a) and water pH (b), concerning the total initially applied. The absorption data were normalized to plant dry weight (c and d). Images (left) and autoradiography (right) of *Salvinia* spp. plants 21 days after contact with ^{14}C-atrazine in water (e); the higher intensity of radioactivity is represented by the black color in autoradiography images. The symbols represent the data, and the bars represent 95 % of the confidence interval. Symbols can cover the bars. Outliers are represented by gray symbols. Boxes followed by the same lowercase letter are not significantly different according to Tukey's test ($p < 0.05$). Ns - non-significant ($p > 0.05$). C15 - 15 μg L^{-1}, C135 - 135 μg L^{-1}.

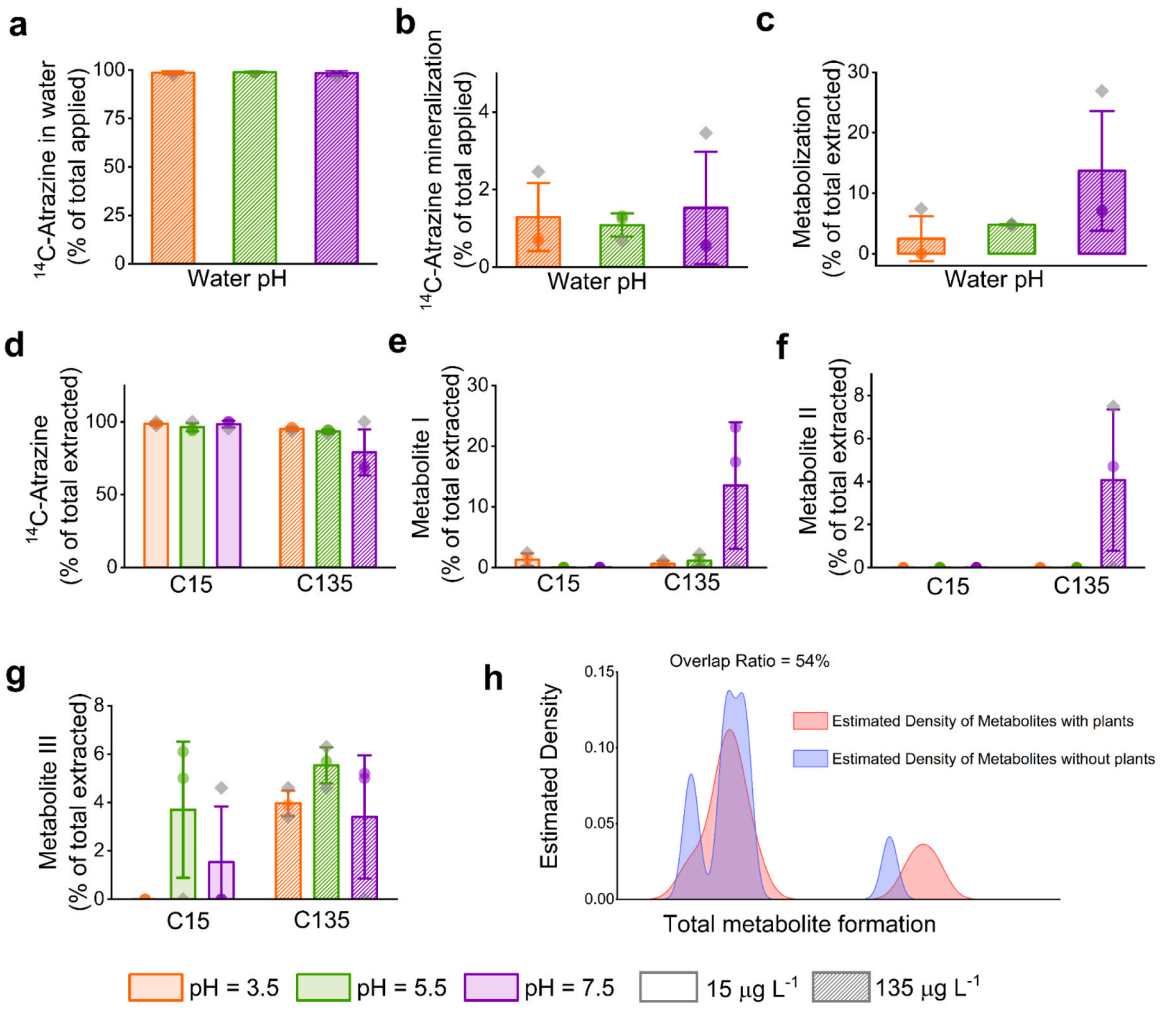

Fig. 5. Mineralization and degradation study of ^{14}C-atrazine in water. ^{14}C-atrazine quantified in water (a), total mineralized to $^{14}CO_2$ (b), and metabolized in biometric flasks (c), without the plants. Total atrazine remaining in water (d) and the amount of each metabolite formed in water in the presence of the *Salvinia* spp. plants (e–g). Similarity between the atrazine degradation in the presence and absence of plants (h) according to Kernel's density estimation. The symbols represent the data and the bars represent 95 % of the confidence interval. Symbols can cover the bars. Outliers are represented by gray symbols.

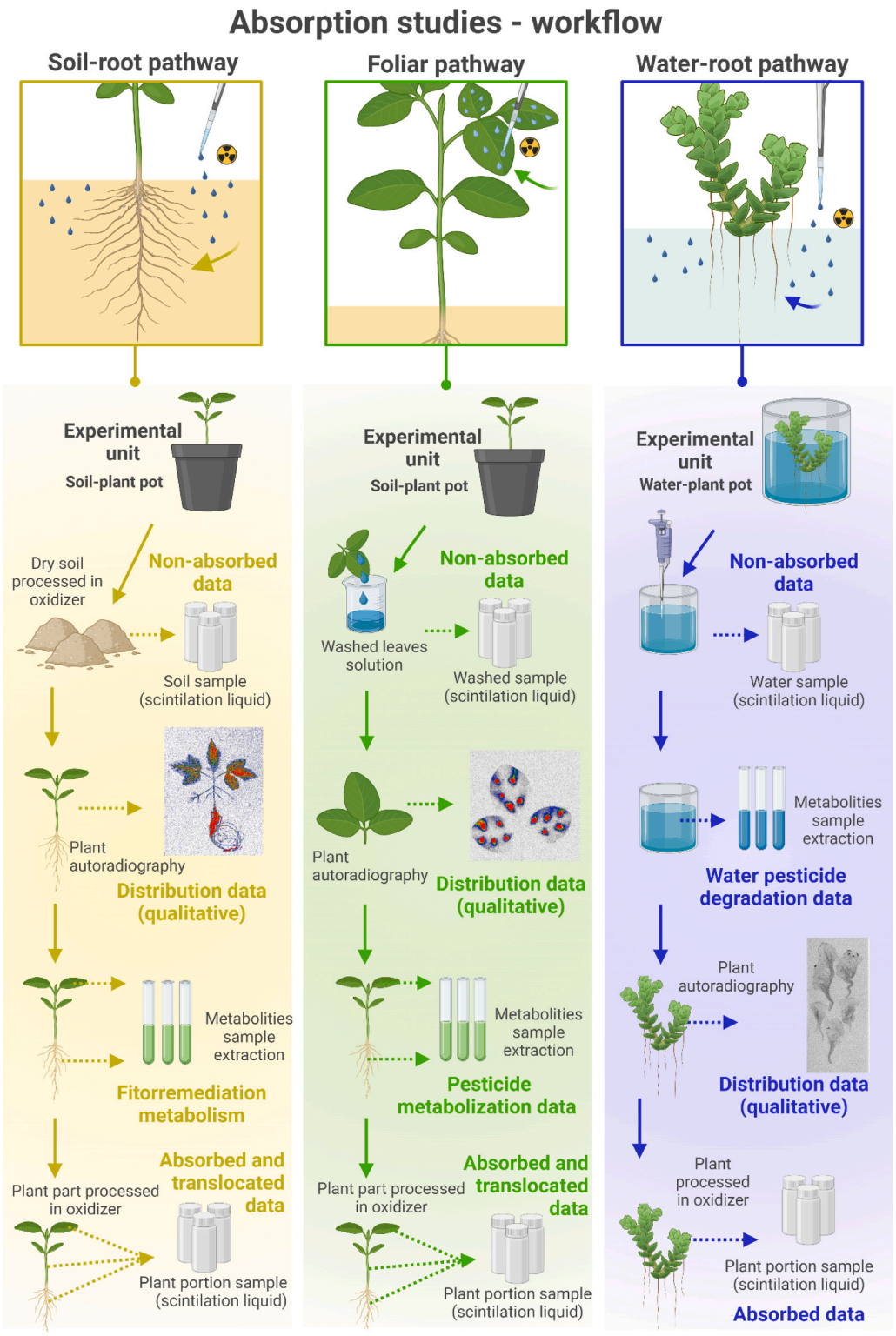

Fig. 6. General overview of absorption and translocation studies using radiometric techniques.