



Near-Complete Genome Sequence of Feline Immunodeficiency Virus from Colombia

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ABSTRACT Feline immunodeficiency virus (FIV) is an important pathogen of domestic and wild felids. Although serological tests suggest the presence of FIV in cats from Colombia, no molecular characterization has been reported. Here, we describe the near-complete genome of FIV subtype A from a Colombian domestic cat.

Feline immunodeficiency virus (FIV) causes AIDS in cats, similar to human immunodeficiency virus (HIV) infection. Both retroviruses belong to the genus *Lentivirus*, and because of their similar genomic organization and pathology, FIV infections have been used as an experimental model for HIV vaccine development and investigation of virus-host interactions (1, 2). FIV strains are classified into six subtypes (A to F), based on *gag* and *env* genes; subtypes A and B are the most prevalent, with worldwide distribution (3). In Colombia, FIV prevalence ranges from 6.7 to 13%, based on serological tests (4, 5), and no data about molecular characterization exist.

Total DNA was extracted from whole blood from a naturally infected male Ragdoll cat from Bogota, Colombia, in 2018, using the DNeasy blood and tissue kit (Qiagen). The near-complete genome was amplified with Platinum SuperFi DNA polymerase (Invitrogen), using primers targeting the primer binding site and the repetition region of the 3' long terminal repeat (6) of FIV proviral DNA (FIVpbs_For, 5'-CAG TTG GCG CCC GAA CAG GGA-C-3' [positions 354 to 375]; FIVR2_Rev, 5'-GTG GGA GCC TCA AGG GAG AAC TC-3' [positions 9381 to 9359, based on the Petaluma reference strain with GenBank accession number [M25381](https://www.ncbi.nlm.nih.gov/nuclot/M25381)]). Purified amplicons were subjected to library construction using the Nextera XT Index and Nextera XT DNA kits (Illumina). The library was sequenced using the MiSeq reagent kit v.3 (600 cycles) on a MiSeq system (Illumina), generating 1,523,068 reads. Reads were trimmed (quality limit, 0.01; minimum read length, 150 nucleotides [nt]) and *de novo* assembled using default parameters (minimum contig length, 5,000 nt) with CLC Genomics Workbench v.12.0.3 (Qiagen). *De novo* assembly of 913,185 reads after trimming (*Q*, >30) generated a contig with 9,056 nt, using 911,799 reads with 24,200-fold average coverage. The annotation of structural and accessory proteins was made based on the FIV Petaluma strain (subtype A). The primer sequences were removed and phylogenetic analyses were conducted with MEGA X (7) and BioEdit Sequence Alignment Editor (8) software.

The near-complete FIV genome sequence of 8,977 nt includes the entire coding regions of structural protein genes *gag* (nt 253 to 1605), *pol* (nt 1494 to 4868), and *env* (nt 5891 to 8461) and accessory protein genes *vif* (nt 4861 to 5616), ORF-A (nt 5617 to 5853), *rev* (nt 5891 to 6136 and 8581 to 8790), and ORF-D (nt 6337 to 6537), as well as the 3' untranslated region (nt 8740 to 8954). Alignment and phylogenetic analysis of

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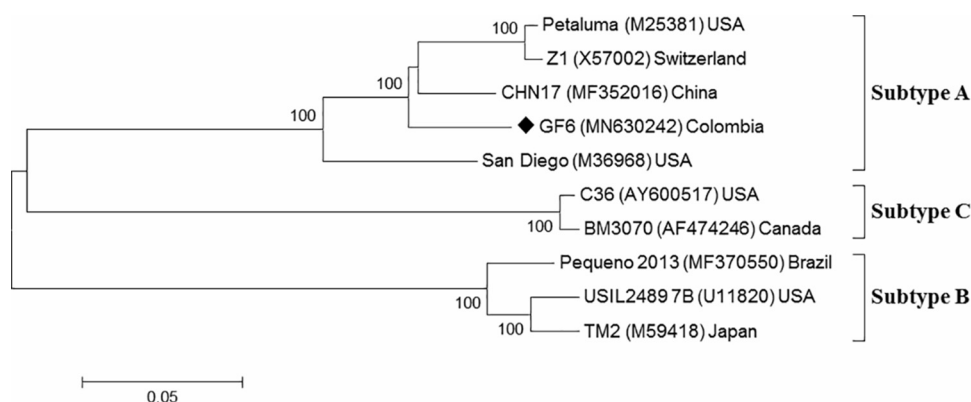


FIG 1 Phylogenetic tree of FIV near-complete genome sequences, constructed with the maximum likelihood method and the general time-reversible model. The diamond marks the sequence obtained in our study. Bootstraps of 1,000 replicates with values of >70 are shown at the nodes. The numbers in parentheses are GenBank accession numbers. The bar represents the substitution number per site.

entire genome sequences demonstrated that FIV isolate GF6 from a Colombian cat belongs to subtype A (Fig. 1). Compared to the Petaluma strain, the full genome of GF6 showed nucleotide identity of 93.7%, whereas amino acid identities of 95.5, 96.0, 89.6, 92.4, 74.3, 90.1, and 83.8% were found for Gag, Pol, Env, Vif, ORF-A, Rev, and ORF-D proteins, respectively.

In South America, studies from Brazil and Argentina demonstrated circulation of subtypes B and E, using partial or complete genome sequences (9–14). Subtype A and an unknown subtype were also described in Brazil (15, 16), but no sequences are available in GenBank. The description of the complete coding sequence plus the 3' untranslated region of FIV subtype A presented here improves the knowledge of the molecular epidemiology of FIV in Colombia and South America.

Data availability. The near-complete genome of FIV isolate GF6 from Colombia is available in GenBank (accession number [MN630242](https://www.ncbi.nlm.nih.gov/nuclseq/MN630242)), and the raw data are available in the SRA database (accession number [PRJNA645463](https://www.ncbi.nlm.nih.gov/sra/PRJNA645463)).

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REFERENCES

- Dunham SP. 2006. Lessons from the cat: development of vaccines against lentiviruses. *Vet Immunol Immunopathol* 112:67–77. <https://doi.org/10.1016/j.vetimm.2006.03.013>.
- Taniwaki SA, Figueiredo AS, Araujo JP. 2013. Virus-host interaction in feline immunodeficiency virus (FIV) infection. *Comp Immunol Microbiol Infect Dis* 36:549–557. <https://doi.org/10.1016/j.cimid.2013.07.001>.
- Duarte A, Tavares L. 2006. Phylogenetic analysis of Portuguese feline immunodeficiency virus sequences reveals high genetic diversity. *Vet Microbiol* 114:25–33. <https://doi.org/10.1016/j.vetmic.2005.11.056>.
- Tique V, Sánchez A, Álvarez L, Ríos R, Mattar S. 2009. Seroprevalence immunodeficiency virus and feline leukemia in cats in Montería, Córdoba. *Rev Med Vet Zoot* 56:85–94.
- Molina VM, Blanco RD, Estepa P, Tamayo S. 2016. Frecuencia del virus de inmunodeficiencia felina (VIF) en el sur del Valle de Aburrá, Colombia (2013–2015). *Rev Cient* 26:374–378.
- Talbott RL, Sparger EE, Lovelace KM, Fitch WM, Pedersen NC, Luciw PA, Elder JH. 1989. Nucleotide sequence and genomic organization of feline immunodeficiency virus. *Proc Natl Acad Sci U S A* 86:5743–5747. <https://doi.org/10.1073/pnas.86.15.5743>.
- Kumar S, Stecher G, Li M, Knyaz C, Tamura K. 2018. MEGA X: Molecular Evolutionary Genetics Analysis across computing platforms. *Mol Biol Evol* 35:1547–1549. <https://doi.org/10.1093/molbev/msy096>.
- Hall TA. 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucl Acids Symp Ser* 41:95–98.
- Caxito FA, Coelho FM, Oliveira ME, Resende M. 2006. Feline immunodeficiency virus subtype B in domestic cats in Minas Gerais, Brazil. *Vet Res Commun* 30:953–956. <https://doi.org/10.1007/s11259-006-3363-8>.
- Lara VM, Taniwaki SA, Araújo JP. 2007. Caracterização filogenética de amostras do vírus da imunodeficiência felina (FIV) do Estado de São Paulo. *Pesq Vet Bras* 27:467–470. <https://doi.org/10.1590/S0100-736X2007001100004>.
- Martins AN, Medeiros SO, Simonetti JP, Schatzmayr HG, Tanuri A, Brindeiro RM. 2008. Phylogenetic and genetic analysis of feline immunodeficiency virus *gag*, *pol*, and *env* genes from domestic cats undergoing nucleoside reverse transcriptase inhibitor treatment or treatment-naïve cats in Rio de Janeiro, Brazil. *J Virol* 82:7863–7874. <https://doi.org/10.1128/JVI.00310-08>.

12. Taniwaki SA, Araújo JP, Brandão PE. 2017. First nearly complete genome sequence of feline immunodeficiency virus from Brazil. *Genome Announc* 5:e00947-17. <https://doi.org/10.1128/genomeA.00947-17>.
13. Teixeira BM, Taniwaki SA, Menezes PMM, Rodrigues AKPP, Mouta AN, Arcebispo TLM, Braz GF, Cruz JCM, Brandão PE, Heinemann MB, Silva MX, Hosie MJ. 2019. Feline immunodeficiency virus in Northern Ceará, Brazil. *JFMS Open Rep* 5:2055116919859112. <https://doi.org/10.1177/2055116919859112>.
14. Cano-Ortiz L, Junqueira DM, Comerlato J, Costa CS, Zani A, Duda NB, Tochetto C, dos Santos RN, da Costa FVA, Roehe PM, Franco AC. 2017. Phylodynamics of the Brazilian feline immunodeficiency virus. *Infect Genet Evol* 55:166–171. <https://doi.org/10.1016/j.meegid.2017.09.011>.
15. Marçola TG, Gomes CPC, Silva PA, Fernandes GR, Paludo GR, Pereira RW. 2013. Identification of a novel subtype of feline immunodeficiency virus in a population of naturally infected felines in the Brazilian Federal District. *Virus Genes* 46:546–550. <https://doi.org/10.1007/s11262-013-0877-3>.
16. Martins NS, Rodrigues APS, da Luz LA, dos Reis LL, de Oliveira RM, de Oliveira RA, Abreu-Silva AL, dos Reis JKP, Melo FA. 2018. Feline immunodeficiency virus subtypes B and A in cats from São Luis, Maranhão, Brazil. *Arch Virol* 163:549–554. <https://doi.org/10.1007/s00705-017-3636-2>.