

# New record and redescription of invasive cassava green mite, *Mononychellus mcgregori* (Flechtmann & Baker) in India on cassava, *Manihot esculenta* Crantz

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## Original research

### ABSTRACT

The unnoticed invasion of mites is attributed to the absence of consistent monitoring over time. This recent investigation identified the presence of the cassava green mite, *Mononychellus mcgregori* (Flechtmann & Baker 1970), on Cassava (*Manihot esculenta* Crantz) for the first time in the Indian subcontinent. The study involves a comprehensive re-description of the invasive mite, *Mononychellus mcgregori* (Flechtmann & Baker), employing an integrative approach. The report also highlights the potential future threat posed by the invasive mite to crop diversity in India.

**Keywords** alien mites; molecular confirmation; morphological identification; spider mite

## Introduction

Cassava (*Manihot esculenta* Crantz), commonly known as Tapioca, continues to be a crop of food security for millions of people, especially in developing countries across the globe. It is an essential alternate energy source to meet the increasing population's demands. Originating in tropical America, cassava was introduced to southern peninsular India in the 17th century by the Portuguese, primarily in Kerala, before spreading to neighbouring states like Tamil Nadu, Andhra Pradesh, and Karnataka. Currently, India cultivates cassava across 172 thousand hectares, yielding 6.257 million tonnes. Tamil Nadu and Kerala lead in production, contributing 3.4 million tonnes and 2.5 million tonnes, respectively, primarily supporting employment through starch extraction and sago export (India Stat 2023). In recent times Cassava pest complexes have emerged and pose a threat to Cassava production in Asian countries. The major arthropod complex is increasing too significantly in coevolved specific pests of cassava. Among the pests, the cassava mealybug (*Phenacoccus manihoti*) and cassava green mites (*Mononychellus* spp.) complexes stand first (Bellotti *et al.* 2012; Graziosi *et al.* 2016). *Mononychellus* mite complex includes *M. tanajoa* (Bondar, 1938), *M. mcgregori* (Flechtmann & Baker, 1970) and *M. caribbeanae* (McGregor, 1950) where these mites are highly host-specific and invasive threats to cassava growing areas. The genus *Mononychellus* is represented by 31 phytophagous mite species. Including the genus *Mononychellus*, 57 species in 12 genera of Tetranychidae have been recorded on *M. esculenta* and other *Manihot* species so far (Byrne 1983; Migeon and Dorkeld 2023; IPPC 2023). Thirty-one species are known under

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*Mononychellus*, four of which viz., *M. tanajoa*, *M. caribbeanae*, *M. progresivus* Doreste, 1981 and *M. mcgregori* attained pest status in the cultivation of cassava worldwide (Parsa *et al.* 2015; Flechtmann and De Queiroz 2015).

The present study reports *Mononychellus mcgregori* (Flechtmann & Baker) for the first time from India and as the original descriptions are not very well detailed, we here provide a complementary description of the species. Further, the present study uses integrative taxonomy, both morphological and molecular studies to confirm the identity of species.

## Material and Methods

### Morphological identification

Mite infested cassava leaves were collected in polybags and brought to the laboratory for detailed observations under a Carl Zeiss® (Stemi 305) binocular stereo zoom microscope. Adult male and female mites were used for micro slide preparation using Hoyer's medium and cleared on a Hotplate at 70 °C for three days. The specimens were examined and photographed using a Zeiss® AxioScope.A1 phase-contrast and DIC microscope attached with a Nikon® DL7500 camera and the illustrations were edited with Adobe Photoshop CS 2020®. Species identification was done by using taxonomic keys (Flechtmann and de Queiroz 2015) followed by confirming morphological characters using original descriptions (Pritchard and Baker 1955; Flechtmann and Baker 1970) and supplementary descriptions (Gutierrez 1987).

In the redescription, the morphometric data are presented with the mean followed by the ranges in parenthesis, while the leg setal formula is presented with the number of tactile setae followed by the solenidia, eupathidia and duplex setae in parentheses. All the measurements are in micrometers.

All microscopic slide mounts are deposited at the mite repository of AINPAA (All India Network Project on Agriculture Acarology), UAS (University of Agricultural Sciences), GVK (Gandhi Krishi Vignan Kendra), Bengaluru, Karnataka, India.

### DNA analysis

#### Isoline cultures of *Mononychellus mcgregori*

Mites were maintained in the laboratory on Cassava leaves for study. Mites were freshly taken from the culture for DNA isolation.

### DNA extraction and amplification

Total DNA was extracted from three to five individual mites using the modified CTAB method (Doyle and Doyle 1987). The mitochondrial cytochrome c oxidase subunit I gene (COI) barcode region was amplified using primers LCO1490 and HCO2198 (Folmer *et al.* 1994). Polymerase chain reactions (PCR) were conducted using compositions of Taq buffer 1x (Tris with 15 mM MgCl2), 2.5 mM of each dNTP, 1 µM of each primer (Sigma-Aldrich®), 0.5 U Taq DNA polymerase (Genei, Bengaluru, India) in a 25 µL final volume.

Amplification of DNA was done in a Bio-Rad Master cycler (Bio-Rad, USA) with the following PCR conditions: initial denaturation at 94 °C 2 minutes, 94 °C 1 minute, annealing at 47 °C for 1 minute, extension and final extension at 72 °C for one minute followed by 10 minutes then holding at 4 °C. PCR amplicons were visualized on a 2% agarose gel using a Bio-Rad gel documentation unit (AINP, Acarology Lab, GVK, Bengaluru). The sequence was obtained through Sanger sequencing and then edited using BioEdit v7.2.5 and aligned using the default parameters. All the sequences were verified for stop codons and insertions/deletions. BLASTs of nucleotide sequences were carried out on the NCBI database to determine the species identity by match percentage levels.

## Results

### Family Tetranychidae Donnadiieu, 1875

#### Subfamily Tetranychinae Berlese, 1913

#### Tribe Tetranychini Reck, 1950

#### Genus *Mononychellus* Wainstein, 1971

Type species: *Tetranychus planksi* McGregor, 1950

*Eotetranychus planksi* (McGregor, 1950) New combination in Pritchard and Baker, 1955  
Fig. 108–110

*Mononychus mcgregori* Flechtmann & Baker, 1970. Description

*Mononychellus mcgregori* (Flechtmann & Baker 1970). New combination by Flechtmann and Baker (1975) (Fig. 109 in Pritchard and Baker (1955) is *Mononychellus mcgregori* (Flechtmann & Baker)

The name *Mononychellus* was proposed as a new combination for the genus *Mononychus* Wainstein, 1960 (with the type species *Tetranychus planksi* McGregor, 1950). This change was necessary because the genus name *Mononychus* was already assigned to *Mononychus* Schueppel, 1824 (Insecta: Coleoptera) and *Mononychus* Agassiz, 1846 (Insecta: Hemiptera) (Wainstein 1970). Tuttle *et al.* (1976) characterised the genus *Mononychellus* Wainstein, 1960 by the following features: duplex setae on tarsus of leg I distal and adjacent; empodium split near the middle into three proximal hairs and tenent hairs on pad like claw (Bolland *et al.* 1998); dorsal striae of opisthosoma with different patterns; dorsal idiosomal setae usually strongly serrate and sometimes borne on small tubercles; two pairs of anal setae (Wainstein 1960, 1971; Meyer 1974; Tuttle *et al.* 1976).

#### *Mononychellus mcgregori* (Flechtmann & Baker, 1970)

Figs. 1–7

#### Diagnosis

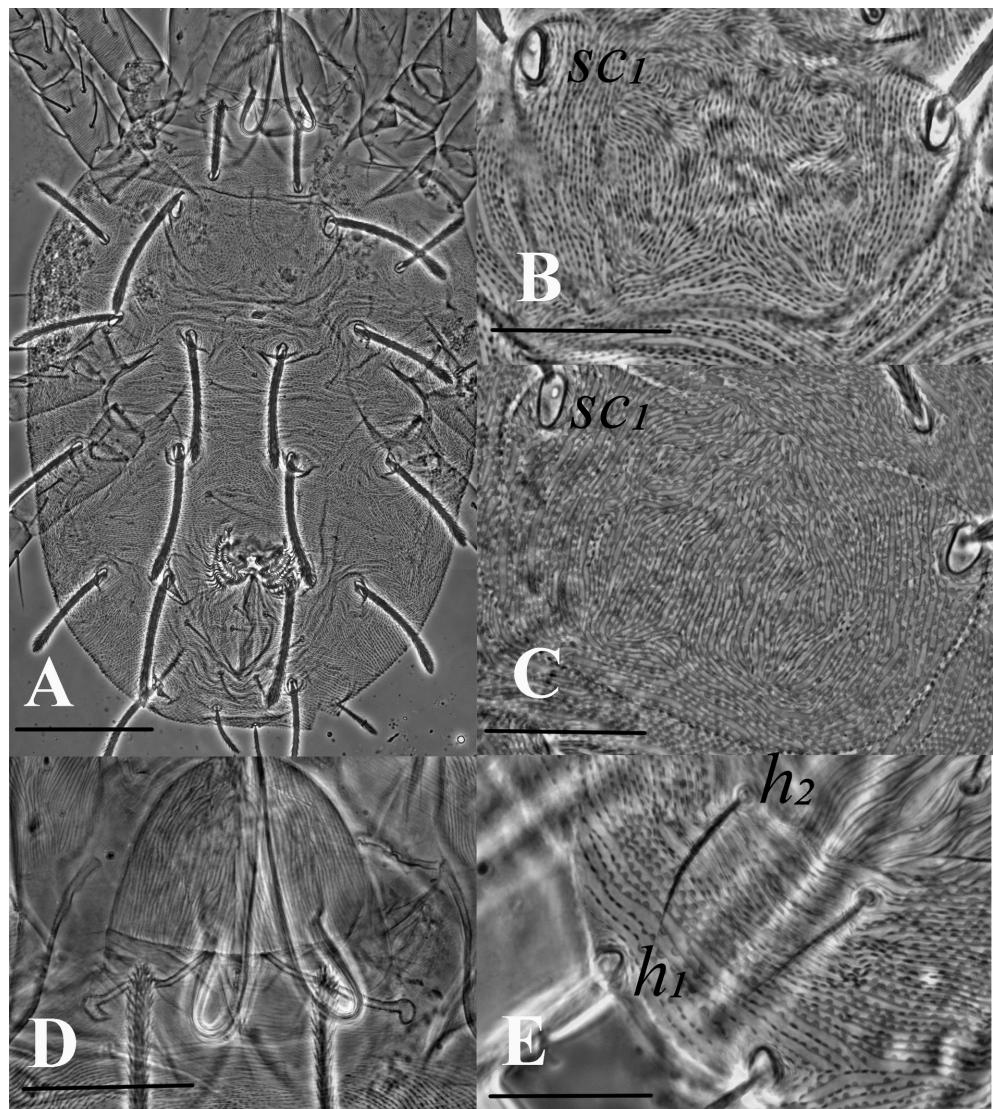
**Female** — Body round to oval, dorsal idiosomal striation more or less transverse, dotted irregular striae around bases of setae. Longitudinal to transverse striae pattern between  $sc_1$  and  $c_1$ , looks like a round anastomous pattern with semi-circular lobes on striations (fig. 1C). Dorsal setae serrated and enlarged distally, set on small tubercles (Tuttle *et al.* 1976; Gutierrez 1987; Flechtmann and De Queiroz 2015; IPPC 2023); bulbous peritreme at gnathosomal region. Terminal sensillum (spinneret) on palptarus wider than long. Opisthosomal setae  $c_1$ ,  $d_1$ ,  $e_1$  and  $f_1$  longer or equal to distance between consecutive setae. Setae  $v_2$ ,  $c_3$ , and  $h_1$  shorter than other setae. Ventral striation at anal region semi-circular lobe like (figs. 1E, 7D) along the striae with regular gaps. Leg tarsus I with four tactile setae and one solenidion proximal to proximal duplex setae. Tibia I with 9 tactile setae and one solenidion.

**Male** — Body smaller than female and narrow towards the caudal end. Terminal sensillum reduced to a minute-thick triangle on palpus. Dorsal striations strongly lobed with long, serrated and tapering setae on tubercles. Legs longer than idiosoma, elongate and end with a bifid claw. Aedeagus elongated, long shaft with a slight bent ventrad and with distal triangular knob. Anterior knob projection extended forward more sharply than the posterior projection.

#### Redescription

##### Female (n=5)

**Dorsum (Figs. 1, 2)** — Length of idiosoma (excluding gnathosoma) 340 (317–360), width (at  $c_3$  level) 252 (228–290). Body more round to oval. Dorsal body striae mostly transverse except in propodosoma (longitudinal to irregular), dorsal striations dotted or lobed (depending



**Figure 1** Adult female *Mononychellus mcgregori* A – dorsal; B – prodorsal striations with dotted lines; C – prodorsal striations with well developed lobes; D – peritreme; E – postanal striations. (Scale bars A, 60µm; B, C, D, 35µm; E, 25µm).

on morphotype and specimens), median prodorsal striations denser, more dotted or lobed, circling the base of the setae (figs. 1B, 1C, 2A). Dorsal setae spatulate and serrate towards the tip, sub equal in length (fig. 1A),  $sc_1$  and dorsocentral setae  $c_1$ ,  $d_1$  and  $e_1$  the longest, setae  $v_2$ ,  $c_3$ , and  $h_1$  the shortest,  $d_2$  and  $e_2$  of almost equal lengths,  $f_1$  twice as long as  $f_2$ . Prodorsal setae  $sc_1$  and hysterosomal setae inserted on quite large tubercles excepted  $c_3$ ,  $f_2$  and  $h_1$ . Setal lengths:  $sc_1$  75 (68–83),  $sc_2$  48 (44–51),  $c_1$  68 (62–78),  $c_2$  65 (57–71),  $c_3$  38 (35–41),  $d_1$  76 (69–84),  $d_2$  68 (57–78),  $e_1$  76 (66–85),  $e_2$  68 (55–80),  $f_1$  62 (55–68),  $f_2$  30 (26–35),  $h_1$  28 (25–32),  $h_2$  20 (15–22) and  $h_3$  20 (15–22). Distances between setal bases:  $v_2$ – $v_2$  44 (41–48),  $sc_1$ – $sc_1$  78 (70–82) and  $sc_2$ – $sc_2$  155 (144–170);  $c_1$ – $c_1$  41 (33–45),  $c_2$ – $c_2$  130 (122–144),  $c_3$ – $c_3$  230 (201–255),  $d_1$ – $d_1$  55 (50–57),  $d_2$ – $d_2$  166 (147–184),  $e_1$ – $e_1$  60 (48–75),  $e_2$ – $e_2$  136 (119–158),  $f_1$ – $f_1$  69 (64–71),  $f_2$ – $f_2$  101 (93–114),  $h_1$ – $h_1$  26 (22–30); Distance between consecutive setae:  $c_1$ – $d_1$  67 (50–82),  $d_1$ – $e_1$  75 (62–91);  $c_3$ ,  $f_2$  and  $h_1$  marginal.

**Venter (Figs. 1E, 2B, 7D)** — Ventral setae smooth and slender. Striations transverse except between 3a and 4a, slightly irregular to longitudinal; genital flap and area immediately

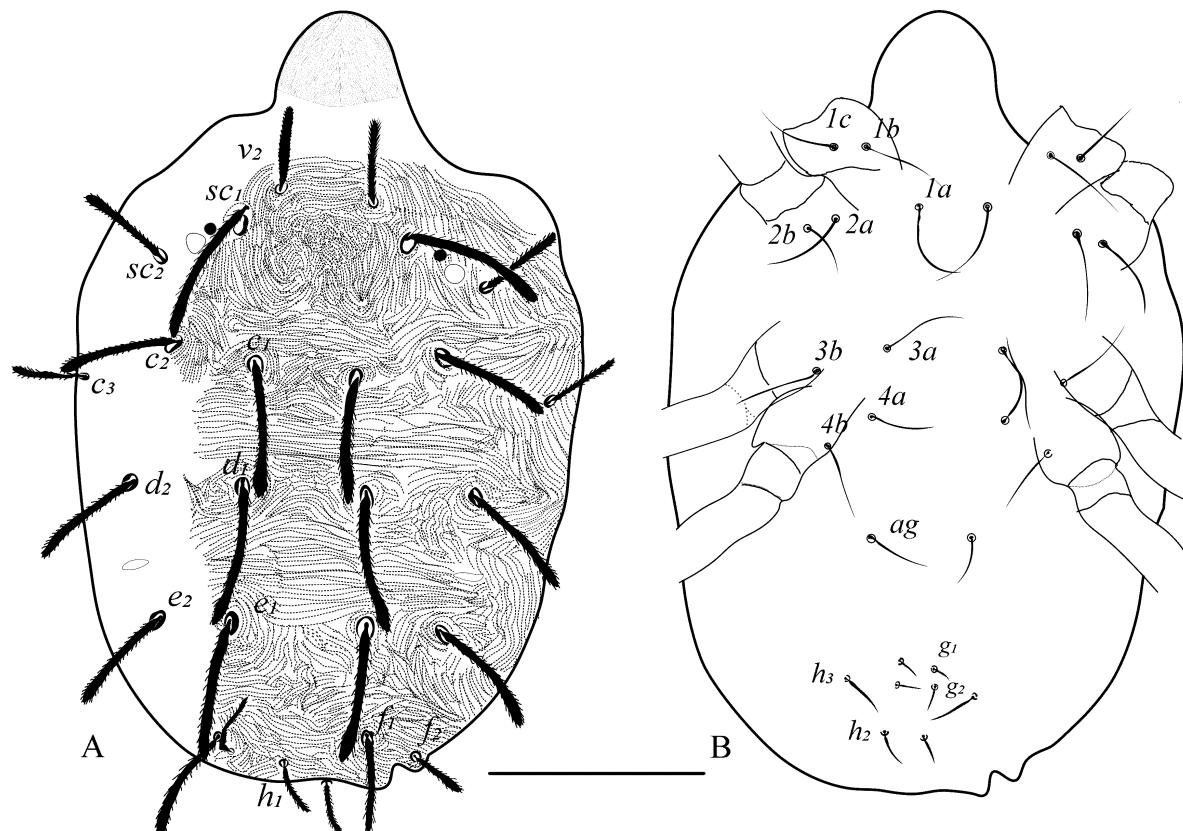


Figure 2 Adult female *Mononychellus mcgregori* A – dorsal; B – venter. (Scale bars A, B 100µm).

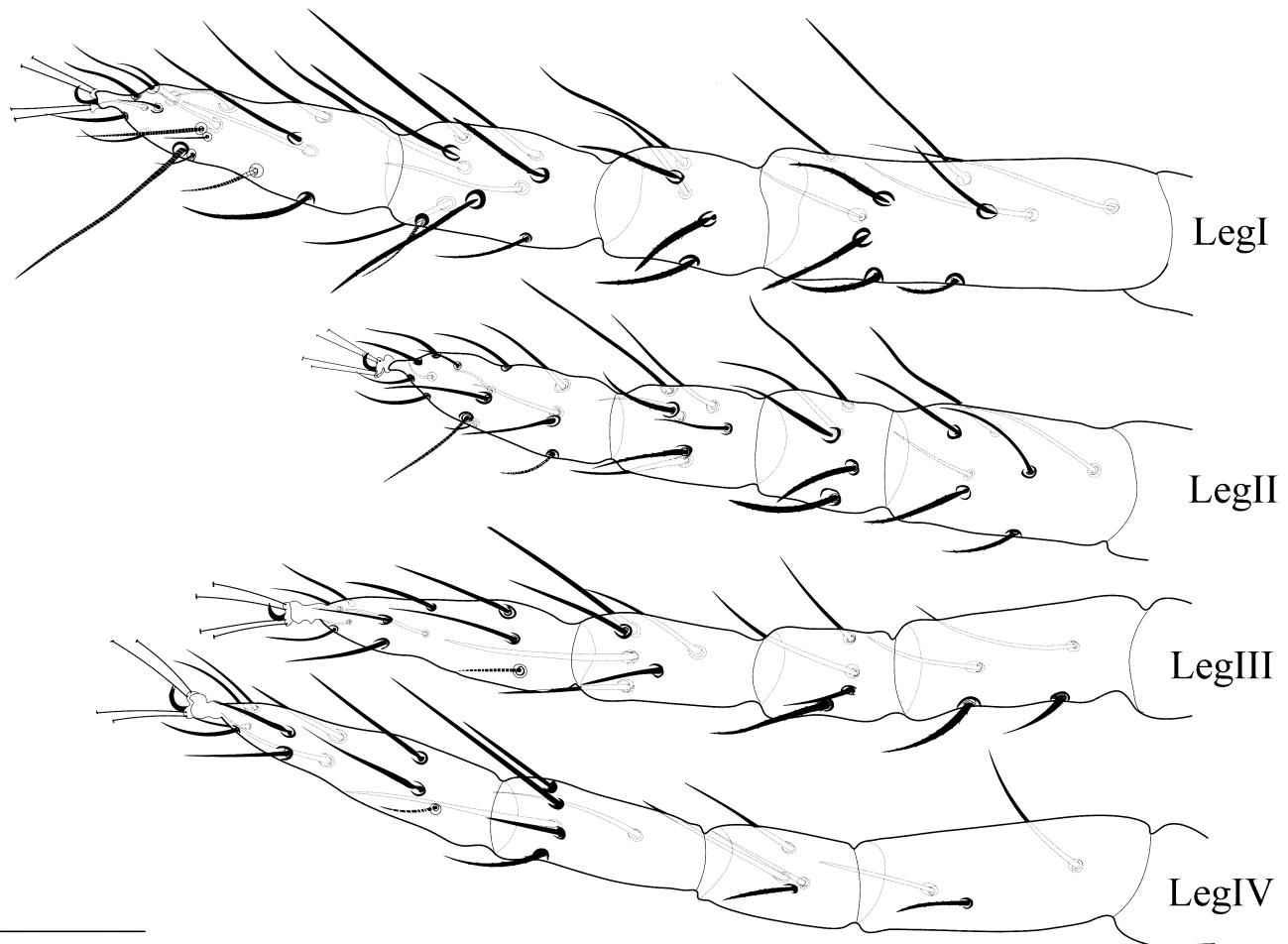
anterior to it with transverse striae, pregenital setae (*ag*) 40 (30–50) long and *ag*–*ag* 48 (45–50). Genital setae (*g*<sub>1</sub>–*g*<sub>2</sub>) and pseudanal setae (*ps*<sub>1–2</sub>) short, tapering. Length of other ventral setae: *m* 28 (27–30), *1a* 45 (40–52), *3a* 45 (40–52), *4a* 45 (40–50). Distances between setae: *m*–*m* 28 (27–30), *1a*–*1a* 29 (27–31), *3a*–*3a* 55 (51–60), *4a*–*4a* 64 (60–72), *ag*–*ag* 48 (45–50). Ventrocaudal setae *h*<sub>2</sub> and *h*<sub>3</sub> tapering, slightly serrated, *h*<sub>2</sub>–*h*<sub>2</sub> 20 (18–22), *h*<sub>3</sub>–*h*<sub>3</sub> 65 (57–80). Pregenital striae transverse with two pairs of genital setae (*g*<sub>1</sub>, *g*<sub>2</sub>) and one pair of *ag* setae. Shape of lobes on ventral striae at *h*<sub>1</sub>–*h*<sub>2</sub>, semi-circular without extending to the dorsum (figs. 1E, 7D).

**Gnathosoma (Figs. 1D, 7A, 7C)** — Stylophore longer than wide. Peritreme enlarged distally like a bulb to a slight hook; out of five specimens, four were asymmetric (one side bulbed and the other side slightly hooked) (figs. 1D, 7C). Palp tibia with thumb claw and two setae. Palptarsus with three tactile setae (*a*, *b*, *c*), a well-developed spinneret, *su* 4 (4–5) width 3 (3–4), two eupathidia, *ul*' 7 (6–7), *ul*'' 5 (5–6) and a solenidion, *ω* 3 (3–4).

**Legs (Figs. 3, 7E)** — Length of legs I–IV (femur to the bent portion of empodium): 244 (230–260), 174 (160–181), 190 (175–212), 210 (201–223), respectively; Leg I with four tactile setae and one solenidion proximal to proximal duplex setae. Tibia with 9 tactile setae and one solenidion. Empodium of leg I–IV split near the middle into three proximal hairs (fig. 7E). Leg setal count as follows: coxae 2–2–1–1; trochanters 1–1–1–1; femurs 10–7–4–3; genua 5–5–4–4; tibiae 9(1sol)–7–6–6; tarsi 11(1sol+2eup+2dup)–10(1sol+2eup+1dup)–8(1sol+2eup)–8(1sol+2eup).

#### Male (n=5)

**Dorsum (Figs. 4, 5)** — Length of idiosoma 230 (220–235) (excluding gnathosoma), width 147 (138–154) at *c*<sub>3</sub> setae. Prodorsum with 3 pairs (*v*<sub>2</sub>, *sc*<sub>1</sub>, *sc*<sub>2</sub>) of setae. Striae between *sc*<sub>1</sub>–*c*<sub>1</sub>



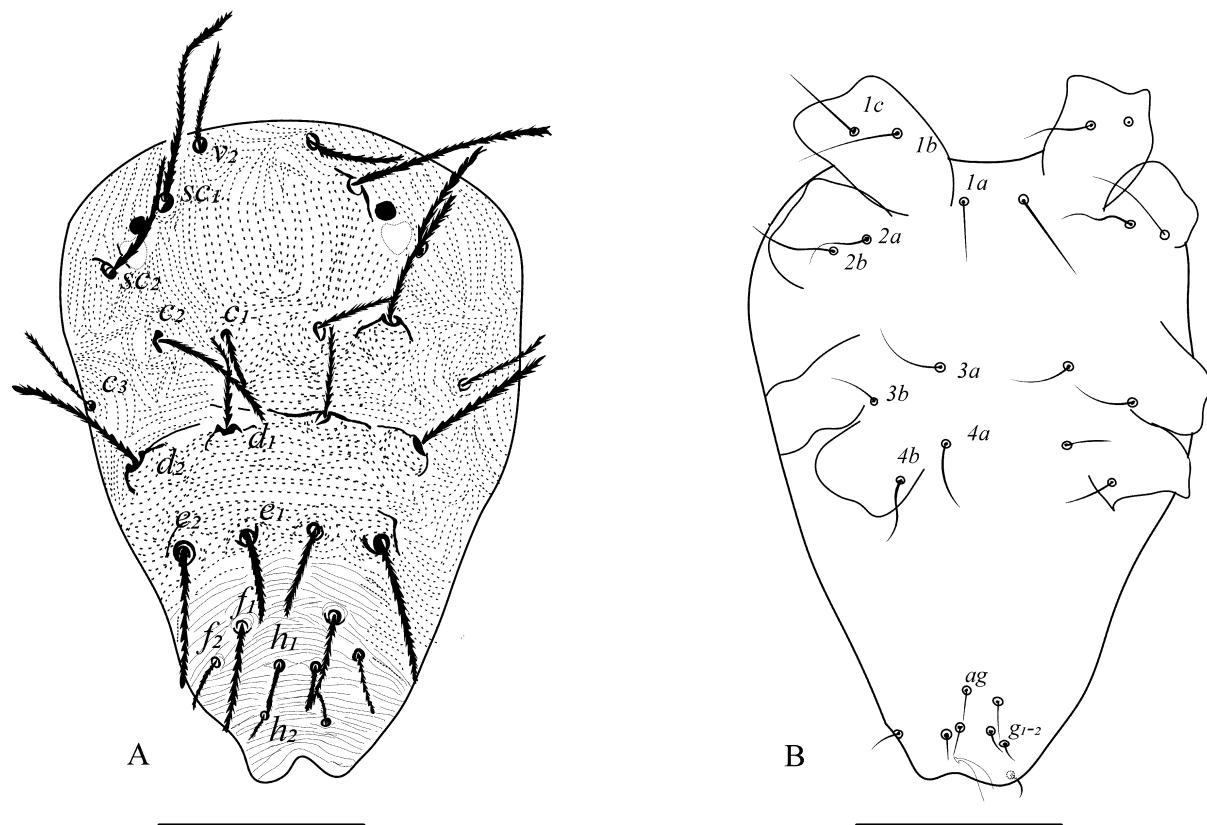
**Figure 3** Adult female *Mononychellus mcgregori* leg I-IV. (Scale bars 40 $\mu$ m).

forming a U-shaped pattern (figs. 4A, 5A). All dorsal setae serrate, tapering,  $sc_1$ ,  $c_2$ ,  $d_2$  and  $e_2$  the longest. Setal lengths:  $v_2$  35 (30-40),  $sc_1$  65 (63-69),  $sc_2$  41 (38-45),  $c_1$  29 (25-37),  $c_2$  52 (45-65),  $c_3$  35 (30-40),  $d_1$  30 (25-33),  $d_2$  57 (46-66),  $e_1$  29 (27-35),  $e_2$  54 (43-65),  $f_1$  33 (30-37),  $f_2$  19 (19),  $h_1$  16 (14-18). Distances between setal bases:  $v_2-v_2$  34 (32-36),  $sc_1-sc_1$  50 (35-60),  $sc_2-sc_2$  98 (95-100),  $c_1-c_1$  26 (25-28),  $c_2-c_2$  75 (67-80),  $c_3-c_3$  125 (115-130),  $d_1-d_1$  31 (27-35),  $d_2-d_2$  92 (86-95),  $e_1-e_1$  19 (18-20),  $e_2-e_2$  64 (60-66),  $f_1-f_1$  29 (27-31),  $f_2-f_2$  43 (42-45),  $h_1-h_1$  10 (9-10),  $h_2-h_2$  17,  $h_3-h_3$  42 (40-45). Setae  $h_1-h_3$  on dorsal side.

**Venter (Fig. 4B)** — Striations sparse and transverse. All ventral setae slender smooth, tapering, shorter than in females;  $m$  18 (17-20),  $1a$  31 (30-32),  $3a$  27 (25-30),  $4a$  25 (23-26),  $ag$  13 (10-16). Distances between setae:  $1a-1a$  18 (17-18),  $3a-3a$  43 (40-45),  $4a-4a$  36 (34-37),  $ag-ag$  8 (8-8). Genital setae ( $g_1-g_2$ ) and pseudanal setae ( $ps_{1-2}$ ) short and tapering.

**Gnathosoma (Fig. 7B)** — All tactile setae on palp slender. Subcapitular setae  $m$  smooth, setaceous. Palptarsus with three tactile setae ( $a$ ,  $b$ ,  $c$ ), with a small spinneret,  $su\zeta$  1 (0.6-1), width 1, two eupathidia  $ul'\zeta$  5 (4-6),  $ul''\zeta$  4 (4-5) and one solenidion ( $\omega$ ) 3 (3-5); solenidion slightly thinner and longer than spinneret.

**Legs (Figs. 6, 7E, 7F)** — Length of legs I-IV (femur to tip of empodium): 271 (245-282), 173 (162-185), 185 (167-192), 214 (195-227) respectively; leg I with six tactile setae and one solenidion proximal to proximal duplex setae. Tibia with 10 tactile setae and two solenidia. Empodium I-II clawlike (Fig. 7F), empodium III-IV as in female (Fig. 7E). Leg setal count as



**Figure 4** Adult male *Mononychellus mcgregori* A – dorsal; B – venter. (Scale bars A, B 55µm).

follows: coxae 2–2–1–1; trochanters 1–1–1–1; femora 10–7–4–3; genua 5–5–4–3; tibiae 11 (2sol)–7–6–6–; tarsi 12(1sol+2eup+2dup)–9 (1sol+2eup+1dup)–8(1sol+2eup)–8(1sol+2eup).

**Aedeagus (Fig. 5B)** — Aedeagus with a long shaft, slightly bent ventrally, featuring a distal triangular knob. Anterior projection of the knob acute, posterior rounded. Ventral margin of the knob with slight depression near the middle (variations can be seen in different focus).

#### Material examined

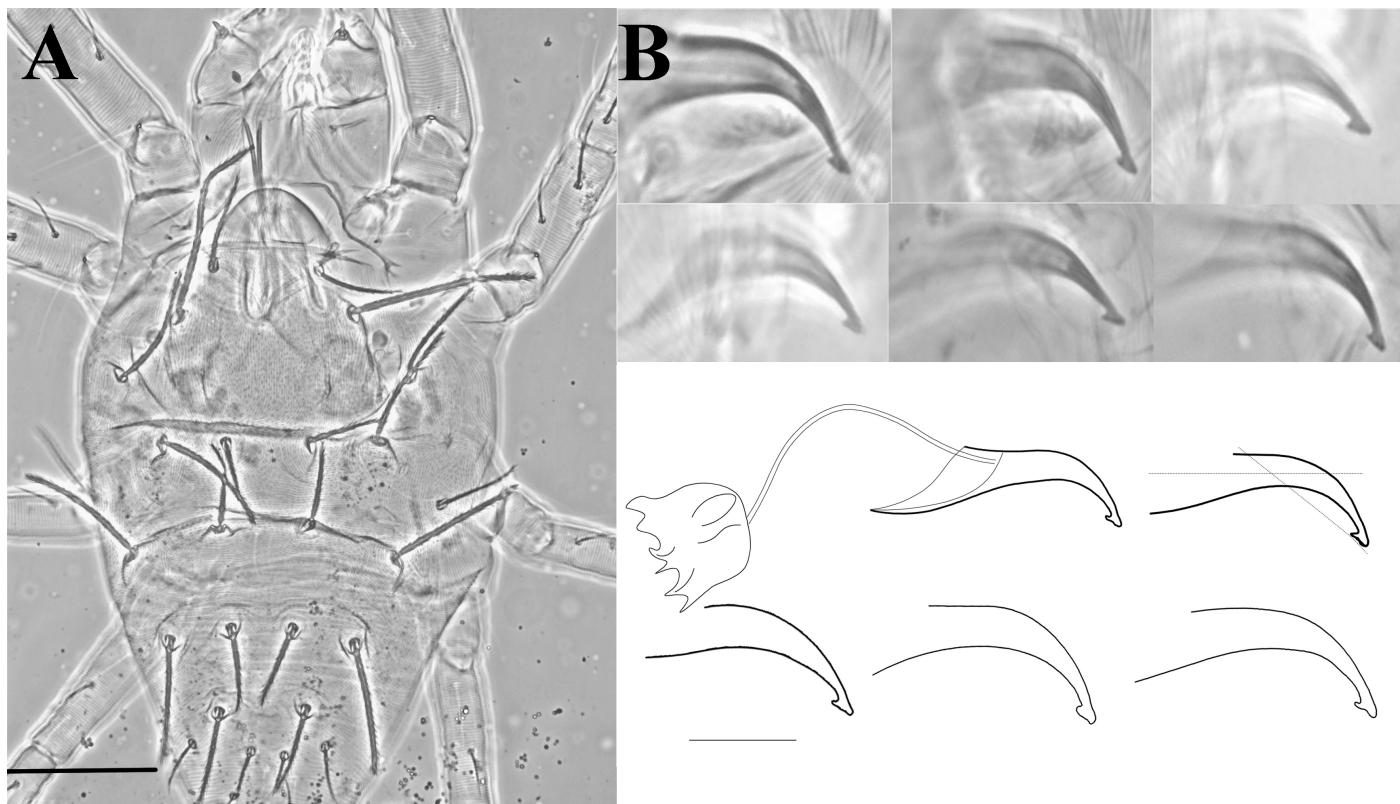
Nine males and 5 females on Cassava (*Manihot esculenta* Crantz) (Euphorbiaceae), Coffee Research Substation (CRSS), Chettalli (12°22'44.9" N 75°50'17.2" E, alt. 1002 m), Chettalli (Kodagu), India, 24 May 2023, coll. P. Dyamanagouda; 10 males, 10 females on Cassava (*M. esculenta*) (Euphorbiaceae), Bengaluru (13°04'36.0" N 77°34'39.5" E, alt. 924 m), GKVK, Bengaluru (Karnataka), India, 04 Sep. 2023, coll. P. Dyamanagouda.

#### Molecular identification

Mite samples collected at Chettalli (Specimen voucher UASB:3596 and NCBI accession number PP074174) and GKVK, Bengaluru (Specimen voucher UASB:3600 and NCBI accession number PP124899) showed similarities of 99.16% and 98.99% with the sequence of *M. mcgregori* (NCBI accession number MN913383) submitted by Ovalle *et al.* (2020) from Colombia, respectively.

#### Remarks

*Mononychellus* species identification was a bit complex earlier due to cryptic morphologies among the species. Pritchard and Baker (1955) introduced a new combination for the type

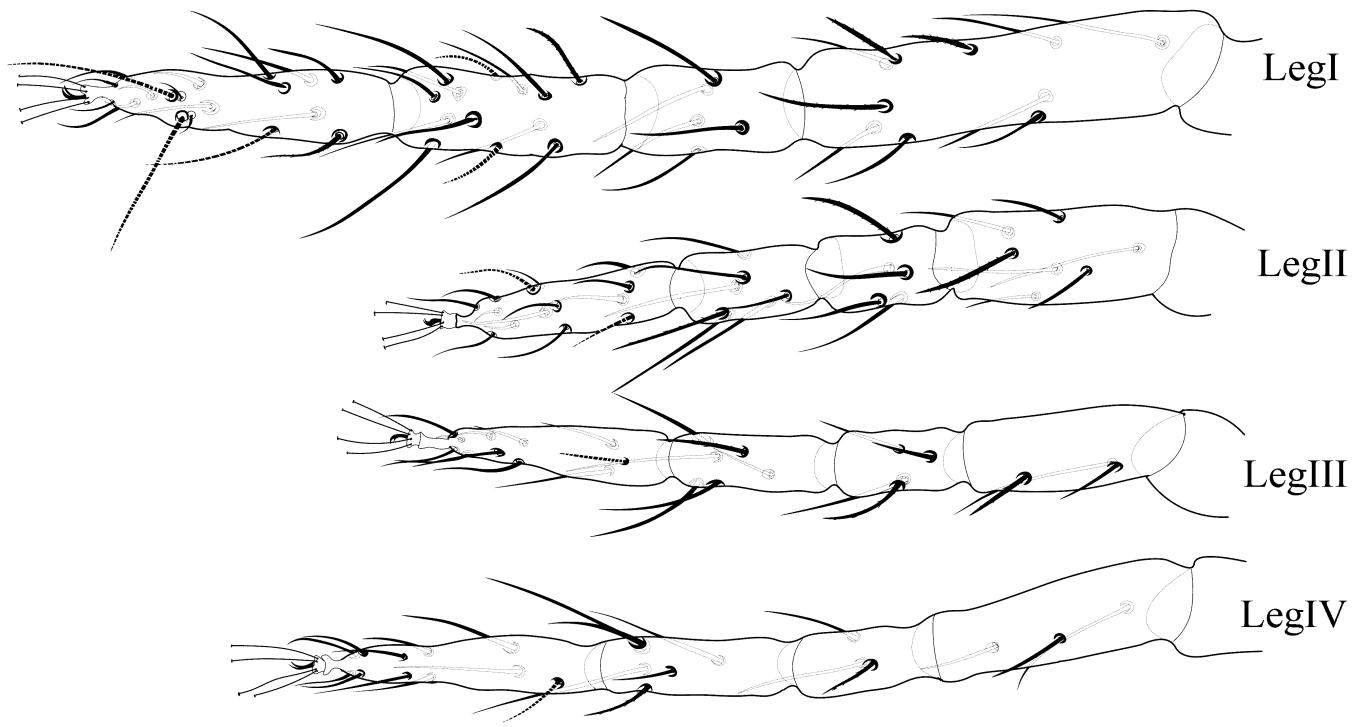


**Figure 5** Adult male *Mononychellus mcgregori* A – dorsal; B – variations in aedeagus of males. (Scale bars A, 80 µm; B, 8 µm).

species *Tetranychus planksi* McGregor, 1950, renaming it as *Eotetranychus planksi*. They noted two morphotypes: one with reticulation at the bases of dorsal setae (Trinidad type) and the other without reticulations at the dorsal setae base (Argentina type). Later, *Eotetranychus planksi* was reclassified as two separate species: *Mononychellus planksi* sensu stricto (McGregor 1950) with reticulation and *M. mcgregori* without reticulation (McGregor 1950; Pritchard and Baker 1955; Flechtmann and Baker 1970; Gutierrez 1987).

Additionally, discrepancies in leg setal numbers, the presence of striations, and other descriptive characters among authors in subsequent publications have been observed (Gutierrez 1987; Flechtmann and De Queiroz 2015). To support this, our investigation observed variability in the lobe pattern present on dorsal striation between mites from two distinct locations. One morphotype (all females from Bengaluru) exhibited dorsal striations resembling dotted or dashed lines, while the other (found in two females from Chettalli) with semi-circular well developed lobes on the dorsal striations (fig. 1B for dotted and 1C for lobed). While, other three female mites from Chettalli are similar to the Bengaluru morphotypes. Despite this variation, all other characteristics remain consistent across both morphotypes in the present study. This variation in dorsal striation of *M. mcgregori* is in congruence with the observations of dotted lines by Pritchard and Baker (1955) (Fig. 109, p. 150) and lobed striations by Flechtmann and Baker (1970) on Argentina morphotypes.

Earlier research has suggested that variations in the dorsal striation pattern and shape could be attributed to several factors such as improperly mounted specimens, environmental conditions, and hybridization effects (Boudreax and Dosse 1963; Monroe 1963; Mollet and Sevacherian 1984). This variation in present study might be due to mounting effect (as only two females observed) or due to genetic diversity due to haploidy nature and gene flow within population (Helle and Pieterse 1965; Dupont 1979). However, even among properly



**Figure 6** Adult male *Mononychellus mcgregori* leg I-IV. (Scale bar 40 $\mu$ m).

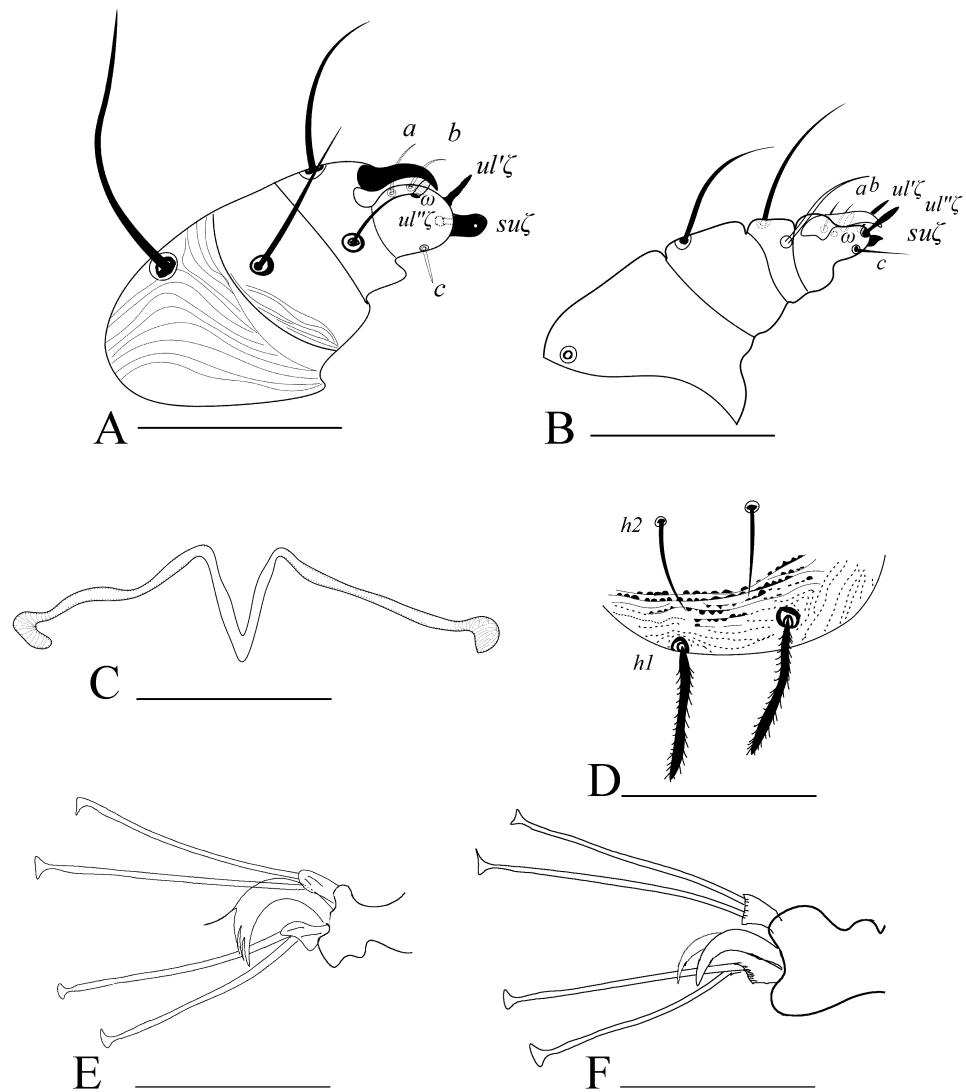
mounted specimens, variations are observed within populations (Boudreux and Dosse 1963; Monroe 1963). Additionally, the dorsal striation shape and pattern of mites is not always a consistent characteristic for taxonomic identification as it varies with each mite form within the population as it is dependent on above factors (Meyer 1974; Jordaan 1977; Auger *et al.* 2013).

Moreover, variations in setal counts in legs of *M. mcgregori* can be seen in earlier literature. Pritchard and Baker (1955) reported one sensory and nine tactile setae without specifying whether these mites were from Argentina or Trinidad. Later, Flechtmann and Baker (1970) provided a full description of *M. mcgregori* based on mites from Argentina, noting that tibia I had one sensory and eight tactile setae.

These kinds of conflicts in the description of *Mononychellus* clearly shows that the genus needs a deep revision. Firstly, descriptions are restricted to females in some sources (Beer and Lang 1958; Estebanes and Baker 1968). Secondly, the cryptic morphology of these mites complicate identification. Lastly, only a few mitochondrial nucleotide sequences (only for *M. tanajoa*, *M. mcgregori*, and *M. caribbeanae*) are present in the GeneBank (NCBI 2024) for comparative analysis.

*Mononychellus mcgregori* is closely related to *M. chemosetosus* (Paschoal) and *M. planksi* (McGregor). *Mononychellus mcgregori* aedeagus is similar to those of *M. planksi* and *M. chemosetosus* (IPPC 2023). However, it can be differentiated from latter species by comparatively longer shaft curved ventrad with anterior acute and posterior round knob. Additionally, reticulations on the prodorsum and the base of each of the opisthosomal setae are absent in *M. mcgregori*.

*Mononychellus mcgregori* is very similar to *M. manihoti* Doreste, 1981 in general morphology and host range but can be differentiated using several characters: 1) the aedeagus of *M. mcgregori* has a long shaft curved ventrad with the anterior projection of the knob acute and



**Figure 7** *Mononychellus mcgregori* A – female pedipalp; B – male pedipalp; C – female peritremes; D – post anal striae and lobes of female; E – female empodium I; F – male empodium I. (Scale bars A, B, C, 30µm; D, 50µm; E, F 15µm).

the posterior rounded, whereas in *M. manihoti*, the aedeagus has a straight shaft without bent and both anterior and posterior projections of the knob are acute; 2) the female tibia I bears 9 tactile setae and one solenidion in *M. mcgregori*, while *M. manihoti* has only 8 tactile setae and one solenidion; 3) the male tibia I bears eleven tactile setae and two solenidia in *M. mcgregori*, whereas *M. manihoti* has nine tactile and three solenidia (Flechtmann and Baker 1970; Doreste 1981).

*Mononychellus mcgregori* shows a high invasive probability to subtropical conditions according to climate models suitability (Lu *et al.* 2012; Rêgo *et al.* 2013; Parsa *et al.* 2015). This situation is particularly important in southern Indian states, notably Kerala, Tamil Nadu, and Karnataka, where cassava is cultivated as a major crop. At the same time, far North-eastern states of India also have a high alert of spread and colonization. Currently, determining the route of entry is challenging and requires further study and intensive surveys. Finally, the ability to feed and establish on multiple hosts is considerable. So, these facts support a significant threat to crop diversity and biodiverse hotspots of India in future.

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