

Review

# Drugging Hijacked Kinase Pathways in Pediatric Oncology: Opportunities and Current Scenario

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**Abstract:** Childhood cancer is considered rare, corresponding to ~3% of all malignant neoplasms in the human population. The World Health Organization (WHO) reports a universal occurrence of more than 15 cases per 100,000 inhabitants around the globe, and despite improvements in diagnosis, treatment and supportive care, one child dies of cancer every 3 min. Consequently, more efficient, selective and affordable therapeutics are still needed in order to improve outcomes and avoid long-term sequelae. Alterations in kinases' functionality is a trademark of cancer and the concept of exploiting them as drug targets has burgeoned in academia and in the pharmaceutical industry of the 21st century. Consequently, an increasing plethora of inhibitors has emerged. In the present study, the expression patterns of a selected group of kinases (including tyrosine receptors, members of the PI3K/AKT/mTOR and MAPK pathways, coordinators of cell cycle progression, and chromosome segregation) and their correlation with clinical outcomes in pediatric solid tumors were accessed through the R2: Genomics Analysis and Visualization Platform and by a thorough search of published literature. To further illustrate the importance of kinase dysregulation in the pathophysiology of pediatric cancer, we analyzed the vulnerability of different cancer cell lines against their inhibition through the Cancer Dependency Map portal, and performed a search for kinase-targeted compounds with approval and clinical applicability through the CanSAR knowledgebase. Finally, we provide a detailed literature review of a considerable set of small molecules that mitigate kinase activity under experimental testing and clinical trials for the treatment of pediatric tumors, while discuss critical challenges that must be overcome before translation into clinical options, including the absence of compounds designed specifically for childhood tumors which often show differential mutational burdens, intrinsic and acquired resistance, lack of selectivity and adverse effects on a growing organism.

**Keywords:** childhood cancer; kinases; chemical inhibitors; clinical trials



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## 1. Pediatric Cancer

The past two decades have witnessed tremendous advances in our understanding of cancer pathogenesis, with most neoplasms resulting from the accumulation of gains in function in proto-oncogenes and losses of tumor suppressors.

In the pediatric setting (between 0 and 19 years of age), cancer is defined as a group of several diseases that have in common the uncontrolled proliferation of abnormal cells that can occur in any region of the body. However, unlike adult tumors that are classified

according to the primary site, the International Classification of Childhood Cancer (ICCI) categorizes pediatric tumors into 12 main groups based on histological findings [1].

Of these, leukemias are the most frequent, representing 35% of all tumors that affect children and adolescents [2]. The second most common group is represented by lymphomas (20%), followed in descending order by tumors of the central (15%) and sympathetic (7%) nervous system, soft tissue sarcomas (6%), bone tumors (5%), kidney tumors (3%), germ cell tumors (3%), retinoblastoma (2%), carcinomas (2%), liver tumors (1%) and other rare pediatric neoplasms.

Several specific characteristics converge on the premise that childhood and adult cancer should be studied separately. First of all, most pediatric tumors have histological findings that resemble fetal tissues at different stages of development, being considered embryonic and carrying different levels of cell differentiation. Furthermore, the spectrum of tumors in the pediatric age group differs from that in adult patients. Medulloblastoma (MB), neuroblastoma (NB), rhabdomyosarcoma (RMS), Ewing's sarcoma (EWS), osteosarcoma (OS), retinoblastoma (RB) and Wilms' tumor (WT), which are the most frequent pediatric solid tumors, are rarely found in adulthood [3].

Moreover, latency periods are shorter in pediatric cancer, with several histologies presenting even shortly after birth. Such rapid proliferation in embryonic tissues relies mainly on genomic errors with lesser contribution of environmental factors [4]. Similarly, unlike what happens in adults, pediatric tumors are usually more aggressive with nonspecific signs and symptoms, confusing them with common childhood illnesses and making early diagnosis difficult.

Additionally, tumors in this age group show clear differences in their presentation, clinical course and response to treatment when compared to adult counterparts. Tumors of the EWS family, for example, in adults, in addition to presenting a more differentiated histology (PNET—primitive neuroectodermal tumor), manifest preferentially with greater volume at diagnosis, affecting soft tissues and with distant metastases, resulting in poorer survival. In children, EWS preferentially affects the bones of the extremities, have a smaller volume, and respond better to chemotherapy. Still, in this population, the chance of survival after 5 years (without metastasis at diagnosis) is 75%, significantly higher than that calculated for adults (50%) [5]. Similar observations have been reported for RMS. In a cohort of 1071 adults and 1529 children, for example, the survival rate in the first group was considerably lower (27% versus 61%), with tumors occurring in unfavorable locations and with rare histologies [6].

Another peculiarity of childhood cancer lies in the fact that specific histological subtypes and clinical behavior are also age-dependent, suggesting differential pathogenic mechanisms and underlying molecular alterations for tumor initiation and progression. The general incidence of acute lymphoblastic leukemia (ALL), for example, is highest in the 1–4-year-old age group, while the highest frequency of lymphomas occurs among adolescents (between 15 and 19 years old). Embryonic tumors (NB, WT, RB, etc.), on the other hand, share a descending incidence, which is highest early in life and almost dissipates after 5 years of age, while the incidence of bone sarcomas reaches a sharp peak at the time of the pubertal growth spurt [7].

Finally, with the methodologic refinements in the identification of genomic alterations, it has become increasingly evident that the spectra of mutations and the subsequent dysregulation of signaling pathways in pediatric neoplasms differ from those that occur predominantly in adult cancer. In fact, it has been stipulated that most pediatric tumors carry between 5 and 10 mutations; however, the average number of mutations in adult tumors varies between 33 and 66 (i.e., colon, breast or pancreas carcinomas) and increases up to 200 in tumors caused by mutagens (such as melanoma/ultraviolet radiation (UV) and lung cancer/smoking) and up to 1000 in tumors with defects in mismatch repair genes, as is the case of nonpolyposis colorectal cancer, among others [8]. In this regard, a recent integrative study based on whole-genome sequencing data from 24 tumor types (914 patients) showed that even though mutation frequencies (SNV and indels) vary

between pediatric tumor types (from 0.02 to 0.49 per Mb), these were 14 times lower than in adult cancers [9]. Of note, a high prevalence of mutations affecting genes related to cancer predisposition syndrome are seen in children affected by different tumor types [10]. In addition, pediatric cancers are usually enriched by gene fusions, driving tumorigenesis and showing impact on both diagnostic and targeted-treatments [11].

In fact, this work identified 52 genes significantly mutated for childhood and juvenile tumors and 102 genes for adult tumors, 25 of which were shared by both groups. *TP53* was the most commonly mutated gene (4% of childhood tumors), followed by *KRAS*, *ATRX*, *NF1* and *RB1* (1–2% of tumors). In adult tumors, *TP53* was also the gene most affected by mutations, albeit tenfold more frequently. Furthermore, the burden of mutations increased with patient age except for tumors characterized by *kataegis* or *chromothripsis* events [9].

Moreover, the mutational identity may also vary. Glioblastoma (GBM) (grade IV astrocytoma), for example, is characterized by mutations in *PTEN* and epidermal growth factor receptor (*EGFR*) amplification in adults [12]; the pediatric counterpart more frequently presents mutations in the N-terminal tail of the histone variant 3.3 [13], in platelet-derived growth factor (*PDGF*) and its receptor (*PDGFR*) [14]. As previously described, gene fusions that are rare in adult tumors appear recurrently in pediatric tumors such as *BRAF/KIAA1549* in pilocytic astrocytoma (PA) [15], *C11orf95/RELA* [16] in supratentorial ependymoma (EPN), *PAX3/FOXO1* and *PAX7/FOXO1* in alveolar RMS [17], and variants involving the *EWS* gene (*EWS/FLI1*, *EWS/ERG* and others less frequent) in EWS [18]; the majority of these fusions involve transcription factors associated with the development/differentiation of the affected tissue.

More recently, with the establishment of cooperative study groups and progress in imaging associated with more accurate anatomopathological and molecular diagnosis, the mortality of children affected by cancer, especially those with leukemia and some types of solid tumors, has shown an important decline. Five-year overall survival rates increased from 56% in the 1970s to 77% in the following vicennial [19].

In accordance, today ALL represents the paradigm of curable cancer in children, with current overall survival rates exceeding 85% in most modern treatment protocols [20]. However, for some aggressive leukemia subtypes and certain solid tumor histologies, the persisting advances in biological characterization combined with new technologies in radiotherapy (RT), chemotherapy (CT) and supportive/rehabilitation care have resulted in marginal survival advantages, and cure rates have stagnated at around 70% [21]. Yet, in underdeveloped areas such as Eastern Europe, Africa and South America, these reductions in mortality have been less expressive, and a considerable portion of children with cancer fail to respond to traditional chemotherapy.

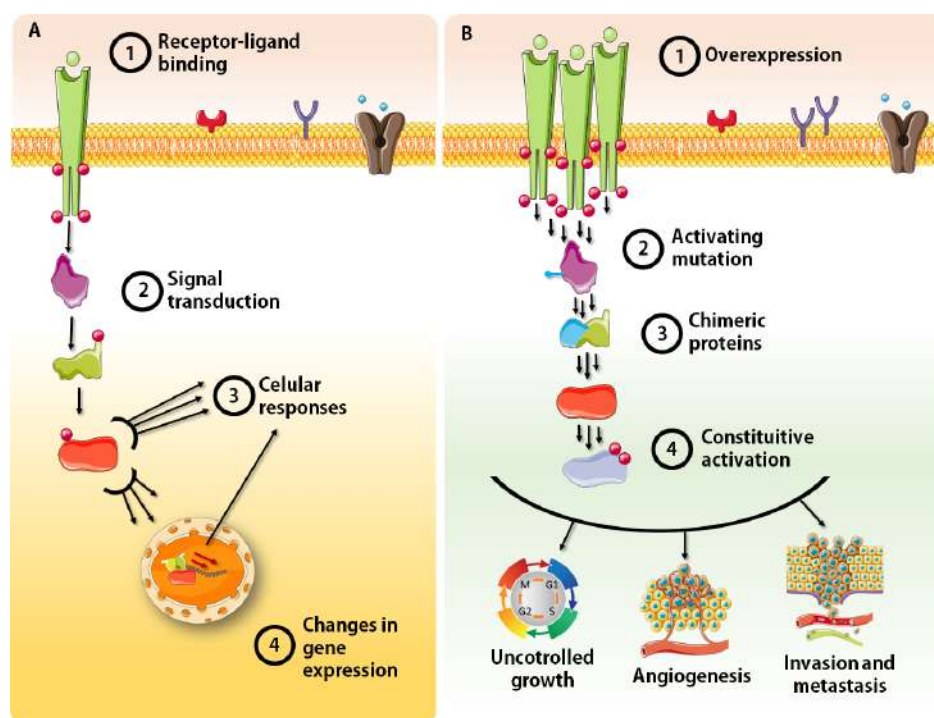
In this scenario, alternative rationally targeted pharmacological options are still needed to overcome clinical resistance, tumor progression, and prevent the adverse side effects of standard therapy.

## 2. Kinases as Cancer Drivers

The human genome encodes more than 500 protein kinases, enzymes responsible for turning protein functions “on” through the transference of  $\gamma$ -phosphate groups from ATP to one of their three amino acids with free hydroxyl groups: serine, threonine or tyrosine. Human protein kinases have been divided into nine classes which are further subdivided into families, and often subfamilies whose actions can alter up to 30% of all cell proteins [22]. The molecular shifts exerted by them through phosphorylation not only can affect the function of a given protein, but it can also stabilize it, localize it in a particular cellular compartment and modulate its association with other proteins [23].

Protein kinases may be triggered or deactivated in many ways, including cis- or autophosphorylation, binding with substrates or activator/inhibitor proteins. Once activated, they act as crucial regulators of many features of cell behavior and specialized functions by coupling reception of extracellular signals, intracellular signaling transduction and cellular responses [24].

Playing fundamental roles in cell division, survival and migration, their dysregulation is commonly associated with human malignancies and contributes to tumor initiation and all stages of cancer progression. In fact, innumerable mutations, translocations, and amplifications that result in constitutively overexpressed or active kinases have been demonstrated in many human cancers [23]. Mutations within the catalytic domain serve to stabilize the kinase in an active conformation and to destabilize cis-inhibitory interactions. Other domains can also be affected and elicit constitutive activity and hyperactive pathways as well. DNA translocations, on the other hand, predominantly create in frame gene fusions leading to chimeric proteins with novel/increased activity, leading to continued cancer cell growth and survival (Figure 1) [25,26]. Moreover, the dysregulation of different kinases has been repeatedly associated with tumor prognosis and categorized as a determinant of patient survival [27,28].



**Figure 1.** (A) Protein kinases may be triggered or deactivated in many ways, acting as key regulators of many features of cell behavior and specialized functions by coupling (1) reception of extracellular signals and (2) intracellular signaling transduction, leading to (3) direct cellular responses or (4) changes in gene expression. (B) In cancer, increased kinase activity may result from gene amplification (1,4), mutations that stabilize the kinase in an active conformation and destabilize cis-inhibitory interactions (2) and translocations that encode chimeric proteins with novel/increased activity (3). This figure was created using Servier Medical Art templates, which are licensed under a Creative Commons Attribution 3.0 Unported License; <https://smart.servier.com> (accessed on 14 December 2022).

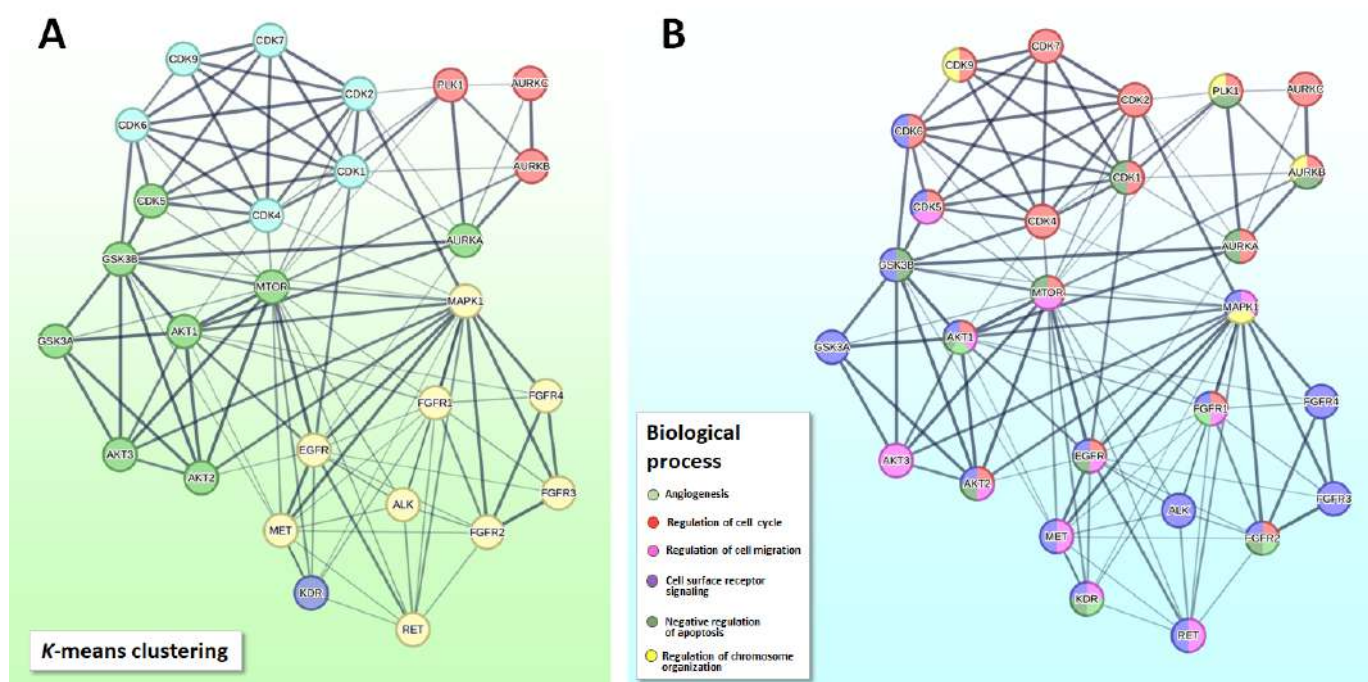
Hence, the so-called “Kinome” (the complete set of protein kinases encoded in the human genome—about 2% of all genes) has become an attractive target for the treatment of a variety of tumors. Even so, despite the great diversity, over the years, it has become more apparent that only certain kinases are among the most frequently occurring drivers of human cancer, including tyrosine receptors (RTK) (i.e., FGFR, EGFR, VEGFR, RET, MET, ALK), members of the PI3K/AKT/mTOR and Mitogen-Activated Protein Kinase (MAPK) pathways, along with central coordinators of cell cycle progression (i.e., cyclin-dependent kinases) and chromosome segregation such as polo-like and aurora kinases [25].



In this way, the present study aimed to present evidence of the involvement of these kinases' dysregulation in the pathophysiology of pediatric tumors, their correlation with clinical outcomes and prospects of their inhibition through in silico analysis, along with an up-to-date revision of compound development and testing.

### 3. Protein Kinases in Pediatric Oncology and Their Association with Tumor Prognosis

As stated above, substrate reversible phosphorylation by protein kinases is nature's main molecular system for organizing cellular signal transduction and regulating cell metabolism, growth and differentiation. The phosphorylation state of a protein determines not only its function, subcellular distribution and stability, but also its interaction with other proteins or cellular components. Intrinsically, signaling pathways are remarkably complex and as our knowledge increases, it has become progressively evident that such molecular networks are not linear but contain modules of multi-protein complexes, many feedbacks, feedforwards and competing protein mechanisms that not only assemble at various intracellular compartments to process, integrate and transmit information that will ultimately specify a particular biological response, but also crosstalk with many other signaling pathways [29]. Thus, even though the kinases that were selected for this review will be treated separately, many, if not all, are directly or indirectly interconnected (Figure 2).



**Figure 2.** (A) Protein–protein interactions accessed through the software STRING v11.5 (available at <https://string-db.org/> (accessed on 2 November 2022)). The parameters evaluated were text mining, experiments and databases. Network edges denote confidence and the minimum required interaction score was 0.700, considered high. (A) K-means clustering; (B) enrichment analysis for biological processes.

In this section, the different roles of protein kinases in oncogenic transformation and tumor prognosis in the pediatric setting were assessed by two different approaches: by a thorough search of published literature, and by a systematic search in publicly available data retrieved from expression arrays accessed through the R2: Genomics Analysis and Visualization Platform (<http://r2.amc.nl> (accessed on 15 October 2022)). For this, datasets were included if they met the following criteria: inclusion of pediatric samples (exclusively or in which adult variants could be omitted), normal counterparts and having information

about clinical features of prognosis (Supplementary Figure S1). Datasets and probes are detailed in Supplementary Tables S1 and S2.

### 3.1. Published Evidence of Kinase Dysregulation in Pediatric Oncology

#### 3.1.1. Receptor Tyrosine Kinases (RTK)

Humans express 58 receptor tyrosine kinases (RTK) which function as entry points for many extracellular signals and the recruitment of the intracellular signaling networks that orchestrate a particular response [30].

These cell surface receptors possess multi-domain identical architectures that are made up of an extracellular ligand-binding domain (which differs between subfamilies), a single transmembrane helix and an intracellular region that contains a juxtamembrane regulatory region (composed of 40–80 amino acids), a tyrosine kinase domain (TKD) and a carboxyl (C-) terminal tail [31].

Generally, RTK activation occurs upon binding of ligands (i.e., growth factors or cytokines) to their extracellular domains. This interaction results in RTK non-covalent dimerization/oligomerization, which juxtaposes the cytoplasmic TKDs and facilitates autophosphorylation in trans of tyrosine residues in the juxtamembrane regulatory region, inducing conformational changes that serve to stabilize the active state of the kinase. Then, a second phase of tyrosine autophosphorylation occurs on phosphotyrosines that recruit downstream signaling proteins that typically contain Src homology 2 (SH2) or phosphotyrosine-binding (PTB) domains. The recruitment of these adapter molecules then initiates a cascade of RTK-specific pathways that determine cell fate [22,31,32].

About 20 different RTKs classes or subfamilies have been described [33]. Their activity is tightly regulated in normal cells; however, constitutive kinase activity acquired through mutation, overexpression and/or autocrine/paracrine stimulation has been strongly associated with pathological disorders, neoplastic transformation and metastasis [34]. Dysregulation of the epidermal growth factor receptor (EGFR/ErbB), the receptor for insulin (IR), the platelet-derived growth factor receptor (PDGFR), the fibroblast growth factor receptor (FGFR), the vascular endothelial growth factor receptor (VEGFR) and the hepatocyte growth factor receptor (HGFR/MET), for example, results in uncontrolled activation of multiple downstream signal transduction pathways and provides a strong drive toward malignancy [35]. Some of these oncogenes are paradigms of certain tumor types, as is the case of the amplification of EGFR/ErbB in breast cancer or MET overexpression in non-small cell lung cancer (NSCLC) [36,37]. Nevertheless, information about RTKs' involvement in the pathophysiology of childhood cancer is less discernible, as illustrated below.

**FGFR.** The signaling cascades of fibroblast growth factor receptors (FGFR1, FGFR2, FGFR3 and FGFR4) play pivotal roles in the regulation of development and tissue repair and regeneration. These receptors are highly conserved and widely distributed and their dysregulation promotes tumor growth, survival and development of drug resistance, as well as the development of angiogenesis and immune evasion [38]. A recent pan-cancer next-generation sequencing profiling demonstrated that ~7% of cancers harbor gain-of-function FGFR aberrations, with varying frequencies between family members (FGFR1 > FGFR3 > FGFR2 > FGFR4) [39]. Gene amplifications or activating mutations have been observed in multiple cancer types, although they are most commonly detected in breast, lung, liver, stomach, uterus and bladder cancer [40]. In fact, most of the aberrations detected in the survey performed by Helsten et al. (2016) involved adult carcinomas, while the percentage of cases positive for *FGFR* aberrations in childhood tumors (NB and OS) was around 3% [39]. Additionally, *FGFR1* fusions (i.e., FGFR1-TACC1), TKD duplications and hotspot mutations (i.e., N5465K and K656E) are frequently observed in certain types of pediatric brain cancer, particularly dysembryoplastic neuroepithelial tumors (DNET) and PA [41]. Moreover, germline mutations in *FGFR1*, either complete or in mosaicism, may predispose low-grade central nervous system (CNS) tumors in children and adolescents [42,43].

Other research groups have also found correlations between altered expression of FGFRs and poor prognosis. *FGFR1* amplification, for example, was correlated with worse prognosis and poor response to chemotherapy in a large cohort of patients with OS [44]. Moreover, *FGFR1* has been correlated with tumor development and lung metastasis in xenographic models, whereas its activation improved survival and radiation resistance, a phenotype which was reversed when *FGFR1* was inhibited [45]. Similar to OS, *FGFR1* copy number gains are frequent in EWS, where patients with activating *FGFR1* mutations present higher incidence of metastatic disease [46]. In vitro, *FGFR1* suppression through interference RNA (RNAi) significantly reduced cell proliferation. Decreased xenograft tumor growth and 18F-fluorodeoxyglucose activity were also observed [47].

*FGFR1* amplification and gene fusions have also been described in RMS with an active role cancer cell proliferation [48,49]. However, opposite expression patterns have been reported [50,51]. Nevertheless, it has already been shown that embryonal (ERMS) histologies present higher *FGFR1* expression levels compared to the alveolar forms (ARMS) [51].

Considering CNS tumors, upregulation of *FGFR1* has been associated with worse prognosis and shorter overall and recurrence-free survival in EPN and NB [52–55]. The analysis of *FGFR1* protein abundance in human MB tissues, through an anonymized, validated MB, and cerebellum tissue microarray (TMA) and immunohistochemistry (IHC) found high levels of *FGFR1* expression in 18% of the tumor tissues. In the case of gliomas, many mutations involving the *FGFR1* gene were described, one of which was associated with radioresistance [56–60]. Otherwise, in WT and MB, there is no published evidence.

Regarding *FGFR2*, information in the literature about its relevance in pediatric tumors is scarce. Besides correlations with higher tumor grade, radioresistance and poorer survival in gliomas [61–63], its phosphorylation (indicative of activation) was seen increased in NB samples (compared to normal tissues) and correlated with cisplatin resistance [62]. Alternatively, low expression of this kinase was observed MB [63] and in RMS when compared to normal myeloblasts [64]. Downregulation or undetectable expression of *FGFR3* was also reported in RMS with no evidence of correlation with the clinical outcome [64]. However, a more recent study described a small population of *FGFR3*-positive cells as strongly tumorigenic with a stem cell-like phenotype [65].

*FGFR3* is also downregulated in WT [66]; however, in pediatric CNS tumors, opposite *FGFR3* expression profiles are observed. In NB, high expression levels are associated with worse overall survival and event-free survival (EFS) [67]. In glioma, its upregulation was associated with increased patient age [52], a feature that denotes a more invasive phenotype in adult counterparts [59]. Moreover, *FGFR3* amplification [68] and fusions (*FGFR3-TACC3*) seem to play a role in tumor metabolism and tumor growth promotion in low-grade gliomas (LGG) [69–71]. Such correlation with poor prognosis was also observed in EPN in which *FGFR3* was associated with shorter overall survival and shorter time to tumor recurrence [52].

Moderate-to-high expression of *FGFR3* mRNA was also observed in 80% of samples from EWS family tumors [71]. Mutations in this gene were also reported in circulating tumor cell samples [72]. However, neither of these studies presented information about correlations with clinical outcomes. Of note, when *FGFR3* is downregulated in OS cell lines (by long noncoding, microRNA or iRNA), there is a reduction in tumor growth and angiogenesis, reinforcing its relevance to this disease [73–76].

Regarding *FGFR4*, little information about its prognostic value has been published in the pediatric setting, with a few reports on glioma, MB, RMS and NB. The prognostic value of this kinase in the first group was initially evaluated by in 2019 by Jimenez-Pascual and Siebzehnrb1, who did not find any correlations between *FGFR4* expression and clinical outcomes [59]. Nevertheless, a recent evaluation of transcriptomic glioma datasets from The Cancer Genome Atlas (TCGA) revealed a direct association of high *FGFR4* expression and dismal prognosis, progressively upregulated in recurrent tumors. In addition, the contribution of *FGFR4* to the malignant phenotype of a highly aggressive GBM subgroup was further validated by increased viability, adhesion, migration and

clonogenicity in vitro, along with abolished xenograft formation in mice and reduced invasiveness in zebrafish xenotransplantation models [76].

In MB, high *FGFR4* expression levels were observed in HD-MBO3 cells and in a small cohort ( $n = 12$ ) of primary MB tissues, even though there was no validation upon TMA [63]. Of note, a pilot study based on an independent blinded set of 112 samples showed that protein levels of *FGFR4* in urine, together with cadherin-1 (*CADH1*) and fibrinogen beta chain (*FIBB*), could be used to discriminate MB patients from healthy control patients with acceptable accuracy. Moreover, the authors reported a positive correlation of urine *FGFR4* detection with the age of affected patients [77].

Additionally, *FGFR4* overexpression in RMS contributes to the failure of cells to complete normal skeletal muscle development, leading to constitutive signaling and unregulated growth in correlation with poor differentiation [78–81]. *FGFR4* mutations in childhood RMS (7–8% of tumors) are more frequently observed within the kinase domain. From those, N535K and V550E increase autophosphorylation of the receptor and promote proliferation and metastatic potential when expressed in vitro [79]. High expression levels of *FGFR4* have also been associated with advanced stage and poor survival in RMS [82,83]. Furthermore, *FGFR4* has been reported as a downstream target of *PAX3* and *PAX3-FOXO1* [81] and thus is commonly altered in fusion-positive RMS [83] and a key contributor to RMS invasion and metastasis [84].

Last but not least, a germline polymorphism in the *FGFR4* gene (rs351855) which results in the expression of an arginine at codon 388 (Arg388), rather than the more common glycine (Gly388), is frequently associated with decreased survival rates, treatment resistance and more aggressive disease in a variety of malignancies, and is associated with an increased prevalence of NB in children [85], and this association may be linked to differences in *FGFR4* degradation rates [86]. It was also observed that cases with the *FGFR4* AA genotype were 2.5 times more likely to have tumors with *MYCN* amplification compared with those with AG and GG genotypes, although such association was not statistically significant [85].

**EGFR.** The epidermal growth factor receptor (EGFR) (also recognized as HER-1 or ERBB-1) is a transmembrane glycoprotein of the ERBB receptor tyrosine kinase superfamily. Overexpression and/or enhanced activity of EGFR activate the downstream pro-oncogenic signaling, including the RAS-RAF-MEK-ERK and AKT-PI3K-mTOR pathways. These consequently activate several biologic expressions that proceed human cancer progression [87].

Overexpression of EGFR has been reproducibly detected in a large number of tumor samples and found to act as a strong prognostic indicator in head and neck, ovarian, cervical, bladder and esophageal cancers, correlating to poorer survival rates [88,89]. In the pediatric setting, however, there are few reports about its prognostic relevance. In gliomas, for example, fewer molecular alterations in the EGFR gene (mutations and amplifications) are observed in children when compared to adult counterparts [90–95]. *EGFR* gene amplification/overexpression is a genetic hallmark in adult GBM (observed ~40% of tumors) [95], whereas this feature is only observed in 25% of pediatric cases; nevertheless, it is associated in a similar manner with higher proliferation and increased tumor grade [96–98].

Likewise, high-level amplification and EGFR overexpression correlate with shorter event-free survival and relapse in high-grade EPN, being considered an independent prognostic marker for intracranial forms [99–101]. In MB patients, high expression of HER-2, another member of the EGFR gene family, was also associated with limited survival and metastasis [102,103]. Moreover, this RTK often co-expresses with HER-4 (more than 50% of samples), suggesting that HER-2/HER-4 heterodimerization may be of particular biological significance in this disease [101].

HER-2 expression was also reported to be associated with the aggressive behavior of NB and to significantly reduce survival [103]. However, a later study demonstrated that EGFR and HER-2 positivity are more frequently found in favorable histological risk groups, including younger age ( $\leq 18$  months), localized disease, and favorable histological group [104]. Similar results were obtained by Izycka-Swieszewska et al. (2010), where



HER-2-negative cases were more often found in the metastatic tumor group, associated with increased mitotic index and higher KI67 expression. *MYCN* non-amplified tumors were more often HER-2-positive than amplified tumors [105]. In contrast, higher expression levels of HER-4 are more often found in patients with metastatic disease [104].

For other pediatric tumors, the biological relevance of EGFR family members remains to be clarified. In EWS, for example, while HER-2 is not considered an important prognostic factor, an association of HER-4 and metastasis was found [106]. In OS, *EGFR* expression is common [107], but correlations between EGFR or HER-2 expression and clinical prognosis have been controversial, as no treatment improvements are achieved when EGFR is inhibited in pre-clinical and clinical trials [106].

In RMS, RB and WT, no strong associations of EGFR expression with clinical data have been found [108–111], although the ERBB family seems to be important for the malignant phenotype of RMS: ERBB1 sustains cell proliferation and growth, ERBB2 regulates myoblast cell transformation and survival and ERBB3 induces myogenic differentiation. Additionally, activation of ERBB2 coupled with inactivation of p53 induces RMS in animal models [106].

**VEGFR (KDR—kinase insert domain receptor).** The vascular endothelial growth factor receptor (VEGFR) family consists of three members: VEGFR1, VEGFR2 and VEGFR3 [112]. These receptors are established players in the formation of new blood vessels and the maintenance and remodeling of existing ones, during development and in adult tissues [113]. As such, in neoplastic growth, VEGFRs play an essential role in tumor neovascularization, providing oxygen and nutrition, and they facilitate tumor cells to metastasize and spread to distant organs [114]. Regarding pediatric cancer, VEGFRs have already been quantitatively evaluated in various types of refractory brain tumors [115]. Both VEGFR1 and VEGFR2 were detected in anaplastic astrocytoma tumor cells, MB and EPN samples [116–120]. Moreover, these receptors are frequently mutated and highly expressed in gliomas, NB and OS; in all cases, there is a negative correlation with unfavorable prognosis, advanced tumor stage, metastasis and shorter overall survival [121–127].

**RET.** Under normal conditions, the RET (“rearranged during transfection”) TKR pathway is activated by glial cell line-derived neurotrophic factor (GDNF) ligands that bind to coreceptors from the GDNF family receptor alphas (GFR $\alpha$ s), playing a major role during sympathetic and enteric nervous system development, where it signals toward proliferation, migration and differentiation. Apart from amplification, the constitutive activation of RET is caused by point mutations and gene rearrangements that drive malignancy in multiple tissues (i.e., papillary and medullary thyroid carcinomas and non-small cell lung carcinomas) [128–131].

RET rearrangements are also found in a high proportion of childhood papillary thyroid cancers [132–134]. Recently, it was observed that pediatric tumors (soft tissue sarcomas or medullary thyroid cancer) harboring either an RET-fused or RET-mutated pathogenic somatic alteration show clinical response to the RET inhibitor Selpercatinib [134].

RET mutations leading to dysfunctional ligand binding have also been described as the second most significant cancer-predisposing gene in the germline of patients with OS [135,136]. Moreover, RET is activated and can promote motility and colony formation in metastatic OS cells, contributing to the higher resistance of this tumor type to different chemotherapeutic agents [137–140]. Furthermore, NB cells and tumor samples demonstrated high RET expression levels [140], and its activation induces invasive spread NB in animal models [141].

**c-MET.** The mesenchymal–epithelial transition factor (c-MET), which is also known as hepatocyte growth factor receptor (HGFR), is an essential molecule for the survival and function of normal cells that promotes tissue remodeling and organ homeostasis [142]. MET’s gain of function either via overexpression, amplification, aberrant splicing or mutations is associated with the constant activation of downstream classic signaling pathways that sustain rapid proliferation, promote cell migration, angiogenesis and survival of cancer cells [143]. Moreover, recent evidence indicates that MET signaling participates in the acquirement of mesenchymal phenotype, tumor plasticity and adaptive responses to

metabolic stress, contributing to the recurrence and metastatic dissemination of cancer cells [144,145].

The c-MET gene was first identified in the human OS cell line (HOS) that had been treated with N-methyl-N'-nitro-N-nitrosoguanidine (MNNG) as a gene able to transform normal fibroblasts [146]. Since then, its involvement in cancer establishment and progression has been repeatedly described in a variety of common and high-risk pediatric solid tumors, including not only sarcomas, but also gliomas, MB, NB, WT and hepatoblastomas, among others [147]. Of note, infantile hemispheric gliomas were recently recognized to be driven by different RTKs, including somatic fusions and alterations involving ALK, ROS1, NTRK and c-MET [148]. Especially in anaplastic, diffuse and PA, c-MET levels often correlate with tumor grade [149].

Cytoplasmic c-MET immunoreactivity is also associated with poor clinical outcome, and tissues with overexpression often exhibit higher vascular proliferation and proliferative index [150–152]. Similar phenotypes have been observed in NB, where overexpression of this receptor promotes invasion and is associated with advanced metastatic stage [153,154].

Likewise, high levels of MET protein are associated with increased proliferative activity invasion and metastasis in WT [155,156], and represent a risk factor for invasion in RB [156].

In childhood sarcomas, several studies have pointed out c-MET as a promising biomarker capable of predicting poor prognosis. Forced expression of MET in primary osteoblasts induces transformation and is essential for the maintenance of the cancer phenotype [157], while loss-of-function approaches in OS cell lines (143B and U2OS) demonstrated that this oncogene promotes cell proliferation, migration and invasion, and inhibits cell apoptosis [158]. However, the study of genomic status of MET and other genes implied in ossification processes in a cohort of 91 children and teenagers showed that *MET* is mainly deleted, although the clinical subgroup with MET amplification presents worse outcomes [159].

In EWS, modest to high MET cytoplasmic/membranous expression is detected in the majority of tumor samples and is significantly correlated with a poor overall survival. However, there were no significant correlations between MET expression and clinical characteristics, including tumor stage, tumor location and age at diagnosis. The same group also detected genetic alterations that result in the formation of truncated MET proteins in 5% of patients and in two cell lines (ES-2 and ES-7) [160].

Finally, this RTK is overexpressed in RMS tumor samples [161–163] and cell lines, contributing to the metastatic and invasive features of this tumor type [163,164].

**ALK.** This RTK was first described in 1994, as a fusion partner in the t(2;5)(p23;q35) chromosomal translocation characteristic of anaplastic lymphoma from which takes its name [165]. In general, ALK activates multiple signaling cascades, such as the PI3K-AKT, CRKL-C3G, MEKK2/3-MEK5-ERK5, JAK-STAT and MAPK pathways, and its role in cancer may vary due to many factors, including not only its fusion partners (more than 30 described so far), but also the tumor type or its genetic background (its effects on NB, for example, are dependent on *MYCN* status) [166].

Next-generation sequencing has revealed the presence of several *ALK* mutations in pediatric cases with RMS, EWS, WT and OS [161,167–170]. Most mutations are located within the kinase domain and can be divided into three groups: ligand-independent mutations (F1174I, F1174S, F1174L and R1275Q), ligand-dependent mutations (D1091N, T1151M and A1234T) and a kinase-dead mutation (I1250T) [167].

Germline gain-of-function point mutations are observed in half of hereditary NB and in 9% of the sporadic forms [170], and correlate with high risk and poor prognosis [171,172], mainly because both the wildtype and mutant forms of ALK induce *MYCN* transcription and potentiate its oncogenic activity in this tumor type [173]. Other ALK-driven pediatric tumors include infantile hemispheric gliomas [149,174], inflammatory myofibroblastic tumors, renal cell carcinomas [167] and pediatric mesotheliomas [175].

ALK in-frame translocations have been described in EPN and EWS as detected by fluorescent in situ hybridization with the break-apart of 5' and 3' probes [160,176]. Finally,

ALK expression is strongly associated with the WNT-activated MB subtype in which, differently from other pediatric tumors, it represents an independent indicator of good prognosis for medulloblastoma patients [177].

### 3.1.2. PI3K/AKT/mTOR Pathway

The phosphatidylinositol 3-kinase (PI3K)/Akt/mammalian target of rapamycin (mTOR) signaling pathway is among the best investigated in human biology, and is considered a key player in both physiological and pathological conditions [178].

The first step of activation of this pathway consists of the recognition of various growth factors and cytokines by RTKs localized at the cytoplasmic membrane. Then, these receptors dimerize and undergo autophosphorylation, activating GRB2 (Growth Factor Receptor Bound Protein 2) and SOS (Ras/Rac Guanine Nucleotide Exchange Factor). These activate Ras through the exchange of GDP with GTP, which then phosphorylates and activates the PI3K [179]. Active PI3K catalyzes the conversion of PIP2 (phosphatidylinositol 4,5-bisphosphate) to phosphatidylinositol-3,4,5-trisphosphate (PIP3), a second messenger that binds and recruits AKT to the cell membrane, which causes a conformational change in AKT and makes it more accessible to the PDK1-mediated phosphorylation of Thr308, followed by the phosphorylation at serine-473 by the mTOR2 complex [180]. This activation then induces a detachment of AKT from the inner surface of the plasma membrane, and the relocation to the nucleus where AKT isoforms phosphorylate and modulate the activity of several transcription factors. More than 100 different effectors have been described, including cyclin-dependent kinase inhibitor kip1 (p27kip1) through the FOXO family of Forkhead transcription factors, glycogen synthase kinase 3 (GSK3) and cell cycle stimulators, including cyclin D1 and c-Myc. AKT can regulate apoptosis by the inhibition of Fas ligand (FasL), BCL2-associated death promoter (BAD), BCL-2-interacting mediator of cell death (BIM) or BCL-2-associated X-protein (BAX), and by the degradation of p53 [181].

AKT also activates mTOR1, which has many different targets, including translation transcription factors that initiate transcription of genes associated with cell survival and growth and factors associated with hypoxia and angiogenesis (Figure 2) [182].

In cancer, this pathway can be dysregulated as a result of the activation of upstream oncoproteins including RTKs, by the loss or decreased level of its negative regulators such as the phosphatase PTEN (phosphate and tensin homolog deleted on chromosome 10), or directly through mutation and overexpression [183].

Undeniably, this pathway is activated in a wide variety of tumors (i.e., prostate, breast, lung and leukemia, among many others), leading to a profound disturbance of cell growth control, metabolic reprogramming and invasion/metastasis, as well as the suppression of autophagy and senescence [179,184–189]. Moreover, increasing evidence points to its critical participation in the maintenance of stemness in a variety of cancers, contributing directly to recurrence and chemoresistance [189].

In the pediatric setting, the PI3K/AKT/mTOR signaling axis has been described as abnormally activated in both hematologic and solid tumors, mainly as a consequence of chromosomal gains amplifying the AKT1 gene (described in rare cases of leukemia) [190–192] or the aberrant expression of PI3K isoforms [193]. Below, the involvement of this pathway's individual members in childhood tumors is explored.

**PI3K.** PI3K is a group of plasma membrane-associated lipid kinases, consisting of three subunits: p85 regulatory subunit, p55 regulatory subunit and p110 catalytic subunit. According to their structure and substrate specificity, these kinases are grouped into three categories (classes I, II and III) [194,195]. PIK3CA (phosphatidylinositol 3-kinase, catalytic,  $\alpha$ -polypeptide), the gene encoding the p110 $\alpha$  subunit, is frequently mutated in ~30% of common human cancers and has been studied most thoroughly [196,197]. Although numerous mutations in this gene have been described, most gain-of-function mutations cluster around two hotspots at exons 9 and 20 [197]; however, contrasting roles for mutations at each exon have been described depending on the tumor type [198,199]. PI3K amplifi-

cations have also been frequently described and correlated with aggressive phenotypes, chemoresistance and poor prognosis [200–203].

The prognostic power of PI3K alterations in childhood cancer has been less explored. In MB, however, although no mutations have been detected [196], the p110 $\alpha$  isoform is typically overexpressed, promoting cell proliferation, chemoresistance and migration [204–206].

Dysregulation of PI3K signaling is also considered an important player in gliomagenesis, with key roles in regulating cell movement and thus contributing to the highly invasive phenotype of GBM. Compared with normal human astrocytes, overexpression of PI3K p110 catalytic subunits, p85 regulatory subunits and phosphorylated Akt (Ser473) was also detected in two pediatric GBM cell lines (GBM6840 and GBM2603) [206]. Likewise, overexpression of the catalytic p110 $\delta$  and regulatory p85 $\alpha$  isoforms was also detected in a panel of primary NB samples and cell lines with active roles in cell growth and survival. Especially, p110 $\delta$  was correlated with *MYCN* amplification [207]. However, this gene is significantly lower in NB samples with loss of heterozygosity at 1p36 and associated with poor clinical outcome [208–210].

The regulatory domain of PI3K, p55, is upregulated in sarcoma stem-like cells and promotes invasion, migration and chemotherapy resistance [210]. In EWS, despite variable expression levels between samples, this positive regulator has an oncogenic role [211]. Moreover, p55 analysis on a human sarcoma TMA (that includes two EWS samples) performed by Yoon et al. demonstrated a 4.1-fold increase compared with normal tissues [210].

**AKT (PKB).** AKT or PKB (protein kinase B) is a serine/threonine kinase that functions as an important regulator of cell growth, survival and glucose metabolism. There are three isoforms of mammalian AKT which are encoded by different genes [212]. AKT1 and AKT2 are ubiquitously expressed and are mostly involved in regulating cellular survival and protein synthesis, involved in glucose transport through the insulin signaling pathway, respectively. The function of AKT3 is not yet fully understood and its expression is almost entirely limited to the nervous system tissue [213–215]. Nevertheless, it has been reported that despite the high similarity, AKT isoforms exert non-redundant specific effects under physiological and pathological conditions [215].

Gain-of-function mutations in all three AKT genes have been identified in ~40% of breast, colon, melanoma and ovarian cancers [216,217]. G49A mutations affecting the pleckstrin homology domain of AKT1, for instance, were identified in ~5% of breast, colorectal and ovarian cancers [217]; however, this mutation was not detected in any of the 100 cases of GBM or 75 cases of MB analyzed by Schüller et al. in 2008 [218]. In the pediatric population, MB samples show p-AKT, and cell lines have shown to be crucially dependent on PI3K/AKT pathway activation; however, the phenotype was attributed to PTEN inactivation as a result of the loss of heterozygosity of chromosome 10q or promoter [219].

In pediatric sarcomas, Akt1 has been shown to contribute to the maintenance of the undifferentiated state of myoblasts pointing towards Akt signaling as a critical RMS nodal point [220]. The AKT pathway is also considered to be an important mediating survival signal in EWS [221]. Likewise, an increasing body of evidence has shown that this pathway is frequently hyperactivated in OS and contributes to disease initiation and development, including tumorigenesis, proliferation, invasion, cell cycle progression, inhibition of apoptosis, angiogenesis, metastasis and chemoresistance [222–224]. The AKT2 gene is significantly upregulated in chemoresistant OS cell lines [224] and tumor samples, being significantly associated with positive recurrence, the presence of metastasis, poor response to chemotherapy and shorter EFS and overall survival [224]. Moreover, the AKT3 isoform was evidently upregulated in OS tissues and positively associated with tumor size [225].

AKT2 also plays an important role in NB by regulating N-myc expression. Of note, attenuation of this AKT isoform impaired proliferation and anchorage-independent cell growth, and decreased the secretion of angiogenic factor VEGF and decreased the potential



to metastasize to the liver in vivo, thus implicating AKT2 in multiple aspects of NB initiation and progression [226].

**mTOR.** mTOR is a serine/threonine protein kinase that forms the catalytic subunit of two structurally and functionally distinct protein complexes, known as mTOR Complex 1 (mTORC1) and 2 (mTORC2) [227]. mTORC1 consists of mTOR, Raptor, G $\beta$ L (mammalian lethal with SEC13 protein 8) and domain-containing mTOR-interacting protein (DEPTOR), and plays active roles in integrating various signals that specify the availability of growth factors, nutrients and energy in order to endorse ribosomal biogenesis, protein translation during cell growth and the expression of metabolism-related genes, while inhibiting apoptosis and autophagy [228]. mTORC2, on the other hand, is composed of mTOR, Rictor, G $\beta$ L, Sin1, PRR5/Protor-1 and DEPTOR, and regulates cytoskeletal dynamics, ion transport and promotes cell proliferation and survival through the activation of Akt [229–231].

mTOR is frequently improperly activated in human cancers and results in alteration of both mTORC1 and mTORC2 signaling pathways, leading to increased cell proliferation and decreased apoptosis. However, among 33 mTOR activating mutations identified in 2014 by Grabiner et al. [231], those that were functionally tested in vitro conferred varying degrees of pathway activation, and, most importantly, a few displayed some substrate preference towards the eukaryotic translation initiation factor 4E binding protein 1 (4EBP1) and ribosomal protein S6 kinase (S6K1), or towards AKT1, implying that such mutations had distinct effects on mTORC1 or mTORC2. Specifically, 4EBP1 activation by mTOR1 is a major contributor to accelerated cell proliferation or increased cell survival; the so-called eIF4E-sensitive mRNAs code for various cell cycle and apoptosis regulators, including cyclins D1 and D3, CDK2, MYC, PIM1, Bcl-2, Bcl-xL and VEGF, among others [232–234].

mTOR overactivation is observed in many childhood tumors, including EPN, MB and PA, high-risk NB, WT and RB, leading to worse prognosis and survival [105,235–241].

Constitutive activation of the mTOR pathway, predominantly through mTORC2, is observed in EWS, with active roles in metastasis formation [241–244]. The metastatic behavior of OS is also dependent on the PI3K/Akt/mTOR cascade, in which mTOR contributes to cellular transformation and poor cancer prognosis via its downstream effectors S6K1, 4EBP1 and eIF4E [245,246]. In RMS, lower disease-free or overall survival is also associated with the activation (phosphorylation) of multiple interconnected Akt/mTOR pathways [246]. Of note, rapamycin treatment can greatly reduce the growth of cell lines derived from these three sarcoma types [242].

**GSK-3.** The glycogen synthase kinase is a ubiquitously expressed serine/threonine kinase existing as GSK-3 $\alpha$  and GSK-3 $\beta$  isoforms (encoded by separate genes), both of which are downstream effectors of AKT [247]. Differentially from other kinases, GSK3 is one of the few whose activity tends to be high in resting cells, and exposure of cells to growth factors, serum or insulin results in its catalytic inactivation [248].

The GSK3 kinases are pleiotropic, phosphorylate many proteins, and interact with multiple signaling pathways [249]. These kinases can modify the activity of transcription factors that have profound regulatory roles in cellular proliferation (such as p53 and NF- $\kappa$ B), transcription factors important for epithelial–mesenchymal transition (EMT) (i.e., Snail) and pro-apoptotic molecules including BCL2 and BAX [250]. Therefore, aberrant activity of GSK3s can result in many diseases and disorders and influence oncogenesis and metastasis [251]. However, since GSK3s are involved in a wide range of signal transduction cascades and a plethora of cellular functions [252,253], their roles in cancer establishment and maintenance can deviate from their chief tumor suppressor effects and also promote neoplastic transformation [254–257]. This dichotomy is also observed in the pediatric setting. Strong evidence provided by Wang et al. (2008) [257], for example, demonstrated that GSK-3 activity is essential for the maintenance of MLL-positive leukemias. MLL rearrangements are in >70% of infant leukemia, and irrespective of the translocation partner, they are associated with poor clinical outcomes [258–260]. Alternatively, as a key suppressor of the Wnt, Hedgehog and Notch pathways GSK3 has attracted much scrutiny. Within these pathways, this kinase is critical in regulating the turnover of the effectors  $\beta$ -catenin, c-Myc

and c-Jun, targeting them for degradation/inactivation, and this inhibits proliferation and stem cell maintenance [250].

The literature about the prognostic value of GSK3A in pediatric cancer is scarce. No evidence was found in the literature about its involvement in EWS, OS, RMS, WT, RB, NB and EPN. However, its role in MB has been explored in vitro and in vivo, showing to be important for cell proliferation and tumor growth [260], a phenotype that seems to be similar in pediatric glioma [261].

On the other hand, the role of GSK3B is more extensively studied. In EWS, this gene can either promote or impair tumor growth and is associated with good prognosis [262–265]. Interestingly, in OS, the same gene acts as an oncogene [265], and is associated with worse response to neoadjuvant chemotherapy [266].

The oncogene status also occurs in alveolar RMS, where GSK3B is directly involved in regulating the transcriptional activity of PAX3/FKHR [267] at the same time that the chimeric protein enhances GSK3B activity, which in turn represses MYOGENIN, a member of the muscle regulatory factor family that orchestrates the terminal differentiation step of skeletal muscle cells [268]. GSK3B is also involved in the maintenance of undifferentiated phenotypes in ERMS [269].

GSK-3B is highly expressed in high-risk NB; however, its expression is not associated with clinical stage, survival or other clinicopathological parameters [270]. GSK-3B has also been involved in the protection of NB cells against chemotherapy by regulating NF- $\kappa$ B signaling [271]. In this regard, several authors have shown that GSK3 inhibitors are able to regulate MYCN mRNA levels and reduce NB cell viability through multiple mechanisms, including p53 and Wnt signaling, BDNF/TrkB/PI3K/Akt, suggesting that targeting this kinase might potentiate chemotherapy [271–274].

Alternatively, a predominantly tumor-suppressive role for GSK3B is observed in MB, in which its accumulation leads to the downregulation of GLI, the most important activator and driver of the SHH medulloblastoma subtype [274]. Constitutive phosphorylation leading to GSK3 $\beta$  activation improves cell survival and contributes to malignant transformation [275]. Dysregulated GSK3B also sustains the survival, immortalization, migration, invasion and maintenance of stem cells in glioma [276–278].

### 3.1.3. MAPK Pathway

The mitogen-activated protein kinases (MAPKs) comprise a group of serine–threonine protein kinases that control numerous cellular processes, including proliferation, differentiation, apoptosis, survival, inflammation and innate immunity [278]. In mammals, MAPKs include three main signaling axes, namely c-Jun NH<sub>2</sub>-terminal kinase (JNK), p38 MAPK and extracellular signal-regulated kinase (ERK), each of which exists in several isoforms [279].

This pathway mediates intracellular signaling triggered by extracellular stimuli such as growth factors and cytokines (ERK), or by intracellular stimuli such as genotoxic, osmotic, hypoxic, oxidative or endoplasmic reticulum (ER) stress (JNK and p38), for example [280,281].

The general cascade pattern includes initial activation of MAP4Ks (membrane-bound GTPases such as RAS, RHO, RAN, RAB and ARF) by RTKs, which phosphorylate intermediate MAP3Ks (i.e., RAF, MEKK). These then mediate phosphorylation and activation of MAP2Ks (MEK1/2—mitogen-activated protein/extracellular signal-regulated kinases, MKK4/7), followed by the positive phosphorylation of MAPKs (ERK1/2, p38 or JNK). Once activated, MAPKs phosphorylate diverse substrates, including transcription factors such as c-Jun, c-Myc, P53 and ATF2, thereby giving rise to the various cellular responses [282,283]. p38 MAPKs have also emerged as important modulators of gene expression by regulating chromatin modifiers and remodelers [281].

Thus, compromised MAPK signaling contributes to the pathology of a wide spectrum of human malignancies [282]. However, while the roles of JNK and p38 pathways are

elusive [284–290], dysregulation of the RAS/RAF/MAPK(MEK)/ERK pathway explicitly drives the oncogenic process [291].

In fact, many of the cancer-associated mutations of components of MAPK signaling pathways have been found in RAS. Missense gain-of-function mutations in all three RAS genes (HRAS, KRAS and NRAS) are found in ~30% of all human cancers [292–294]. Other perturbations in GDP–GTP regulation, persistent receptor tyrosine kinase-mediated activation of GEFs, and miRNA deregulation are additional mechanisms of RAS activation in cancer and result in constant input signals with downstream kinases [291,295].

In this regard, the frequency of genomic alterations in the MAPK pathway as a whole parallels the direction of the signaling cascade: RAS > BRAF > MEK, and ERK mutations are exceptionally rare [296,297].

**RAF.** RAF has three isoforms (ARAF, BRAF and CRAF/RAF1), sharing a high similarity of domain organization. These cytoplasmic serine/threonine-specific protein kinases are essential effectors of the MAPK pathway through the association with activated RAS. This binding leads to their homo- or heterodimerization and activation with the phosphorylation of ERK.

Altered activation of RAF members results in increased proliferation in a broad range of human tumors [298,299]. The most common gain-of-function mutation in the members of the family occurs in BRAF codon 600, in which a valine is substituted for glutamic acid (BRAF-V600E). This point mutation is notably widespread in pilocytic astrocytoma (15%), melanomas (63%) and papillary thyroid carcinomas (more than 50%) [300,301]. Overexpression of full-length RAF or the truncated catalytic domain also leads to hyperactivated ERK signaling, resulting in increased malignant behavior [302].

**ERK1/2.** ERK1 and ERK2 are the prototypes of the eight isoforms of ERK and are activated by MAPK/ERK kinase (MEK) 1 or 2. Upon activation, ERK detaches from cytoplasmic anchoring proteins and translocates to the nucleus to exert its transcriptional regulation. Despite the well-recognized importance of ERK activation in cancer malignancy, mutations in these genes have rarely been reported as drivers in human cancers. Nonetheless, The Human Protein Atlas classifies them with enhanced expression compared to normal tissues.

The most compelling evidence of MAPK activity in cellular processes contributing to the development and progression of childhood tumors is represented by the duplication/rearrangement of *BRAF* at 7q34 leading to *KIAA1549:BRAF* fusion product, which is the most common molecular alteration in sporadic PA, occurring at the highest frequency in tumors of the posterior fossa [303–305]. ERK2 was identified as differentially expressed in tumor samples compared to normal tissues [306]. RAS/MAPK activation was associated with metastatic disease in MB [307,308]. Associations of ERK hyperexpression with distant metastasis and poor overall survival were also reported for childhood sarcomas, including RMS and OS [309–312]. For other pediatric tumors, activation of this pathway results from their interaction with dysregulated microRNAs [313,314].

#### 3.1.4. Cell Cycle Kinases

The cell cycle is a complex and well-ordered series of irreversible events through which a cell duplicates its DNA and grows to produce two daughter cells with identical genomes [315,316]. Transitions from one state to the next are driven by many oscillating regulators that determine whether cells proceed through G1 into the S phase, and from G2 to M, each of which are characterized by distinct molecular features and functional outputs [317]. Central to this process are the cyclin-dependent kinases and other key regulators such as kinases from the Polo and Aurora families.

##### Cyclin-Dependent Kinases

Cyclin-dependent kinases (CDKs) comprise 13 key regulatory enzymes involved in cell proliferation through the regulation of cell cycle checkpoints and transcriptional events in response to extracellular and intracellular signals. These intracellular serine/threonine

kinases, whose catalytic activities are regulated by interactions with the adaptor molecules cyclins and CDK inhibitors (CKIs), orchestrate the evolution through the sequential phases, including entry into the cell cycle from quiescence, the G1/S phase transition, DNA replication in the S phase, nuclear breakdown, chromosome condensation and segregation, and cytokinesis [318].

CDKs coordinate cell cycle regulation at different stages to ensure the coherence, integrity and maintenance of every step in a sequential manner. CDK1 and CDK2, for instance, are necessary to direct the transition from S to G2, but only CDK1 governs the G2/M transition and mitotic progression [319]. Other CDKs regulate the cell cycle indirectly by activating other members of the family (CDK7, CDK20) or transcription (CDK7, CDK8, CDK9, CDK12, CDK19) [320,321].

Changes in the expression and regulation of CDKs induce unscheduled proliferation and chromosomal instability, well-known hallmarks of cancer and tumor aggressiveness [322–327]. Amplification or mutation of genes encoding CDKs, cyclins or endogenous inhibitors of CDKs have been described in many solid cancer types, and are recurrent events in the development of breast cancer [328] and GBM [329], for example. Such alterations are also described as molecular drivers in childhood tumors [330–334].

**CDK1.** Cyclin-dependent kinase 1 (CDK1) is vital in governing cell division and transition from G2 to the M phase [335]. Its dysregulation is common in many tumors of diverse origins, leading to chromosomal instability via replication stress and enhanced proliferation of cells. A recent pan-cancer integrative analysis based on TCGA and GTEx databases performed by Liu et al. (2022) showed that CDK1 expression levels are increased in many tumor types when compared to normal tissues and are generally associated with poor clinical prognosis [336]. For example, CDK1 expression is positively and highly associated with advanced cancer stages in lung and endometrial cancer [337,338]. Similar results were reported for other tumors, such as breast [339–341]. Moreover, CDK1 expression is positively correlated with the expression of the stemness marker SOX2, indicating a direct action on tumor maintenance and chemoresistance [342,343].

Regarding pediatric tumors, this kinase is associated with lower overall survival and EFS rates in EPN [344,345], RMS [346,347] and NB [348]. In silico analyses have also demonstrated that CDK1 is differentially expressed in RB [349] and plays a key role in the development of OS, since its negative regulation or depletion leads to significant decreases in proliferation while inducing apoptosis [350–353].

Furthermore, according to the literature, a well-described relationship exists between CDK1 and EWS, WT and high-grade gliomas (HGG). Specifically, this kinase expression has been directly associated with tumor progression [354], being considered a hub gene for GBM [355].

**CDK2.** Cyclin-dependent kinase 2 (CDK2) drives the entry of cells into the S and M phases of the cell cycle. Except for a few exceptions (i.e., testis), the majority of normal tissues have low expression of this serine/threonine kinase [356], and its activity is not essential for normal development [357]. However, CDK2 has been associated with cancer progression and aggressiveness across several malignancies [339,358], contributing not only with genomic instability and under-replication of DNA in the late S phase [359] but also through interactions with other proteins in a wide range of biological processes such as DNA damage response, intracellular transport, protein degradation and signal transduction, among others. Differentially from other kinases, several investigations have demonstrated that CDK2 is not upregulated or amplified; instead, its dysregulation results from altered binding partners or alterations due to post-translational modifications [360]. In tumors with MYCN overexpression, as is the case of NB, interaction with CDK2 appears to be critical for senescence avoidance and immortalization [361], being associated with worse prognosis and considered a suitable therapeutic target in this tumor type [362,363]. CDK2 inhibitors effectively induced cell cycle arrest or apoptosis in MYC-driven MB [364].

The literature also shows overexpression of CDK2 in HGG compared to normal tissue and low-grade forms, with a direct association with worse prognosis due to immune cell



infiltration [365]. Moreover, this kinase plays a central role in the development of NB. Even though there is no well-established relationship between this kinase and the development of this tumor type, Zhang et al. (2016) [366] demonstrated that the inactivation of TAZ (a biomarker of aggressiveness in RB through miR-125a-5p) inhibited proliferation and tumor formation by decreasing cyclin E and CDK2 expression [367]. Similarly, although there is no clear and explicit description in the literature about the relationship of this kinase with RMS, Knudsen et al. also discussed the relationship between sustained CDK2 levels in RD cells irrespective of the exposure of cells to differentiating culture media, explaining the inability of those cells to arrest growth and thus contributing to oncogenesis [368]. Moreover, there are reports of apoptosis induction in several sarcoma cells after CDK1 and CDK2 co-depletion [369,370]. Of note, in a microarray-based study, CDK2 was found to be overexpressed and associated with poor prognosis in EWS [371].

**CDK4/6.** Cyclin-dependent kinases 4 and 6 are highly homologous key components of the cell cycle to drive the passage from G1 to S phase. Upon interaction with any D-type cyclin (CCND1, CCND2 or CCND3), these interphase kinases phosphorylate Rb to release E2F from Rb and initiate the transcription of genes required for cell cycle progression. Besides proliferation, other roles of cyclin-D/CDK4/6 have been confirmed, including the regulation of senescence, apoptosis, migration/invasion and angiogenesis [372].

Consequently, the complex CCND/CDK4/6 shortens G1, and hence, its constitutive activation represents a driving force of tumorigenesis. These proteins are generally concurrently studied and, in many cases, they present themselves with similar patterns, being simultaneously dysregulated [373,374].

CDK4 was identified as a major risk factor for disease progression in Paget's disease [375] and its overexpression and/or hyperactivation is implicated in many types of human cancers [376–381]. Point mutations at the CDK4 locus (CDK4R24C) have also been reported [382].

Co-overexpression of both CCND1 and CDK4 is common in hepatoblastoma, a rare malignant liver tumor of childhood, and usually positively correlated with tumor recurrence [383]. In parallel, enhanced kinase activity of CDK6 has been associated with other childhood tumors [384]. This kinase plays an important role during hematopoiesis and is frequently altered in hematological malignancies of different immunophenotypes [385,386]. MLL-AF9 oncofusions in myeloid leukemia, for example, induce high CDK6 levels, acting as a blocker of myeloid differentiation and contributing to the maintenance of an immature phenotype [387]. This MLL fusion-driven activation of CDK6 (through MLL-AF4 and MLL-ENL) has also been described in infant leukemia [388].

Likewise, CDK4 and CDK6 have been described with similar frequencies in WT compared to normal mature kidneys, even though only CDK4 showed correlation with relapse [389]. However, a more recent study by Haruta et al. (2019) showed that WT samples with chromosome 12 trisomy does indeed show upregulation of this kinase, but that stronger expression is associated with better overall survival [390].

Activation of the CDK4/6 pathway is also a powerful driver of sarcomagenesis [391]. Amplification of 12q13-15 also occurs in OS, and a recent copy number analysis of pediatric high-grade OS detected a recurrent gain of chromosome 12q14.1 in ~25% of samples, which resulted in CDK4 overexpression. In vitro, higher expression of CDK4 was considered a predictive biomarker for resistance to cisplatin [392]. Indeed, elevated CDK4 expression is correlated with metastasis potential and poor prognosis in this tumor type [393–395]. Consistent with these findings, a recent study demonstrated that about 50% of OS samples present CDK4 somatic variants, 9.5% of which were identified as gain-of-function CNVs correlated with metastasis and death [396].

The inhibition of CDK4/6 also represents a promising precision medicine-guided therapy for other childhood sarcomas. A parcel of PAX3/PAX7-FOXO1-positive RMS tumors with amplification of the chromosomal region 12q13-q14, for example, also presents elevated CDK4 levels relative to non-amplified, fusion-negative forms [397,398]. In addition, in Brazilian cohorts, amplification or overrepresentation of CDK4 was evinced

through qRT-PCR and immunoreactivity in both forms of RMS (ERMS and ARMS), along with several leiomyosarcoma samples [399,400]. Similarly, using a human TMA, Saab et al. (2007) demonstrated CDK4 expression in 82% of ARMS and 63% of ERMS tumors [401]. CDK6 was detected at high levels in six RMS-derived cell lines, reinforcing the prospects of its inhibition as a therapeutic opportunity [402]. In a similar manner, a shRNA-based screening demonstrated that CDK4 (together with CCND1) is required for survival and anchorage-independent growth in EWS [403].

More recently, a systematic evaluation of CDK4/6 as targets in a series 16 pediatric cancer types indicated that further preclinical evaluations are still needed to affirm the dependence of tumors on CDK4/6. Nevertheless, the results provided evidence for benefits in EWS, malignant peripheral nerve sheath tumors and MB [391]. Of note, within MB subgroups, CDK6 and CDK14 co-amplifications were identified in 20% samples from patients with relapsed group-4 MB [330].

Shubert et al. (2022) also pointed out that patients with atypical rhabdoid tumor/malignant rhabdoid tumor, NB or HGG may also benefit from anti-CDK4/6 therapy [391].

CDK4 and CDK6 are both highly expressed in NB compared to normal tissues [404]. Moreover, like CDK2, CDK4/CDK6 exert oncogenic roles in this tumor type, especially in MYC-amplified forms [405]. Additionally, when co-amplified with MDM2/FRS2, CDK4 and CDK6 are associated with poor prognosis and atypical clinical features, including poorly differentiated or undifferentiated histology and metastasis at diagnosis and at relapse [406].

In line with Schubert et al. (2022) [391], CDK4/6 upregulation also plays an important role in the pathogenesis and progression of high-grade gliomas with potential actionability [407–409]. However, the use of CDK4/6 inhibitors alone did not show satisfactory results, suggesting the use of combinatorial intervention [410].

CDK4 was likewise found overexpressed in EPN and associated with adverse outcomes [411]; accordingly, its inhibition restricted cell proliferation and reduced the expression of genes associated with the cell cycle and DNA repair (*CCNB1*, *TOP2A*, *CDK2*, *BRCA1* and *RAD51*), and induced morphological changes that culminated in cell death [412]. Considering EPN subgroups, De Almeida Magalhaes et al. (2020) showed that *CDK6* is overexpressed in ST-EPN-RELA tumors compared to other ST-EPN subgroups [413], even though others have suggested that the dysregulations of the p16-CDK4/6-pRB-E2F pathway might also compose the genetic background underlying the aggressive biology of posterior fossa EPN in infants less than 1 year old [414].

**CDK5.** The cyclin-dependent kinase 5 (CDK5) represents an unusual member of the family of cyclin-dependent kinases, which is activated upon binding to p35 and p39 proteins, which are not cyclins. Conversely, interactions with CCND1 or CCND can attenuate CDK5 activity [415]. CDK5 is expressed ubiquitously, but with higher activity in the nervous system, participating in neuron migration, neurite outgrowth and synaptogenesis. Nevertheless, increasing evidence points to a diverse array of functions in other tissues, ranging from cell proliferation to cytoskeleton remodeling and cell motility by regulating actin dynamics [416–418].

Apart from neurodegenerative disorders, amplification and increased expression of CDK5 have been described in multiple tumor types and are associated with worse prognosis and stemness [419–431]. Mutations located in key domains of CDK5 that influence its structure and post-translational modifications have also been described as contributors to tumorigenesis [432].

The participation of CDK5 in pediatric tumors is purported; however, the stimulation of cancer-related signaling pathways by this kinase remains obscure, and reports are scarce. CDK5 was found to be hyperactivated in NB and its inhibition resulted in cell cycle arrest and morphological differentiation [433,434]. Moreover, as a crucial regulator of neuronal signal transduction, CDK5 can be found differentially expressed in gliomas, progressively

augmenting with tumor grade, suggesting an active role not only in tumorigenesis but in aggressiveness as well [424,435,436].

CDK5 also appears to be a central regulator of OS tumorigenesis, with high levels of expression being associated with low survival and increased angiogenesis [437,438]. Interestingly, CDK5 also plays a role in osteoblastic differentiation. Fu et al., for example, demonstrated that CDK5 inhibition promotes the expression of *Runx2*, *ALP*, *OCN* and *OPN* in mesenchymal stem cells, the mineralization of MC-3T3E1 cells and suppresses the migration of the OS cell line MG-63 [439]. Additionally, the CDK5/p35 complex strongly inhibits the Wnt/beta-catenin signaling pathway, also able to stimulate osteoblastic differentiation [440].

The WNT pathway defines a molecular subgroup of MB [441]; thus, it may be assumed that CDK5 might also contribute to this tumor malignancy. In fact, *Cdk5* expression has been demonstrated in different MB cell lines and in a reduced cohort of patients; however, its deletion did not alter proliferation, reflecting the more favorable prognosis of MB with WNT activation [441]. Nevertheless, a role for CDK5 in tumor immune evasion through the regulation of PD-L1 was suggested [442].

**CDK7/9.** Cyclin-dependent kinase 7 (CDK7) and 9 (CDK9), apart from directing cell cycle progression, have critical roles in transcription initiation and elongation as regulators of the phosphorylation of the carboxy-terminal domain (CTD) of RNA polymerase II (CDK7 is a component of TFIIF, and CDK9, a subunit of pTEFb) [443–445]. CDK7/9 also controls many transcription factors, functioning to either promote their activity and/or regulate their turnover [445]. Recently, other uncovered transcription-associated functions have been revealed, including epigenetic modifications and mRNA-3' termination [446–448].

CDK7/9 levels are elevated in several cancer types and are associated with clinical outcomes [449–455]. In many cases, they can indirectly impact gene expression profiles by aberrantly controlling the functioning of transcription factors that are critical in specific tumor types, as is the case of estrogen- or androgen receptor-mediated transcription in breast and prostate cancer, respectively [449,456,457]. MYCN-dependent transcription can also be affected, as demonstrated in NB cells, or contribute to histone-3 methylation in diffuse intrinsic pontine glioma (DIPG) [445,458].

With regard to other pediatric tumors, CDK7 has been shown to be upregulated in a panel of OS cell lines and tumor samples, being associated with worse prognosis and higher metastasis rates [459]. Accordingly, CDK7 knockdown in SJS-1 cells reduced phosphorylation of the RNAPII CTD and reduced tumor volume and weight in xenograft models compared with tumors derived from wild-type cells [460]. Similarly, higher levels of expression of CDK9 have been associated with lower Huvos grade and lower survival rates, characterizing this kinase as a suitable therapeutic target, as determined through siRNA assays [461,462].

Descriptions about the relationship between CDK7 expression and EPN, RMS, EWS, WT, RB and NB are rare. Nevertheless, the use of THZ1 (CDK7 inhibitor) has exposed positive scenarios, considering that EWS cells are sensitive to this compound and that reduced EMT capacity of RB cells is observed after treatment [463,464]. In contrast, CDK9 kinase is widely expressed in RMS, where it impedes the physiological cellular differentiation [448,465–467]. Additionally, inhibition of CDK9 demonstrated a general disruption of transcription [465]. Similarly, this kinase is widely expressed in pediatric sarcomas, such as EWS, and its pharmacological inhibition (PHA-767491) enhanced the mithramycin-mediated suppression of the EWS-FLI1 transcriptional program, leading to a shift in the IC<sub>50</sub> and striking regressions of mouse xenografts. Furthermore, this kinase is upregulated in NB, increasing with the degree of differentiation of the tumor [468]. Of note, Poon et al. (2020) demonstrated that CDK9 inhibitors are able to downregulate MYCN to varying degrees and to induce apoptosis, as detected by induction of poly (ADP-ribose) polymerase (PARP) cleavage [469].

### Polo-Like Kinases

Polo-like kinases (PLKs) comprise a highly conserved multifunctional family of kinases of five members: PLK1, PLK2, PLK3, PLK4 and PLK5 [470–472]. These serine/threonine kinases are traditional controllers of cell cycle progression, with major roles in the formation of the mitotic spindle, chromatid separation, regulation of the anaphase-promoting complex, DNA damage response and cytokinesis [472,473]. Structurally, these proteins share an N-terminal highly conserved catalytic domain and a regulatory domain fundamental to the functionality and localization of PLKs, called Polo-Box (PBD) and located at the C-terminus [474].

PLKs are differentially expressed depending on the tissue and cell cycle phase [475]. Alterations in the expression of PLK genes have already been described in different types of cancer (breast, OS, leukemia, gliomas, among others) and have generally been correlated with dismal prognosis [476].

**PLK1.** PLK1 is the most studied member of the family. This protein plays key roles at different points of the cell cycle, especially during the progression of mitosis [477]. Nevertheless, other non-mitotic functions such as cell survival, genomic maintenance, cell fate and DNA damage control are also regulated by PLK1 through the interaction of with effector pathway components, including the oncogenes AKT, MYC, MDM2, B-catenin and the tumor suppressors P53, PRB, BRCA2 and PTEN [478,479].

A plethora of studies have firmly established the active role of this kinase in oncogenesis and its prognostic value along with its potentiality as a therapeutic target [480–486]. Childhood cancer is not an exception. Higher levels of PLK1 have been observed in a variety of cell lines, including EWS, OS, NB and RMS [483,484]. Moreover, this protein has been described as overexpressed in MB samples, where it is associated with higher recurrence and lower survival rates [487,488]. Furthermore, other studies have validated this positive correlation between PLK1 expression and higher cell proliferation in, mainly in undifferentiated tumors with the presence of massive choroidal invasion [205,476,489–491]. PLK1 overexpression is also present in unfavorable NB and associated with poor prognostic markers such as lower age at diagnosis and MYC amplification [492]. This interaction between PLK1 and MYC has also been observed in OS, in which the kinase contributes to MYC stabilization [493].

### Aurora Kinases

Three members of the Aurora family of serine/threonine kinases have been identified in humans: Aurora kinase A (AURKA), Aurora kinase B (AURKB) and Aurora kinase C (AURKC). These kinases (named after the resemblance or their localization to the poles of the mitotic spindle to the way aurora borealis are observed at one of the poles of the earth) have pivotal parts in the execution of mitosis (AURKA and AURKB) and meiosis (AURKC), and even exerting conserved function, they cannot fully compensate for the loss of one another [494,495].

**AURKA.** Aurora kinase A is involved in the centrosome maturation process and promotes the transition from G2 to mitosis. AURKA levels increase along late S and G2 phases and reach a higher peak in mitosis, followed by proteasome-dependent degradation [496,497]. This kinase is early localized at the centrosome and regulates the progression of mitosis by phosphorylation of multiple substrates, promoting mitotic entry through the activation of Cyclin-B/CDK1 [498]. Moreover, AURKA progressively associates with the mitotic poles and the adjacent spindle microtubules, contributing to chromosome separation and bipolar spindle [499,500].

Among the three human aurora kinases, AURKA has been the family member most consistently associated with cancer. Amplification of the chromosomal region 20q13 where the AURKA gene is located is commonly observed in cancer cells [501–503]. Nevertheless, according to Mou et al., almost 90% of tumors present in the TCGA database show AURKA overexpression [504]. Indeed, high levels of AURKA expression can endorse abnormal cell cycle progression, resulting in genomic and chromosomal instabilities, which are hallmarks



of highly proliferative tumors [505,506]. Thus, AURKA expression not only enhances proliferation, but may also influence other processes, including apoptosis evasion, EMT, drug resistance and metastasis [507–517].

Besides AURKA mitotic functions, other non-canonical and kinase-independent activities have been gradually discovered in cancer cells. After mitosis, most AURKA proteins degraded, but a remnant population may be still detected inside the nucleus, pointing to the possibility that the kinase could work as a transcriptional regulator [518]. In this regard, AURKA overexpression has been associated with the upregulation of stem cell markers such as SOX2 and NANOG, imposing participation in the maintenance of the self-renewal capacity of cancer stem cells (CSCs) [519].

In pediatric tumors, AURKA polymorphism rs8173 G > C has shown to decrease WT risk [520]. Conversely, AURKA plays an active role protecting MYCN from ubiquitinylation and proteolysis in NB, thus contributing to more aggressive phenotypes and poor survival probability [521,522]. Moreover, this role has also been described in RMS, where AURKA not only stabilizes MYC but also PAX3-FOXO1 [523]. Moreover, in this tumor type, AURK is overexpressed and considered a key factor in the observed aneuploidy and chromosomal instability [524]. AURKA is also closely related to the oncogenic process of EWS and is considered a chemotherapy resistance and may act as a potential biomarker for prognosis [525,526].

AURKA overexpression has also been associated with OS, as many cell lines are highly sensitive to its inhibition [527,528]. Similar patterns have been seen for WT, where AURKA inhibition impaired tumor growth and induced apoptosis both in vitro and in vivo; however, such effects were improved in RB1-deficient cell lines compared to those with MYC amplification [529].

Moreover, expression profiling of pediatric brain tumors has shown that AURKA was consistently and highly overexpressed (up to 106-fold) in tumor samples from all glioma grades and from patients varying from 4 months to 82 years old; however, mRNA expression showed only weak correlation with the Ki-67 labeling index, and significant associations with poor patient survival were only observed for GBM [530]. Overexpression of AURKA is also linked with survival in MB patients [531].

**AURKB.** The second member of the family, Aurora kinase B (AURKB) is one of the most intensively studied kinases because it provides catalytic activity to the chromosome passenger complex (CPC), formed by AURKB, INCEP (inner centromere protein), survivin and borealin (Cell 2002;13:3064–77). The CPC governs highly different processes, such as chromosome alignment, histone modification and cytokinesis [532–535]. Additionally, AURKB kinase activity is essential for faithful chromosome segregation and functions to correct any improper kinetochore attachment to the spindle [532,536–538]. Finally, Aurora kinase B (AURKB) is also essential in mitotic DNA damage response, protecting against DNA damage-induced chromosome segregation errors, including the control of abscission checkpoint and prevention of micronuclei formation [539]. Consequently, AURKB dysregulation results in aneuploidy and genomic instability and in turn promotes cell cycle progression and survival of cancer cells [540–543]. Indeed, expanding evidence at the gene, mRNA and protein levels supports a carcinogenic role of AURKB. Overexpression has been reported in clear cell renal cell carcinoma (RCC) and cervical carcinoma, among many others, with clear associations with clinicopathological parameters such as stage and tumor volume, chemoresistance, tumor progression and poorer survival [544–553].

In the pediatric setting, overexpression of AURKB was closely correlated with poor prognosis and carboplatin resistance in NB patients [552,554]. Similar profiles were observed for pediatric ALL and AML patients, especially in T-cell and E2A-PBX1-translocated ALL cases. Further in vitro assays demonstrated that AURKB is an essential protein for the proliferation and survival of acute leukemia cells [555].

The importance of Aurora kinases as potential therapeutic targets for childhood brain malignancies is highlighted by AURKB being highly and consistently overexpressed in the majority of high-grade gliomas, but despite reflecting the presence of aneuploidy,

at least in EPN, it did not emerge as a prognostic factor [556,557]. For other tumor types, however, data about the prognostic value of AURKB are less explored and primarily rely on experimental assays using pharmacological inhibitors. In this context, OS, EWS and RB are included [558–560].

**AURKC.** Differentially from AURKA and AURKB, Aurora kinase C (AURKC) is limited to cells that undergo meiosis (sperm and oocyte). This kinase is located on human chromosome 19q13.43, regulated by promoter methylation and when expressed in germ cells, can undergo alternative splicing resulting in three protein variants [561–563]. As the major enzymatic component of the CPC during meiosis, it plays a specific role during human female meiosis and preimplantation embryo development [564].

A body of evidence shows that overexpression of AURKC in mitotic cells leads to centrosome amplification and multinucleation [565]. Its upregulation and other CPC components occur in cancer cells and may correlate with clinical characteristics [566,567]. In line with this, AURKC is overexpressed in tumors of the reproductive system and in breast and prostate cancer cell lines [568,569]. Nevertheless, varying degrees of CpG islands hypermethylation leading to lower AURKC mRNA levels have been described in WT, suggesting that this kinase might not be of importance in this childhood tumor [570]. Likewise, AURKC expression was not associated with survival or risk status in neuroblastoma patients [571]. In OS, knockout of AURKC displayed no changes in cell proliferation, migrated less and formed fewer colonies in soft agar compared to wild-type cells. Moreover, whole-transcriptome sequencing revealed over 400 differentially expressed genes which included genes encoding proteinaceous extracellular matrix components, suggesting that therapeutics targeting this aurora kinase isoform could decrease cancer cell metastasis and disease progression, the most limiting characteristic of survival [572].

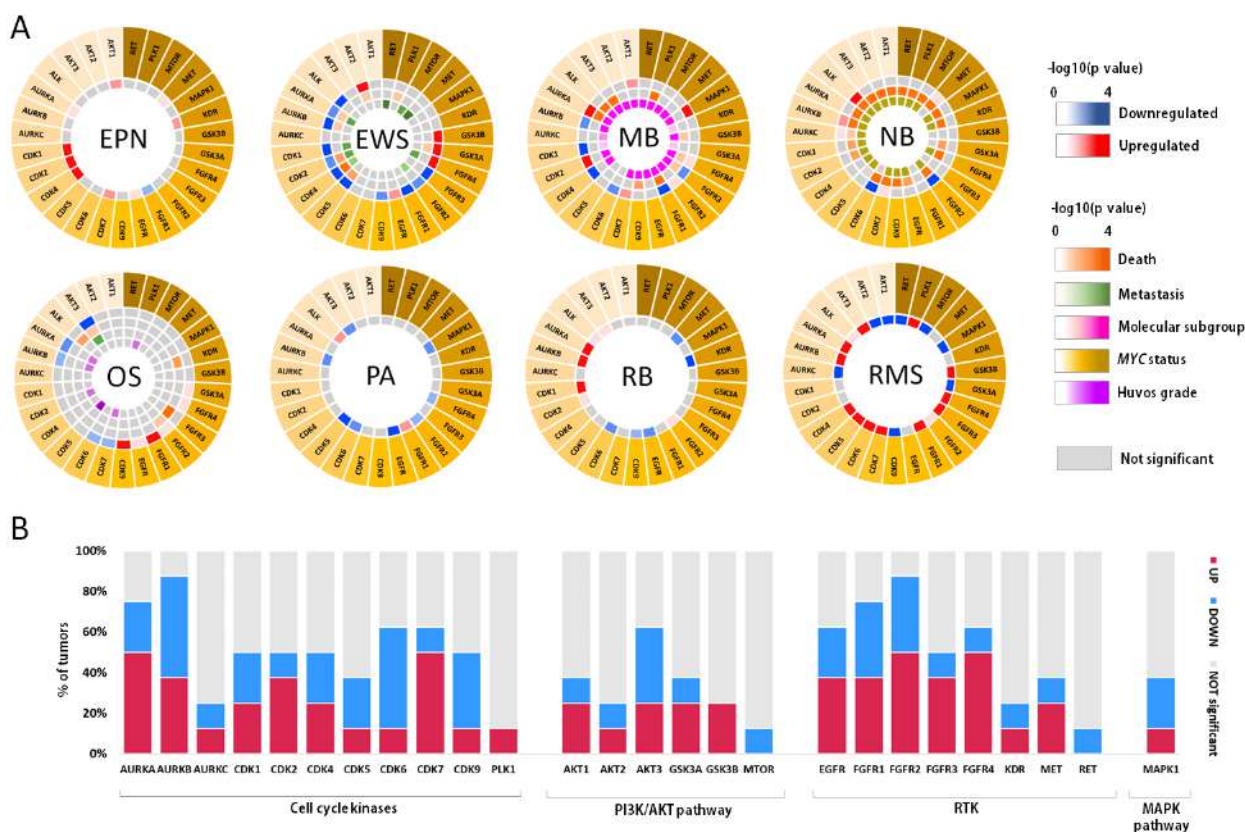
### 3.2. *In Silico Analysis of Different Kinases Expression and Their Association with Clinical Prognosis*

According to our systematic *in silico* analysis of the selected group of kinases, comparisons of expression patterns between pediatric tumors and normal samples showed varying results, and despite what was expected from the data already published, few commonalities were found (Figure 3A,B, Supplementary Table S1). Considering CNS tumors, overexpression of *AKT1*, *AURKA*, *CDK2* and *CDK7* was observed in EPN and MB, but not in PA. Similarly, EPN showed higher levels of *CDK1*, *CDK4*, *EGFR*, *KDR* and *MET*, while for MB, the upregulation of *FGFR2*, *FGFR4* and *MAPK1* was highlighted. Conversely, EPN samples demonstrated low levels *AKT3* and *FGFR1*, while *CDK1/4/6*, *FGFR1/3* and *AURKB* were downregulated in MB tissues. PA, on the other hand, exhibited high levels of only *ALK* and *FGFR1*, and downregulated genes included *AKT3*, *AURKB*, *CDK5*, *EGFR*, *FGFR2*, *FGFR4* and *MAPK1*.

Neuroblastoma was the tumor type with the more reduced number of hub genes. In this tumor type, *CDK6* and *FGFR2* were less expressed than in normal tissue, whereas *CDK7*, *MET*, *ALK* and *AURKB* stood out as upregulated in tumor samples. Concomitantly, RB showed high levels of *AKT3*, *ALK*, *AURKA*, *AURKB*, *CDK1*, *CDK2* and *FGFR2* genes and low levels of *CDK6*, *CDK9*, *EGFR*, *KDR* and *MET*.

Among sarcomas, RMS was the tumor type with the most altered kinase profile, including high levels of *AKT3*, *ALK*, *AURKA*, *AURKB*, *CDK4*, *CDK5*, *CDK6*, *CDK7*, *GSK3B*, *PLK1* and all *FGFR*. Then, *AKT2*, *EGFR*, *FGFR4* and *GSK3s* (A and B) showed higher expression in EWS, contrasting the low levels of *ALK*, *AURKs* (A and B), *CDKs* (1, 2, 4, 5 and 9) and *FGFRs* (1, 2 and 3). Finally, our analysis of OS samples demonstrated upregulation of the receptor genes *EGFR* and *FGFR* (1, 2 and 4), as well as *CDK9* and *GSK3A*. Alternatively, *CDK6/7*, *AURKA/B* and *AKT3* had low expression profiles.

Notwithstanding, further analysis showed that, in the same line as reported in the literature, the expression of most of the selected kinases is indeed associated with clinical features of worse prognosis, including associations with *MYC* amplification in NB and molecular subtypes in MB, and metastases in bone sarcomas (Supplementary Table S2).



**Figure 3.** (A) Polar plots of differentially expressed kinases in pediatric tumors obtained through the analysis of available data on the R2: Genomics Analysis and Visualization Platform (<http://r2.amc.nl> (accessed on 15 October 2022)). Tumor abbreviations: EPN—ependymoma; EWS—Ewing sarcoma; MB—medulloblastoma; NB—neuroblastoma; OS—osteosarcoma; PA—pilocytic astrocytoma; RB—retinoblastoma; RMS—rhabdomyosarcoma.  $p$ -values are represented by differential coloring gradients. The external inner circle corresponds with “tumor versus normal tissue” results. The other concentric layers represent data related to associations with clinical features: metastasis, death, molecular subgroup (MB), MYC status (NB) and Huvos grade (OS). For actual  $p$ -values, refer to Supplementary Table S2. (B) Percentage of tumors with altered expression of each kinase. Few commonalities were found.

#### 4. Kinases as Druggable Targets—Evidence and Limitations

The gradual advancements in genetics and biochemistry during the second half of the last century not only contributed to the better understanding of molecular events underneath signaling pathways in both natural and pathological settings, but also laid the foundation for the development of modern targeted agents. Perhaps the most expressive example of that trajectory involves chronic myeloid leukemia and the “Philadelphia chromosome”. After its simple description (250 words) by Nowell and Hungerford, it took a decade to properly identify the chromosome pairs involved in the translocation [573]. It was only after the introduction of the G-bands by Marina Seabright that Janet D. Rowley from the University of Chicago that it was possible to identify the little chromosome as a result of the reciprocal translocation between chromosomes 9 and 22, specifically, t(9;22)(q34;q11) [574,575]. However, its molecular characterization only came to light between 1982 and 1984 [576–578], demonstrating the in-frame juxtaposition of the *ABL* oncogene (on chromosome 9) with the *BCR* gene (on chromosome 22), resulting in the hybrid *BCR/ABL* gene that gives rise to a chimeric protein with high tyrosine kinase activity and with a critical role in the development of leukemia [579]. Later, the discovery of this tumor-specific protein led to development of imatinib mesylate, providing an incredibly

successful treatment that converted a fatal cancer into a manageable chronic condition, and pioneered an era of target-directed therapy [580].

More recently, the emergence of integrative laboratorial methods such as kinome-wide siRNA screens, next-generation sequencing (NGS) and phosphoproteomics have dramatically intensified the assortment of kinase inhibitors for the treatment of human cancers, currently accounting for about a quarter of all drug discovery research and development efforts.

Moreover, the increasing number of databases and analytical and visualization tools has facilitated advanced drug discovery not only by gathering information about the prognostic value of specific genes in oncology, but also it is now possible to access chemical structures and docking, affinities and structural features of approved small-molecule inhibitors in more easily, accessible and systematic ways, thus accelerating the discovery and optimizing screening to more direct translational assays.

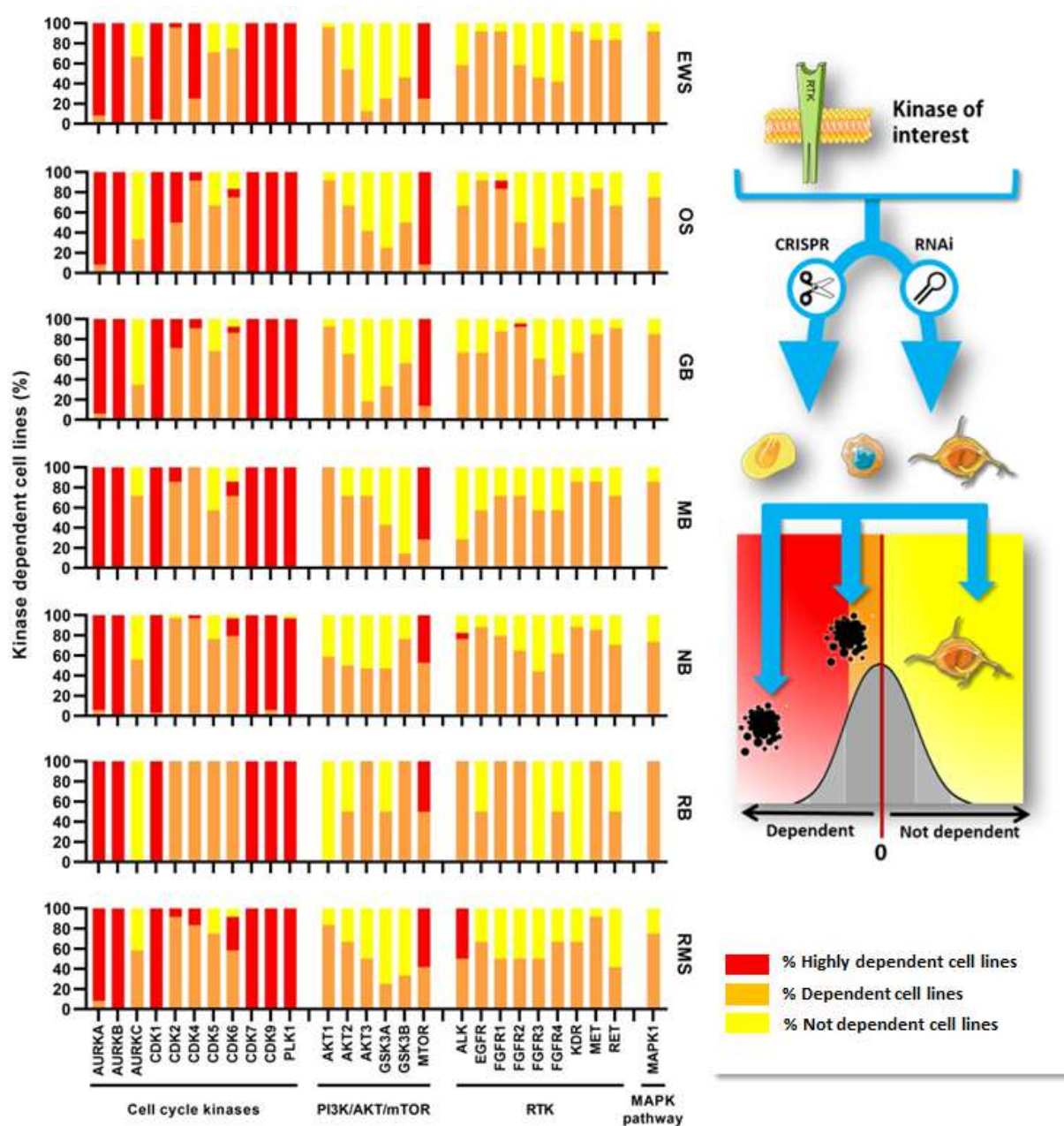
In this regard, to further illustrate the importance of the selected kinases' dysregulation in the pathophysiology of pediatric cancer, other bioinformatic tools were used (Supplementary Figure S2). As a first step, we analyzed the vulnerability of different cancer cell lines against their inhibition through the Cancer Dependency Map portal (<http://depmap.org> accessed on 14 December 2022), a platform that provides information about how dependent different cell lines are on a specific gene depletion based on CRISPR and RNAi knockout experiments. The results are presented as a score generated by the platform itself: greater than zero ( $>0$ ) indicates that the cell line is not dependent, less than zero ( $<0$ ) indicates that the lineage is dependent and scores below  $-1$  indicate that the analyzed gene is essential for the survival of the cell lineage.

Initial screening showed that all or most cell lines are dependent on the kinases analyzed, with comparable scores between adult or pediatric origins (Figure 4; Supplementary Table S3). Interestingly, a similar pattern occurs across the different tumors, irrespective of histology. Almost 100% of the cell lines are highly dependent on cell cycle kinases, especially AURKA/B AURKB, cyclin-dependent kinases CDK1 and CDK7/9, and PLK1. Cell lines were also highly dependent on mTOR. Conversely, cell lines were less vulnerable to the depletion of AURKC, AKT3, GSK3A and FGFR3, with more than 50% of cell lines presenting scores above 1 (in line with published data reviewed above).

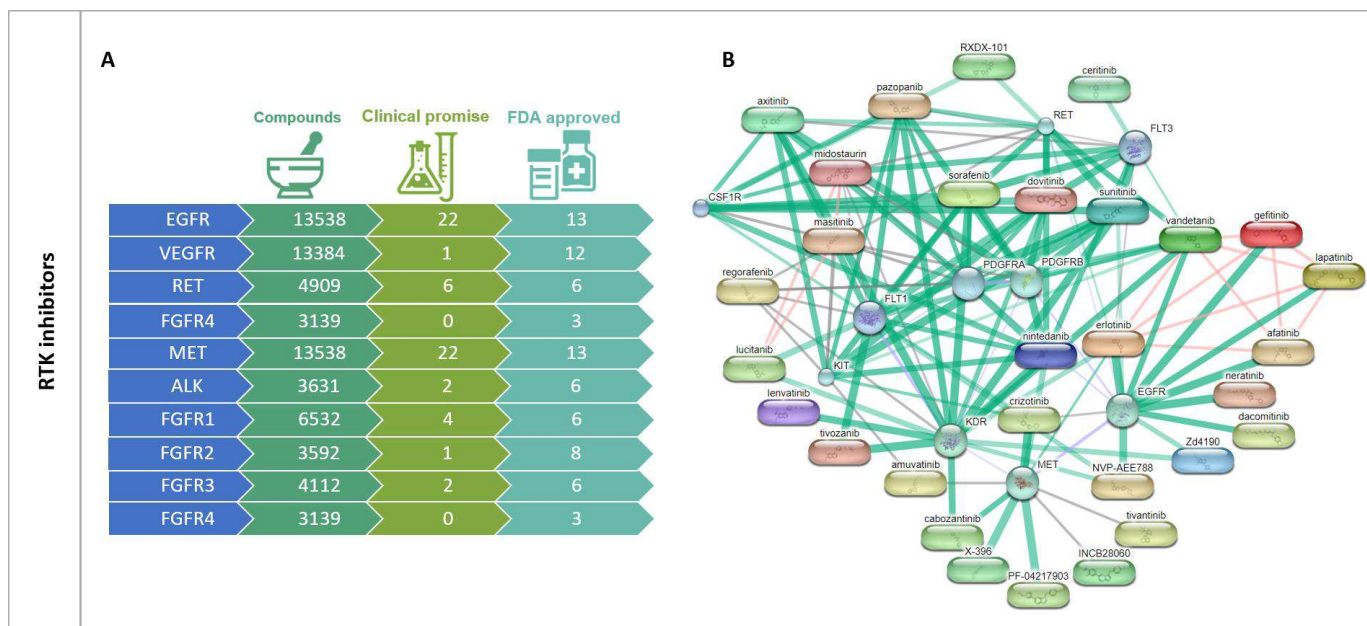
Then, aiming to further exemplify information on individual kinase-targeted compounds and their analogs, we performed a search through the CanSAR knowledgebase (<http://cansar.icr.ac.uk> accessed on 14 December 2022), an integrative platform that compiles multidisciplinary data and provides useful drug discovery predictions. The analysis of predicted compounds for our selected group of kinases showed more than 55,000 potential compounds that are able to target RTK, with EGFR and VEGFR representing the most druggable ones. As shown in Figure 5, FDA-approved drugs are already identified for all the RTKs, and more than 20 additional drugs are being studied as novel clinical candidates.

For PI3K, although the CanSAR analysis revealed more than 20,000 potential compounds, only the mTOR inhibitors perhexiline and everolimus are described as FDA-approved drugs. Nonetheless, 15, 4 and 2 clinical candidates are being investigated as specific inhibitors for mTOR, AKT1 and GSK3B, respectively (Supplementary Figure S3). Approved drugs for MAPK are also scarce, including only sorafenib, regorafenib, dabrafenib and encorafenib, all of which target RAF1. For this specific kinase, CanSAR identified 4764 promising compounds and three clinical candidates (Supplementary Figure S3). Among the cell cycle kinases, CanSAR identified only approved drugs targeting CDK4 and CDK6. However, many clinical candidates (more than 15) targeting the other kinases of this group are being studied. According to the platform, cell cycle kinases are the second druggable category of kinases with the highest number of compounds described as potential specific drugs, totaling more than 48,000 (Supplementary Figure S3).





**Figure 4.** Percentage of pediatric cell lines dependent on the selected group of kinases. Dependency data were imported from the DepMap consortium (CRISPR (DepMap 22Q2 Public + Score, Chronos; <https://depmap.org/> portal/ (accessed on 30 October 2022)) and classified as highly dependent, dependent or not dependent. The data were plotted on a histogram where it is possible to see the vulnerability of pediatric cell lines mainly to cell cycle kinases and PI3K/AKT/mTOR families. Cell lines selected included Ewing sarcoma (EWS), osteosarcoma (OS), glioma (GB), medulloblastoma (MB), neuroblastoma (NB), neuroblastoma (NB), retinoblastoma (RB) and rhabdomyosarcoma (RMS). Dependency scores for each cell line are shown in Supplementary Table S3.



**Figure 5.** (A) Schematic illustrations of RTK druggability identified by the CanSAR database, including the total number of compounds with predicted interaction capacity with each kinase, as well as FDA-approved drugs and clinical candidates. (B) Interaction networks of RTK inhibitors and associated binding proteins according to STITCH (search tool for known and predicted interactions between chemicals and proteins available at <http://stitch.embl.de> (accessed on 1 November 2022)). Compounds are represented as pill-shaped nodes, while proteins are shown as spheres. Small nodes represent proteins of unknown 3D structures, while large nodes show proteins with known or predicted structures. Nodes that are associated with each other are linked by an edge: thicker lines represent stronger binding affinities. Networks were constructed considering a minimum required interaction score of 0.700, and based on associations reported in curated databases (gray lines), or on both databases and experimental/biochemical data (green lines). Purple lines represent functional links between proteins.

Further, for compilation of the preclinical results on kinase inhibitors, and considering that information in the literature often appears scattered and fragmented, the following section shows published evidence for (1) solid tumors that showed differential gene expression through in silico analysis (refer to Figure 3), (2) compounds considered “FDA approved” or with “clinical promise” and (3) compounds that have been assessed in vitro or in vivo before entering clinical trials. Thus, in the following subsections, experimental data on individual kinase inhibitors are detailed within each category. For further information, see Supplementary Table S4.

#### 4.1. RTK Inhibitors

**Erlotinib (Tarceva®).** This compound is a quinazoline derivative that selectively and reversibly inhibits EGFR [581]. Erlotinib is an FDA-approved drug for the treatment of NSCLC and pancreatic cancer in combination with gemcitabine chemotherapy [582]. In the pediatric setting, this inhibitor has shown contrasting results. As monotherapy, Erlotinib was not efficient in reducing cell growth in a panel of NB cell lines, albeit effective indirect responses were obtained in xenograft tumors [583]. Similar results were obtained in sarcomas. This compound alone was ineffective in OS cells, in which the STAT3 cascade pathway has been pointed as the molecular mediator of both intrinsic and acquired resistance. In EWS, this compound alone or in combination did not inhibit growth of tumor xenografts and even led to a decrease in the therapeutic activity of cyclophosphamide when compared to single-agent activity [584,585]. Erlotinib had no effect on tumor progression in genetically engineered ARMS mouse models [586].

In contrast, satisfactory results were obtained in CNS tumors, where erlotinib therapy inhibited MB migration *in vitro* and successfully diminished the levels of phosphorylated EGFR in EPN models [587,588]. Treatment with this drug was also cytotoxic in Y79 and WERI RB cells in a dose-dependent manner, leading to cell cycle arrest and reduced migration, while oral administration dramatically reduced the growth of Y79 tumor grafts [589]. Regarding patients, phase I clinical studies have been developed in order to evaluate the acceptable tolerability profile in cases of brain and refractory solid tumors including RMS, soft tissue sarcomas, NB or germ cell tumor. Children appeared to tolerate erlotinib similarly to adult patients, and drug disposition was similar between these populations. The combination of temozolomide and erlotinib was well tolerated and it was also suggested in combination with radiotherapy [590,591].

**Vandetanib (Caprelsa®).** This is a multitargeted tyrosine kinase inhibitor with potent effects against VEGFR2/3, EGFR and RET [592,593]. This compound is approved to treat medullary thyroid cancer that cannot be removed by surgery and is locally advanced or has metastasized, and has demonstrated modest efficacy in patients with metastatic breast cancer [594,595]. Regarding pediatric tumors, vandetanib has been shown to inhibit the proliferation of NB cells mediated by the induction of G1-phase cell cycle arrest at lower concentrations and by apoptosis at higher concentrations. Migration and invasion were also markedly decreased compared with the control group [596,597]. Treatment also decreased (p)RET expression in five other NB cell lines and strongly impaired tumor growth *in vivo* in both MYCN/KI AlkR1279Q and MYCN/KI AlkF1178L mice, and was able to sensitize cisplatin-resistant NB subcutaneous tumor growth with less severe liver toxicity compared with high-dose cisplatin [598–600]. Moreover, vandetanib, in combination with 13-cis-retinoic acid, reduced tumor vascularity and induced apoptosis in NB xenografts [601]. Indeed, according to Craveiro et al. (2017), the narrow target spectrum of Vandetanib along with a favorable toxicity profile makes this drug ideal for multimodal treatment approaches. These authors tested this compound against SHH-TP53-mutated and MYC-amplified MB cell lines and found that it leads to a dose-dependent reduction in cell viability, interferes with clonogenicity and has pro-apoptotic effects after 48 h. Of note, combinations with GDC-0941 (clinically available PI3K inhibitor) and etoposide resulted in complete loss of cell viability [602]. The combination of vandetanib and celecoxib displayed a synergistic or additive antitumor effect on OS *in vitro* and *in vivo* [603]. However, combinations of gefitinib and vandetanib only inhibited the proliferation of EWS cell lines at very high concentrations (>1  $\mu$ M vandetanib, >5  $\mu$ M gefitinib), indicating the action on off-target effects [604].

**Gefitinib (Iressa®).** This drug, also known as ZD1839, is a member of the 4-anilinoquinazoline class of compounds that specifically and selectively inhibits EGFR [605]. In preclinical studies, gefitinib treatment was associated with growth inhibition and increased apoptosis in human cancer cell lines, and antitumor effects against xenografts of human tumors [606,607]. Gefitinib was shown to inhibit proliferation in juvenile PA primary cell cultures with an IC<sub>50</sub> determined between 1.6 and 9.6  $\mu$ M [608]. In addition, this compound was shown to inhibit invasion and metastasis of intratibial OS xenografts via inhibition of macrophage receptor interacting serine-threonine kinase 2 (RIPK2) [609]. Moreover, in children, Gefitinib has been tested for refractory solid tumors and CNS malignancies, showing similar pharmacokinetics as in adults [610].

**Regorafenib (Stivarga®).** This drug is an oral multikinase inhibitor that targets VEGFR1/3, FGFR and other receptor kinases [611]. This compound is already approved to treat metastatic cases of colorectal cancer and advanced hepatocellular carcinoma (HCC) previously treated with Sorafenib [610]. *In vitro*, regorafenib exhibits antiproliferative effects against a panel of 33 pediatric tumor cell lines, including MB (D341 Med, Med-Meb-8A), OS (IOR-OS-18), EWS (EW7, ORS, POE, SIM, STA-ET-1) NB and (SJ-NB-8, SK-N-BE(2), SH-SY5Y), with a mean half maximal growth inhibition of 12.5  $\mu$ mol/L [612]. Particularly in the last, Regorafenib has shown to be effective against through the inhibition of RAS/MAPK, PI3K/Akt/mTOR and Fos/Jun pathways [612]. Similarly, regorafenib



demonstrated antitumor activity in animals bearing subcutaneous RMS, EWS (STA-ET-1 and EW7) and NB (SJ-N-B8 and SK-N-AS) xenografts with tumor growth inhibition ranging from 73% to 93%. Moreover, when associated with radiation and irinotecan, it induced 100% regression in an MB patient-derived xenografts (PDX) model [610].

**Dacomitinib (Vizimpro®).** Also known as PF-00299804, this drug is an orally administered, second-generation, irreversible inhibitor of EGFR, HER2 and HER4, which has shown positive anticancer activities in some preclinical and clinical trials, being approved by the FDA for the treatment of metastatic NSCLC [613,614]. Besides its ATP-competitive action, dacomitinib covalently binds to Cys773 located in the ATP-binding cleft of EGFR, which irreversibly blocks ATP binding and inactivates the receptor [615]. For pediatric MB, dacomitinib has shown to block EGFR/HER signaling in DAOY cells and in orthotopic xenografts, extending median survival as a single agent; however, it was antagonistic when used in combination with standard frontline chemotherapy (4HPC, vincristine or cisplatin) [616].

**Lapatinib (Tykerb®).** This compound is an oral dual tyrosine kinase inhibitor that inhibits human EGFR and blocks the EGF receptor 2 (HER2) [617]. It was FDA-approved to treat HER2-positive advanced or metastatic breast cancer as monotherapy or in combination with other drugs [618]. Lapatinib has been tested in childhood solid tumors (including RMS, EWS and NB) and leukemia cells by the NCI-supported Pediatric Preclinical Testing Program (PPTP) [619]. In this study, among 23 cell lines, fifteen achieved at least 50% growth inhibition, and the median IC<sub>50</sub> value for lapatinib against the entire cell line panel was 6.84 μM (range 2.08 μM to >10.0 μM). In vivo, however, lapatinib presented little activity against the 41 xenograft models of pediatric tumors [619,620].

**Cetuximab (Erbix®).** This compound, available as Erbitux® (Merck Sereno), is a human–murine chimeric monoclonal antibody that competes to bind to the extracellular domain of EGFR and has been approved for the treatment of colorectal and head and neck cancer [620]. Information about preclinical use of this compound is scarce. However, a report showed that the proliferation of RMS cell lines was not influenced by this EGFR inhibitor [621]. However, later, it was shown that the combination of cetuximab and actinomycin D was highly effective in EGFR-positive RMS cells (RD and Rh30, of embryonal and alveolar origin, respectively), synergistically inhibiting cell growth and inducing apoptosis [622].

**Sunitinib:** Sold under the brand name Sutent®, this drug is a small-molecule multitarget inhibitor functioning on PDGFR, VEGFR, KIT, Flt-3 and RET [623–625]. In a preclinical study, this drug demonstrated limited growth inhibitory effects in a panel of 23 pediatric cell lines that included OS, ALL, EWS, RMS, MB, EPN, NR, GBM, WT and others [625]. However, in vivo, it presented growth inhibitory activity against pediatric solid xenograft models of EWS, RMS and NB [625]. A later study showed decreased cell proliferation and phosphorylation of VEGFRs NB cells after treatment with sunitinib, and tumor growth, angiogenesis and metastasis in tumor xenograft models [624]. Moreover, in combination with an mTOR inhibitor (rapamycin), it showed a synergic cytotoxic effect, which was more effective than the traditional chemotherapeutic agent cyclophosphamide [624].

**Lenvatinib (Lenvima®).** This drug is a synthetic, orally available type I tyrosine kinase inhibitor exhibiting powerful antiangiogenic activity currently used to treat certain types of thyroid cancer and potentially other tumor types [626]. Lenvatinib was initially reported in 2008 as a multitargeted RTK inhibitor of VEGFR1/2/3, but it also inhibits FGFR1–4, PDGFR-α and KIT [627–629]. Preclinical findings in sarcomas indicated that lenvatinib was able to inhibit tumor growth in xenografts obtained through direct implantation of patient tumor specimens in nude mice. The experiment showed positive results in 7 out of 10 xenografts accompanied by marked decrease in microvessel densities. However, in vitro, Lenvatinib did not show potent effects on tumor viability in OS-derived cell lines [629]. Others showed that the drug was able to inhibit tumor cell migration and invasion in U2OS cells [630]. Further, in a phase I/II study, lenvatinib as a single-agent reported a response rate of 7% and a median progression-free survival of 3 months in a



cohort of 31 children and young adults with OS, although many patients had treatment-related adverse events of grade  $\geq 3$  [631]. In other pediatric tumors, the effects of lenvatinib remain to be investigated.

**Pazopanib (Votrient®).** This compound is an FDA-approved pan-VGFR inhibitor, even though it also targets PDGFR- $\alpha$  and - $\beta$ , FGFR1/3, KIT as well as BRAF proteins [632]. In a pan-cancer study, pazopanib was unable to affect the viability of any of the treated cell lines, which included SK-N-BE(2) (N-Myc amplified) and SH-SY5Y (non-N-Myc amplified) NB cell lines, the KHOS OS cell line, and the RMS cell lines RH30 and RD. However, in combination with topotecan, this compound showed significant antitumor activity in vitro and halted tumor growth in NB xenograft-bearing mice, but after 50 days, gradual growth was observed [633,634]. The combination of pazopanib with trametinib showed antitumor effects in vitro and in vivo against a panel of seven OS cell lines, in which treatment reduced proliferation and colony-forming capacity and increased the percentage of apoptotic and dead cells. In MNNG/HOS and KHOS xenograft models, both drugs induced a significant inhibition of tumor growth compared to the untreated controls [635]. The in vivo antitumor activity of pazopanib was also tested by the PPTP Program in a subset of sarcoma models that also included EWS and RMS. Although objective responses were not observed for any of the sarcoma xenografts studied, treatment prolonged survival [636]. Even with modest benefits, pazopanib has been approved for line treatment of metastatic non-adipocytic soft tissue sarcomas after the failure of standard chemotherapy. Its efficacy in patients with OS is limited to case reports [637]. One metastatic extraosseous EWS was also reported as successful after treatment with pazopanib [638].

Regarding CNS tumors, EPN cells showed to be sensitive to Pazopanib with a viability reduction of around 35% at 1  $\mu\text{mol/L}$  [639]. Additionally, treatment with Pazopanib reduced the mobility of MB cell lines, inducing clumping of the actin microfilaments (which facilitated cell detachment), as detected by wound healing assays and Fluor-555-coupled phalloidin [640]. Further in vivo tests demonstrated delayed growth of group-3-MB cells transplanted into the cerebellum of mice and prolonged survival (by 10 days) of mice treated once daily by gavage with 60 mg/kg compared to untreated controls [641]. Alternatively, for patients with recurrent high-grade gliomas as part of phase I or II clinical trials, this drug has not been beneficial [642].

**Cabozantinib (Cometriq®).** This compound is an orally available multitarget tyrosine kinase inhibitor that inhibits VEGFR1/2/3, MET, KIT, RET, AXL and FLT3. FDA-approved since 2012, it is currently used to treat metastatic medullary thyroid cancer, RCC, HCC and differentiated thyroid cancer [643,644]. In preclinical studies, reports of its anticancer effects include the inhibition of metastasis, angiogenesis and tumor growth [645–647]. In vitro, cabozantinib has been shown to diminish the cell viability of EWS and OS cells in a dose-dependent manner [160]. Moreover, it also interferes with migration and the microenvironment by inducing the production of osteoprotegerin and causing a decrease in the synthesis of the RANK ligand by osteoblasts [648]. Positive effects on decreasing proliferation were also observed in MB with no differences between cell lines corresponding to different molecular subgroups [649]. Cabozantinib also exhibited anti-proliferative effects in NB cells and reduced cell migration in vitro and significantly inhibited tumor growth of orthotopic xenografts on a daily basis [650].

**Nintedanib (Ofev®).** This drug, commercially available under the brand names Ofev and Vargatefi, is an indolinone-derived inhibitor of multiple kinases including VEGFR, FGFR and PDGFR. Recently approved for the treatment of idiopathic pulmonary fibrosis and advanced non-small cell cancer of adenocarcinoma tumor histology, it exerts its antitumor activity by reducing proliferation, migration and angiogenesis [651–653]. Considering pediatric tumors, nintedanib has been shown to inhibit growth in EWS (A673, CHP100) and OS (SaOS2) cell lines, with a key role in controlling OS lung metastatic growth by blocking the fibrogenic reprogramming of OS stem cells (OSCs) [654,655]. Growth inhibition was also observed in a panel of 13 RMS cells, with the PAX3–FOXO1 fusion-gene-positive ones more sensitive to treatment [656]. Moreover, there are reports of EPN cells being sensitive

to nintedanib treatment, while this drug is able to extend the survival of mice bearing ST-RELA xenografts [657,658].

**Midostaurin** (Rydapt/Tauritmo<sup>®</sup>). Also known as PKC412 and CGP 41251, this small molecule acts as a multikinase inhibitor targeting PKC $\alpha/\beta/\gamma$ , Syk, Flk-1, Akt, PKA, c-Kit, c-Fgr, c-Src, FLT3, PDGFR $\beta$  and VEGFR1/2. Presenting anticancer roles in vitro and in vivo, it is currently approved for the treatment of acute myeloid leukemia and advanced systemic mastocytosis [659,660]. Midostaurin has been shown to be an efficient anti-sarcoma agent. Indeed, it inhibited EWS cell proliferation in a dose- and time-dependent manner and decreased tumor growth in vivo [661,662]. Moreover, the combination of midostaurin with the cytokine oncostatin M has been shown to be efficient in reducing in vivo tumors, pointing to this combination as a potential adjuvant treatment for OS [663].

**Axitinib (Inlyta<sup>®</sup>)**. Also known as AG-013736 this is an oral VEGFR1/3 and PDGFR inhibitor explored to control angiogenesis [664]. Currently, this compound is approved for treatment as monotherapy or in combination with other drugs for renal carcinoma, and is under phase I, II and III clinical trials for many other tumor types [664]. Pre-clinical studies in EPN showed that this drug inhibited PDGFR $\alpha$  and PDGFR $\beta$ , and reduced the expression of mitosis-related genes including *ASF1B*, *MKI67*, *HMGA1*, *BRCA2*, *ESPL1*, *TACC3*, *CDC25A*, *RAD51AP1*, *AURKA*, *BUB1B*, *CENPE* and *HELLS*. It also decreased proliferation resulting from cellular senescence [639]. Similar antiproliferative effects were observed in MB 2D and 3D cell cultures, without affecting normal brain cells. Of note, the compound efficiently crossed the blood–brain barrier (BBB), reducing growth rates of experimental brain tumors without acute toxicity in juvenile rats [649]. In GBM, the cytotoxic activity of Axitinib was also reported in vitro and in vivo, characterized by an anti-angiogenic effect and survival prolongation [665]. Moreover, combinations of axitinib and other therapeutic targets have been explored with satisfactory results [666]. Indeed, the combinatorial treatment of Axitinib and PLK4 inhibitor has shown to be beneficial in MB and RMS [667]. Combinations with etoposide or gemcitabine also showed favorable effects on preventing tumor progression in an orthotopic group-3-MB xenograft models [649,668]. Furthermore, in immunodeficient and immunocompetent orthotopic GBM models, axitinib + G47 $\Delta$ -mIL12 resulted in an extensive decrease in vascularity, increased macrophage infiltration and significant tumor necrosis [669]. Such a antimetastatic effect was also observed in NB [670].

**Ramucirumab (Cyramza<sup>®</sup>)**. This is a humanized monoclonal antibody that acts by binding to VEGFR-2, thus limiting angiogenesis and the proliferation and migration of human endothelial cells [671]. Preclinical studies in NB, RB, OS, RMS and EWS have also shown that ramucirumab enhances anti-tumor activity by abrogating endothelial cord formation, while in vivo, it has also induced tumor growth delay. However, modest or no response was observed in OS [672]. This compound was approved by the FDA in 2014 and indicated for the treatment of gastric cancer, NSCLC, colorectal cancer and HCC [673–675]. As a well-tolerated drug, its combinatorial use was also approved, even though its use in clinics is limited due to a lack of specific markers and high costs [676].

**Alectinib (Alecensa<sup>®</sup>)**. Also known as CH5424802, this is an orally available selective ALK inhibitor already approved by the FDA for lung cancer treatment [677]. The compound is able to bind wild-type ALK and its fusions and its anticancer effects have been widely described. Noteworthy, it has shown acceptable results after treatment of intracranial EML4-ALK-positive tumors in rats with a high brain-to-plasma ratio, and permeability independent of P-glycoprotein transport [678]. Moreover, despite heterogeneous intratumoral distribution, alectinib delayed tumor growth in an NB mouse model, leading to increased survival, providing an option for future clinical treatment [679–681]. An interesting point in this regard is that Alectinib may improve sensitivity to chemotherapeutic since it increases the intracellular accumulation of ABCB1/ABCG2 substrates such as doxorubicin (DOX) and rhodamine [682]. Moreover, it has also shown effectiveness in combination with the histone deacetylase inhibitor vorinostat in NB harboring the ALK R1275Q mutation and

after intensive radiotherapy for the treatment of a rare intraosseous RMS with FUS-TFCP2 fusion, evidencing the potential of this drug to treat extremely aggressive tumors [683,684].

**AEE-788.** This drug is an orally bioavailable bispecific EGFR/HER2 inhibitor that exerts significant anti-tumoral activities and radio-sensitizes EGFR-overexpressing cells [685]. By targeting this receptor, the compound efficiently reduced clonogenicity, proliferation and survival of EPN cells and prolonged the survival of tumor-bearing mice, probably due to the increase in apoptosis of endothelial cells (as shown by others in cutaneous cancer xenografts) [686,687]. AEE788 also inhibited cell proliferation and prevented epidermal growth factor- and neuregulin-induced HER1, HER2 and HER3 activation in chemosensitive and chemoresistant (cisplatin selected) MB cells in vitro and in vivo [688].

**Crizotinib (Xalkori®).** This drug is an orally available aminopyridine-based ATP-competitive inhibitor of ALK that has shown positive results against NSCLC [689]. In turn, in pediatric tumors, growth-suppressive activities have been reported in some tumor types, such as PA, EPN, EWS and MB [160,690,691]. This drug was also able to induce apoptosis and autophagy in a dose-dependent manner in RMS cells, reducing cell migration and invasion, as well [692]. However, despite these promising results, this compound lacks clinical significance in patients with FOXO1-rearranged ARMS [693]. Similarly, crizotinib responses in NB are variable and mostly dependent on the mutation variants present in the tumor, considerably limiting its applicability [694–696]. Moreover, the literature widely illustrates that despite initial effectiveness, the vast majority of tumors treated with this compound will develop resistance within a few years [697].

**Capmatinib (Tabrecta®).** This compound is an orally bioavailable inhibitor of c-MET [597]. The information about the effects of this compound in pediatric oncology is limited. There are reports of its action in pediatric HGG in which this compound appeared to be more efficient than crizotinib in terms of specificity, potency and brain availability, resulting in a higher cellular response compared to crizotinib treatment in vitro and in vivo [698]. Nevertheless, in a phase I dose escalation study that included EWS and OS patients, only mild responses were observed [699].

**Tepotinib (Tepmetko®).** This compound is a phenylmethyl-pyrimidine derivative developed to disrupt MET phosphorylation that received approval from the FDA and the Japanese Ministry of Health, Labour and Welfare for the treatment of patients with metastatic NSCLC harboring METex14 skipping alterations who progressed following platinum-based cancer therapy [700]. According to PubChem (CID 25171648), this compound has been investigated in the treatment of neuroblastoma.

**PF-04217903.** This compound is an ATP-competitive small-molecule inhibitor with 1000-fold selectivity for c-MET compared with more than 150 kinases, making it one of the most selective c-MET inhibitors described to date. In vitro, it inhibited tumor cell proliferation, survival and migration/invasion in cell lines where c-MET is activated by different mechanisms, including c-MET gene amplification, HGF/c-MET autocrine loop formation and c-MET overexpression [701]. In vivo, oral administration or subcutaneous minipump infusions led to a robust tumor growth inhibition at doses of 30 mg/kg with suitable tolerability. Reductions in microvessel density were also observed [701]. Considering pediatric tumors, similar results were obtained when two highly metastatic OS cell lines were injected by tail vein into immunodeficient mice. In this experiment, mice were treated with PF-04217903 (30 mg/kg) or vehicle control by gavage 5 days on and 2 days off for 30 days. Mice injected with MNNG-HOS cells (which has constitutively activated MET) treated with PF-04217903 had a tenfold reduction in the number of metastatic nodules, while those with injected MG63.2-derived tumors (which have high levels of total and phospho-MET) had a 37% reduction in nodules compared to control mice [702]. This compound has also shown potential for the treatment of malignant peripheral nerve sheath tumors in NF1 patients [703].

**Tivantinib.** This compound, also known as ARQ 197, was described as an orally bioavailable small-molecule c-MET inhibitor with antitumor activity. Tivantinib inhibited cell viability with similar potency in both c-MET-addicted and nonaddicted adult carci-

noma cells, pointing to alternative mechanisms of action [704]. Despite this, the failure of a pioneer phase I clinical trial in pediatric tumors was attributed to the lack of selection for MET amplification during patient enrollment. In the study, which comprised 36 patients, including 4 glioma, 4 MB, 4 EPN, 4 EWS, 4 OS, 3 RMS, 2 WT and 2 NB, sub-optimal responses were achieved when tivantinib was given with food to children with refractory solid tumors is 240 mg/m<sup>2</sup>/dose. Moreover, while the drug was well tolerated, its pharmacokinetic profile was also variable, discouraging further investigation in this setting [705]. However, two of those patients (alveolar soft part sarcoma) who responded to tivantinib administration and were transitioned to a follow-up protocol experienced extended progression-free survival receiving 360 mg twice every day without adverse events [706].

**Lorlatinib (Lorbrena®).** This small molecule represents an orally available, ATP-competitive inhibitor developed by Johnson et al., and further investigated for the treatment of ALK-positive NSCLC [707]. Also named PF-06463922, the drug has shown minimal toxicity in adults and there has been much interest in its prospective use in NB treatment. In this regard, Infarnato et al. described higher potency of PF-06463922 across ALK variants in a panel of 10 NB cell lines, with IC<sub>50</sub> values for inhibition of F1174L- and F1245C-mutated ALK significantly lower than those seen for its predecessor, crizotinib (0.2–10 nmol/L) [708]. Moreover, this compound at 10 mg/kg/day induced complete tumor regression in xenograft mouse models of NB, and in (PDX) harboring the crizotinib-resistant F1174L or F1245C mutations within 3 weeks [708]. Similar 10-fold lower IC<sub>50</sub> values were obtained by Guan et al. (2016). In another group of cell lines, PF-06463922 inhibited growth, reduced levels of tyrosine 1278 (Y1278) phosphorylation on ALK, and induced apoptosis. Comparatively, treatment reduced tumor volume in subcutaneous and orthotopic xenograft models of NB, as well as in the Th-ALKF1174L/MYCN-driven transgenic NB mouse model [709]. PF-06463922 has also been tested sporadically in patients affected with NB. Two recent articles portray favorable responses in a 3-year-old boy with ALK-fusion-positive HGG and an adolescent with relapsed, refractory, metastatic ALK F1174L-mutated NB. The first, considering that the compound is able to cross the BBB, was treated through a nasogastric tube at a dose of 95 mg per square meter of body surface area once daily [710]. Histology after tumor resection showed a marked decrease in the proliferative index of the tumor and since the tumor was not seen on postsurgical MRI, therapy stopped. After 6 months, metastatic lesions were identified on cranial nerve VII and treatment was restarted at a dose of 95 mg per square meter administered by mouth once daily, achieving a near-complete response after 1 month [711]. In the second case, the patient had already shown no response to the first-generation ALK inhibitor crizotinib (240 mg/m<sup>2</sup>/dose given twice daily combined with the standard cytotoxic chemotherapy regimen). The tumor was reduced with continuous 95 mg/m<sup>2</sup>/dose lorlatinib and the only significant side effect observed was grade 2 hypercholesterolemia. However, differentially from the infant, she relapsed after 13 months of treatment and died from progressive disease 3 months later [712].

**Ceritinib (Zykadia™).** Formerly known as LDK378, it is an oral ALK inhibitor that also targets insulin-like growth factor receptor IGFR, insulin receptor and ROS1. This compound was approved by the FDA through an accelerated process to treat ALK-positive metastatic NSCLC [713]. Preclinical studies in the pediatric setting have indicated antiproliferative effects and improved inhibition (11-fold) compared to crizotinib [714]. However, in an exploratory study with a panel of NB cell lines, it was noted that inhibition occurs irrespective of ALK mutational status, and cell lines that carry other driver mutations (i.e., MYC amplification) are sensitive to treatment as well. The same authors further treated a child with ALK-I1171T high-risk NB that was not responding to conventional treatment due to an underlying congenital genetic condition, Fanconi anemia. Monotherapy with ceritinib was well tolerated and resulted in tumor shrinkage and complete clinical remission including all metastatic sites [715]. This compound can be given with food and penetrates the human brain, and thus presents itself as an option for the treatment of CNS tumors with ALK alter-



ations such as EPN and MB [715–718]. However, in orthotopic PDX (from a 10-year-old boy with a multiple recurrent GBM), it was observed that even though ceritinib-treated mice lived longer, the drug had only a moderate effect [719]. Monotherapy was also inefficient in treating a 16-year-old patient with a long history of OS lung metastases, despite acceptable results in primary tumor cells of six other patients and the HOS cell line [720]. Similarly, Ceritinib treatment led to decreased cell proliferation, cell cycle arrest and apoptosis in a dose-dependent manner in a panel of RMS cell lines, all of which lack intrinsic ALK phosphorylation (PAX3-FOXO1-positive Rh30, Rh41 and -negative Rh18 and RD cell lines). The work showed that the compound affects the IGF1R signaling pathway without effects on the migratory ability of cells. Moreover, in subcutaneous Rh41 xenografts, a reduction in tumor growth was observed after approximately 2 weeks, albeit subsequent evaluation of tumor characteristics showed no difference in proliferation or vascularization between the treatment groups and controls [721]. Others also showed that even though LDK378 reduces cell viability and induces cell death in RMS cell lines at low micromolar concentrations irrespective of ALK expression levels or phosphorylation status, cells are far less sensitive compared with Karpas 299 non-Hodgkin's lymphoma cells carrying the NPM–ALK fusion gene [722].

**Brigatinib (Alunbrig®).** Originally named AP26113, this next-generation ALK inhibitor was first described in 2016 and is considered highly CNS-penetrant [723,724]. This compound was granted approval for the treatment of patients with metastatic ALK+ NSCLC and intolerance to crizotinib [725]. In an NB setting, preliminary indication of efficacy was observed after exposure of several NB cell lines, including CLB-BAR (MYCN amplification, ALK ( $\Delta$ 4-11) and amplified, ALK addicted), CLB-GE (MYCN amplification, ALK (F1174V) amplification, ALK addicted), IMR32 (MYCN amplification, WT ALK) and CLB-PE (MYCN amplified, WT ALK), in which treatment inhibited cell growth and ALK phosphorylation in a dose-dependent manner. However, while  $IC_{50}$  values varied between 75 and 100 nM in ALK-addicted cell lines, the compound was unable to inhibit growth of both non-ALK addicted NB cell lines, IMR32 and CLB-PE. The effects of brigatinib were further validated in vivo through two complementary models. The first used transgenic *Drosophila melanogaster* flies expressing two gain-of-function variants, F1174L and R1275Q, which disrupt the eye morphology, giving a “rough phenotype”. The authors showed that larvae grown on food containing Brigatinib displayed a concentration-dependent improvement of the rough eye phenotype. Then, brigatinib was used as a single agent to treat BalbC/NUDE mice bearing ALK-addicted CLB-BAR xenografts. In this model, the compound also showed to be effective, with robust and potent anti-tumor activity [726].

**Entrectinib (Rozlytrek®).** This compound (also called RXDX-101, NMS-E628, NMS-01191372, Rozlytrek) is a selective, oral tyrosine pan-TRK, ALK and ROS1 inhibitor that has demonstrated preclinical efficacy in tumors with NTRK1/2/3, ALK and ROS1 alterations [727]. This inhibitor can pass through the BBB and has clinically proven to be effective against primary and metastatic brain diseases, with no adverse off-target activity [728].

Entrectinib also displays promising anti-tumor activity in NB, evinced by diminished Ki-67 and activation of caspase-3 in ALK wild-type, amplified or mutated cell lines [729]. In vivo growth inhibition and substantially reduced phosphorylation in TrkB-expressing NB xenografts were also observed after treatment as a single agent or in combination with irinotecan or temozolomide (TMZ), eliciting increased EFS when compared to controls [730]. Moreover, the ability of entrectinib to inhibit p-TrkB, p-PLC $\gamma$ , p-Akt and p-Erk suggested that this compound may have improved efficacy compared to other targeted inhibitors previously evaluated in NB [172]. However, despite durable responses in pediatric patients with intracranial tumors or NB harboring NTRK1/2/3 or ROS1 fusions, its utility may be hampered by the appearance of acquired resistance in this tumor type [730,731].

**X-396.** This compound, also known as Ensartnib, is an aminopyridazine-based second-generation ALK/MET inhibitor that holds much clinical promise with increased potency as compared with crizotinib and other second-generation ALK inhibitors such as alectinib and ceritinib [732]. X-396 significantly reduced growth (by 40% at a 3 nM concentration)

and ALK phosphorylation in SY5Y NB cells that harbor ALK-F1174L. Biochemical IC<sub>50</sub> values for MET inhibition were 2-fold higher [733]. Ensartinib was significantly more effective than crizotinib at inhibiting the intracranial growth of the SH-SY5Y NB model harboring the F1174L mutation [732]. Furthermore, the activity of X-396 administered alone or in combination with liposomes carrying ALK-siRNAs (that are active irrespective of ALK gene mutational status) was later tested in a mouse model by Di Paolo et al. (2011). These authors corroborated previous *in vitro* data with a second NB cell line (LAN-5) and showed that in subcutaneous NB models, the compound acted in a dose-dependent manner, with adequate bioavailability, moderate half-life, high mean plasma and tumor concentrations. Moreover, against human NB orthotopic xenografts obtained by implanting of Luciferase stably transduced NB cells, SH-SY5Y-Luc and LAN-5-Luc, into the adrenal gland of nu/nu mice, significant dose-dependent anti-tumor activity was also observed, with even more reduced tumors and prolonged survival with the combination with the liposomal formulation [734].

**Erdafitinib (Balversa™).** This compound is an oral pan-FGFR inhibitor with quinoline structure [735]. Known as JNJ-42756493, this compound is already approved by the FDA for the treatment of advanced or metastatic urothelial carcinoma, and is now under clinical trials that also include childhood CNS tumors [736]. It inhibits FGFR1/2/3/4 with increasing IC<sub>50</sub> values of 1.2, 2.5, 3.0 and 5.7 nM, respectively [737]. This compound inhibited proliferation on five different NB cell lines (SK-N-AS, SK-N-BE(2)-C, SK-N-DZ, SK-N-FI and SK-N-SH) as monotherapy, but showed variable synergistic, additive and antagonistic effects when combined with commonly used cytotoxic agents such as cisplatin, vincristine and doxorubicin [738]. Additionally, IC<sub>50</sub> for FGFR4 inhibition by this compound on the A-204 RMS cell line was determined as 4.5 nM, while treatment of mice xenografts resulted in a 58% volume reduction after 21 days of treatment with daily doses of 30 mg/kg [735].

Erdafitinib has also been tested alone and in combination with cisplatin, vincristine and radiotherapy on the SHH-MB cell lines DAOY and UW228-3. Under all conditions, the cell lines showed dose-dependent decreases in viability and proliferation after 48 and 72 h [739].

**Dovitinib.** Also known as TKI258, this is a multi-targeted tyrosine kinase inhibitor with potent activity against FGFR1/3, VEGFR1/2/3 and to different extents, PDGFR-β, Flt3, c-Kit and CSF-1R, that showed promising results as an antitumoral and antiangiogenic compound [658]. This compound is already in clinical trials in adult patients [740]. However, in pediatric neoplasms, information about preclinical studies is limited. Preliminary results of Dovitinib in NB cells, which express high levels of FGFR, indicated anticancer-activity in this tumor type [741]. Similar results were reported for RMS, albeit it was demonstrated that this inhibitor is not as potent as other FGFR inhibitors (i.e., ponatinib) [656,742]. In addition, due to its ability to cross the BBB, this compound has been indicated as a suitable candidate for the treatment of CNS tumors. In this regard, *in vitro*, it reduced the capacity of EPN cells to re-adhere and proliferate in a dose-dependent manner [658]. In DIPG and GBM, however, true effects on viability were observed at high dovitinib concentrations (>400 nM) [743]. Of note, others showed that despite killing glioma cells *in vitro* (up to 55% of cells at the assay end point), the drug exerted minimal anti-tumoral effects *in vivo*, suggesting a microenvironment-mediated therapeutic resistance mechanism [744].

**Masitinib (Masivet®).** This compound, also known as AB1010, is an orally administered, novel, potent and selective phenyl aminothiazole-type tyrosine kinase inhibitor of KIT, used in the treatment of canine mast cell tumors acting as a blocker of mast cell degranulation, cytokine production and migration of bone marrow cells [745,746]. Masitinib is under clinical investigation in several human malignancies that harbor similar canine KIT mutations (i.e., gastro-intestinal stromal tumors, ovarian and prostate cancer). In fact, this inhibitor acts on several mutated forms of KIT, and other receptors, including PDGFR, FGFR3 and focal adhesion kinase (FAK) [747,748]. Noteworthy, a brain tumor xenograft

model using pediatric GBM cells suggested that masitinib may potentiate the effects of TMZ, providing decreased tumor growth relative to either drug used as a monotherapy [749].

#### 4.2. PI3K/AKT/mTOR Pathway Inhibitors

**Everolimus (Afinitor®).** Everolimus (Afinitor, Novartis) is an orally administered rapamycin derivative approved by the FDA and the European Medicines Agency for the treatment of RCC [749]. This compound reduces tumor cell proliferation and induces apoptosis and autophagy through the phosphorylation inhibition of mTOR [750,751]. Pre-clinically, the combination of everolimus with sorafenib yielded enhanced antiproliferative and proapoptotic effects, potentiated antiangiogenesis and reduced the metastatic potential of OS [751]. Prolonged exposure to everolimus also improved the CNS retention of dasatinib and extended the survival of mice bearing pediatric high-grade glioma tumors [752]. Comparatively, everolimus is synergistic with carboplatin in low-grade glioma models [753]. However, in the literature, there is significantly more information about clinical experience because, since its approval, everolimus has become widely accepted by the medical community where treatment options may be limited. One major clinical example involves subependymal giant cell astrocytomas (SEGA), tumors that are frequently diagnosed in patients with tuberous sclerosis complex (TSC) [754]. Loss of function of either TSC1 or TSC2 leads to downstream constitutional activation of the mTOR complex [755]. Besides surgical excision, patients with large or recurring SEGAs did not have robust treatment options, and Everolimus has been shown to induce tumor shrinkage and presents additional clinical benefits including seizure control [756,757].

**Palomid-529.** Also known as RES-529, this compound is a small-molecule drug dual novel inhibitor of mTOR complex 1 (mTORC1) and mTOR complex 2 (mTORC2). Palomid 529 likewise inhibits both VEGF-driven and bFGF-driven endothelial cell proliferation [758]. Due to its potential to penetrate the BBB without restriction by the ABCB1 and ABCG2 efflux transporters, the anti-glioma effects of this drug have been investigated [759–761]. In childhood cancer, a single report proved a potent inhibition of viability, cell cycle progression and proliferation of the OS cell line U2OS [762].

**OSI-027.** This compound is an orally bioavailable selective ATP competitive inhibitor of mTOR and off-targeted PI3K $\alpha$  (100-fold selectivity for mTOR relative to PI3K $\alpha$ ) that has been studied in the treatment of many tumors [763]. OSI-027 is active in vitro against cell lines and primary cells of pediatric pre-T-ALL, with superior efficacy to rapalogs and in vitro synergy with a number of conventional cytotoxic agents [181]. Preliminary studies combining OSI-027 treatment with alpelisib demonstrate similar antineoplastic results inhibiting PI3K/mTOR signaling MB, EWS and RMS cell lines [764–767].

**VS-5584.** This dual inhibitor of mTORC1/2 and class I PI3-kinases has shown anti-tumor potential in a broad spectrum of tumor types in vitro and in vivo [768–770]. Noteworthy, evidence supports that this compound has an active role in reducing stem cell viability in multiple mouse xenograft models of human cancer (30-fold more potent compared to non-stem cells) [769]. Thus, the activity of VS-5584 was recently explored in OS, in which treatment dramatically suppressed growth and cell migration and synergized with CCT128930, an AKT2 inhibitor [771–773]. In the same way, this drug is cytotoxic, showing apoptosis induction and a robust limitation of the colony-forming ability in NB cell lines. Delay of tumor growth was also observed in mice subcutaneously inoculated with BE(2)-M17 cells and treated with VS-5584 (25 mg/kg, three times per week) for 2 weeks [773].

**Dactolisib (BEZ235).** This drug, also called BEZ235 or NVP-BEZ235, is a reversible PI3K/mTOR inhibitor belonging to the imidazoquinoline class already tested in a variety of cancers in preclinical studies. In sarcomas (EWS, OS and RMS), dactolisib showed promising results in vitro, such as a reduction in cell proliferation, G1 cell cycle arrest and decreased in cell migration [774,775]. Interestingly, it was also shown that BEZ235 elicits strong cytostatic effects in EWS cells and results in a global modulation of the transcriptome affecting other pathways related to splicing and metabolism. This drug also reduced

the expression of EWS/FLI1 by 50%, reinforcing its potential for EWS treatment [773]. However, its capacity to induce apoptosis is uncertain. Mild results were obtained by Giorgi et al. (2018), and when OS cells were treated with a similar inhibitor range, U2-OS and MG63 presented no significant differences in apoptosis induction, although the drug was efficacious with either doxorubicin or vincristine [774–776]. In RB, GBM, and MB, decreased viability and proliferation in a dose-dependent pattern was observed in most cell lines [777–780]. When tested *in vivo*, dactolisib could reduce tumor volume, vascularity and metastasis and improve animal survival, especially when combined with other drugs, such as topotecan, carboplatin, vincristine or the SMO inhibitor LDE225 [773,781–783].

**SF-1126.** This is a pan and dual first-in-class soluble PI3K/mTORC inhibitor that exhibits antitumor and antiangiogenic activity against several malignancies [784,785]. In the literature, there are few reports of this inhibitor in pediatric preclinical models. SF-1126 promoted a decrease in cell viability of a panel of EWS cell lines and CD15+ stem cell population in SHH-driven MB [786,787]. Moreover, SF1126 has been shown to enhance the cytotoxicity of doxorubicin in NB cells, leading to p53-mediated activation of apoptosis [788]. Treatment of NB tumors with SF1126 also reduced MYC expression and inhibited growth *in vivo*, leading to tumor shrinkage and reduced neovascularization [789].

**Triciribine.** Triciribine is a pan-AKT 1 inhibitor with anticancer effects in various tumor types [790]. This compound has been shown to decrease the survival of SH-SY5Y NB cells in both 2D and 3D culture models, affecting the migratory abilities of their sphere-forming units [791]. Triciribine demonstrated activity in EWS cell lines as well, with a mean IC<sub>50</sub> of 24  $\mu$ M, with robust synergy when combined with dasatinib; however, it did not affect tumor growth *in vivo* [792]. Moreover, Smeester and colleagues (2020) tested this drug in OS and showed that ATK inhibition in HOS and SJS-1 cell lines leads to decreased cell proliferation, migration and colony capacity, and increased apoptosis. Further assessment in an orthotopic OS model also demonstrated reduced tumor growth (volume and weight) and metastasis after triciribine 40 mg/kg three times weekly [793].

**Sapanisertib.** Also called MLN0128, INK-128 or TAK-228, this is an ATP-competitive mTOR inhibitor already tested for safety in an adult cohort [794]. In studies including pediatric models, this inhibitor has shown promising results *in vitro*, reducing cell viability and colony formation and inducing apoptosis in sarcoma cells (EWS, OS and RMS) without affecting human osteoblast and osteocyte cells (normal bone cells); effects were improved by combination with MK2206, an AKT-specific inhibitor [795,796]. Similarly, the inhibition of mTOR (oral gavage for 21 days-3 mg/kg twice daily 3  $\times$ /week in EWS and RMS, or 2.5 mg/kg, daily) in OS resulted in tumor volume reduction, without observable side effects [795,796]. Comparable results were obtained for brain tumors (MB, NB and GB), with reduced cell invasion at low concentrations [797,798]. Interestingly, it was also observed that sapanisertib promotes metabolic alterations, such as disrupting glutathione synthesis and reducing glucose and lactate (a common feature of cancer cells) [797,798]. When tested in murine models, it reduced tumor weight and size, improving animal survival [797–799]. However, in combination with trametinib (1 mg/kg; 3  $\times$ /week; p.o. + MAPK inhibitor; 1.5 mg/kg; 5  $\times$ /week; p.o.), despite showing antitumor effects (reduction in angiogenesis and improvement in animal survival), some adverse effects were observed, including weight loss and skin redness [800]. Finally, in RB models, sapanisertib inhibited growth and increased apoptosis, whereas it inhibited cell migration and angiogenesis [801].

**LY-2090314.** This drug belongs to the ATP-competitive class of GSK-3 inhibitors with limited activity against additional kinases. Preclinical data suggested partial anticancer activity as a single agent against solid-tumor-derived cancer cell lines *in vitro* and in xenograft models, although it seemed to potentiate platinum-based chemotherapy [802]. Only a single report of its activity against pediatric tumors was found in the literature. Kunnimalaiyaan et al. (2018) tested LY2090314 in a panel of NB cell lines with different genetic backgrounds: SH-SY-5Y (non-amplified MYCN or single-copy, wild-type TP53, F1174L ALK mutation), NGP (1p alteration, MYCN-amplified, wild-type ALK, TP53 mutated, MDM2-amplified) and SK-N-AS (1p deletion, MYCN single-copy, H168R TP53 mutation, wild-type ALK),



and found that this GSK-3 inhibitor at nanomolar range promoted growth inhibition in a time- and dose-dependent manner irrespective of the cell line markers. Reduced growth resulted mainly due to apoptosis induction, evinced by a 2-fold increase in the expression of cleaved PARP and caspase-3/7 activity. Downregulation of survivin and cyclin 1 was also observed [803].

**Tideglusib.** This compound represents another GSK-3 inhibitor, although it acts in a non-ATP competitive manner. Evidence of its antineoplastic effects with a pediatric scope includes in vitro experiments in OS and NB cell lines. In both models, treatment showed a significant reduction in cell proliferation in a dose-dependent manner, cell cycle arrest, and apoptosis induction, even though micromolar concentrations are required to achieve comparable results to LY2090314 [804–806]. Nevertheless, inhibition of GSK-3 by Tideglusib importantly compromises stem cell characteristics of both cell types. In the OS, treatment decreases stem cell markers, including OCT4, CD133 and SOX2, while in NB, it decreases neurosphere self-renewal. In mice models, tideglusib treatment (10 or 20 mg/kg in OS- and NB-derived tumors, respectively) promoted a reduction in tumor growth with few side effects [804,806]. Of note, PDX-derived cell cultures of both variants of RMS (embryonal and alveolar) treatment with tideglusib substantially reduced  $\beta$ -catenin phosphorylation at 60 nM; however, tumor-bearing mice treated with 200 mg/kg of tideglusib daily by oral gavage did not benefit in terms of survival or myodifferentiation [807].

**MK-2206.** This compound is an orally bioavailable allosteric and non-ATP-competitive AKT inhibitor tested in several tumors [808]. In OS, for instance, this drug was able to induce cytotoxic effects both in vitro and in vivo [223,796,809,810]. Similarly, in NB cells, MK-2206 diminished cell viability and increased apoptosis in cells with high expression of FOXO3a [810,811]. In vivo, the drug promoted inhibition of tumor growth and increased animal survival, effects that were even improved by combination with etoposide [812]. Of note, EWS and RMS cells were not sensitive or had less sensitivity to AKT inhibition [809].

**Ipatasertib.** Also known as GDC-0068, this compound is an ATP-competitive pan-AKT inhibitor developed by Array BioPharma/Genentech Inc. Having similar activity against Akt-1 and Akt-3, it is effective against several tumor types [813]. So far, Choo and colleagues are the only group that has tested this compound in childhood sarcomas. The drug induced a reduction in cell viability; however, RMS cells were more sensitive to PI3K/AKT pathway inhibition than OS cells [814].

#### 4.3. MAPK Pathway Inhibitors

**Sorafenib (Nexavar®).** Sorafenib is an inhibitor of VEGFR2/3, PDGFR, KIT, FGFR-1, RAF and RET, approved by the U.S. FDA for the treatment of unresectable HCC and advanced RCC [815]. In preclinical models of MB, this compound reduced cell viability and increased apoptosis in established cell lines and primary tumor cultures. Moreover, it induced cytoskeletal alterations that ended in impaired cell migration [640]. In vivo, sorafenib (100  $\mu$ L of 10  $\mu$ mol/L administered three times a week for five weeks) was able to reduce the volume of subcutaneous tumors [816]. Similar results were obtained NB, in vitro and in vivo. Of note, in this model, sorafenib also impaired angiogenesis and G1 cell cycle arrest [817,818]. Conversely, despite reducing initial viability in EPN and PA, growth-factor-driven rescue was also seen, reducing the potential of using sorafenib for treatment of these tumors [691]. Indeed, in pediatric patients with PA, sorafenib induced progressive tumor growth acceleration as a result of ERK upregulation, which resulted in premature termination of the study [819]. In bone sarcomas, dubious results were also observed. In OS, sorafenib treatment blocked cell proliferation and was able to reduce tumor growth in murine models [820–822]. However, other studies showed that sorafenib was only able to reduce tumor growth when combined with everolimus, or palbociclib, probably due to the capacity of sorafenib to induce mTORC activation [821,823]. At the same time, in EWS and RMS, sorafenib only showed efficacy when combined with doxorubicin or with ceritinib, respectively [823,824].

**Regorafenib.** Also called BAY 73–4506, this is a new-generation multi-tyrosine kinase inhibitor that already showed antitumor and antiangiogenic effects. This inhibitor diminished cell proliferation in a cell line panel from the Innovative Therapies for Children with Cancer (ITCC), which includes 5 MB, 7 EWS, 7 NB, 7 OS and 7 RMS cell lines [610]. Regorafenib also induced cell cycle arrest and promoted apoptosis in NB cells [612]. In vivo, 10 mg/kg/d or 30 mg/kg/d treatment resulted in tumor growth inhibition in RMS, EWS and NB orthotopic models and improved EFS in EWS, NB, OS and RMS [610,825,826].

#### 4.4. Cyclin-Dependent Kinases Inhibitors

**Milciclib.** This is a second-generation ATP competitive pan-CDK inhibitor, developed by Tiziana Life Sciences, that also acts on TRKs (from tropomyosin receptor kinase A) [827,828]. In preclinical trials performed in different tumors, such as MB and gliomas, it showed promise given its ability to cross the BBB, even though there are reports of MDR transporters limiting the penetration into the brain [829–831]. Moreover, MYCN-amplified the Grp3-MB cell lines MB002, Sd425 and D283 and the MYCN-amplified NB cell line Kelly are particularly sensitive to MILCICLIB treatment, evinced by cell cycle arrest and massive apoptosis [829].

**Terameprocol (CINelim™).** This is a semi-synthetic inhibitor developed by Erimos Pharmaceuticals LLC from a plant lignan, showing antiviral and anti-cancer potential [829,832]. The drug is considered a global inhibitor of the transcription process, which in turn acts by preventing, for example, the synthesis and activation of survivin, by competing with the transcription factor Sp1 for specific Sp1 DNA-binding domains within gene-promoter regions during DNA synthesis [833]. To date, only a single in vitro study (that included the childhood GBM cell line SF188) showed that this inhibitor was able to reduce the proliferation capacity of the cells in a dose-dependent manner, and showed synergism with TMZ under simultaneous exposure for 48 h. Increased effects were also observed when combined with ionizing radiation. Moreover, as expected, this compound induced significant arrest in the G0/G1 phase, decreasing the mitotic index and almost killing all cells at 30  $\mu$ M [833].

**UCN.** UCN-01, or 7-Hydroxystaurosporine, is a synthetic derivative of staurosporine with antineoplastic activity that acts on AKT, CDKs and calcium-dependent protein kinase C (in an ATP-competitive manner), and is able to act synergistically with others [834]. In experimental models described in the literature, UCN-01 showed promising results by inducing apoptosis in leukemic and colon cancer cells; moreover, according to the pediatric tumors included in this work, this inhibitor also showed potential against OS tumor cells, reducing viability, proliferation and migration [835,836]. Similar results were observed in a panel of NB cell lines (with genetic backgrounds differing in MYC, p53 and BCL2 statuses), where this inhibitor was the most effective compound in reducing cell proliferation (compared to BiCNU, docetaxel, flavopiridol, staurosporine) and induced apoptosis measured through both caspase activation and caspase-3 and PARP cleavage [837].

**BMS-387032.** Also called SNS-032, this is a small aminothiazole molecule that acts as an ATP-competitive cyclin CDK inhibitor, especially for CDK2/7/9. Preclinical studies have shown that as a cell cycle blocker, it causes cytotoxicity and prevents tumor cell growth in several models [838–840]. Regarding pediatric tumors, this inhibitor showed positive results in the OS cell line U2-OS, evoking downregulation of RNA polymerase II Ser2 phosphorylation and some degree of caspase activation at all doses tested [839]. Similarly, this CDK inhibitor showed encouraging results in a panel of 109 NB cell lines, consisting of 19 parental cell lines and 90 sublines with acquired resistance to 14 different anticancer drugs. Doses between 58.3 and 14,615 nM were able to reduce viability in a great proportion of cell lines and impaired the growth of the multidrug-resistant cisplatin-adapted UKF-NB-3 subline UKF-NB-3(r)CDDP(1000) injected into the right flank of NMRI:nu/nu mice. Of note, p53 status did not affect the response of NB cells; however, ABCB1 expression conferred resistance to this drug [841–843]. Other interesting results, albeit not in child-derived cell lines, were also published, including the inhibition of hypoxia-mediated GBM

cell invasion and cell-mediated capillary formation of HUVEC cells when co-cultured with U87MG cells in the presence of the drug [844,845]. Nevertheless, further studies with this inhibitor were stopped due to its high toxicity and low selectivity [846–848].

**Seliciclib (Roscovitine®).** Formerly known as Roscovitine, CYC202 or R-roscovitine, this is a selective ATP-competitive pan-CDK inhibitor that blocks cell proliferation in almost all phases of the cell cycle. Seliciclib is a potent inhibitor of CDK9/cyclin T, CDK7/cyclin H, CDK2/cyclin E and CDK1/cyclin B. The negative influence of seliciclib on CDK7 and CDK9 also portrays a role for this inhibitor in modulating RNA polymerase II CTD phosphorylation [848]. Its antitumor activity has been explored in a wide spectrum of hematological and solid malignancies as a single agent and in combination with other cytotoxic agents [848–850]. Among pediatric tumors, Roscovitine showed promising results, in EWS, where it was able to reduce cell proliferation and induce caspase-dependent activation (half minimal dose 10  $\mu\text{mol/L}$ ) in a panel of six cell lines, while it slowed A4573-derived tumor growth in mice after intraperitoneal injection [851]. Similar results were found in OS, with reduced proliferation and migration at doses up to 90  $\mu\text{M}$  [437,851]. Moreover, in NB, the drug resulted concentration-dependent cytotoxicity, both in vitro and in vivo, with doses between 10 and 200  $\mu\text{M}$  [851–855]. Roscovitine also reduced MB viability, with IC<sub>50</sub> values of around 25  $\mu\text{M}$  [856]. Moreover, treatment of Pzp53med cells (derived from a mouse Ptc+/-/p53-/- tumor) with 10 nM roscovitine resulted in reduced levels of E2F1, FASN, Bmi1, cyclin D2, cdk2 and cdk4. Synergistic effects were also observed when combined with C75, an inhibitor of FASN [857].

**Ribociclib (Kisqali®).** Also known as Kisqali® (Novartis, Basel, Switzerland), this compound is a highly specific inhibitor of CDKs 4/6 that received FDA approval for use in the upfront treatment of hormone receptor-positive (HR<sup>+</sup>), HER2-negative breast cancer in 2017 [858]. With respect to the pediatric tumors, this compound demonstrated adequate results in EWS, causing cell cycle arrest mainly in combination with IGF1R inhibitors [859]. Moreover, in NB, this drug was able to reduce proliferation in vitro and in vivo, with doses between 0 and 10,000 nmol/L [331,860,861]. Most importantly, ribociclib showed high CNS penetration (>10 nM) in vivo, suggesting prospects for its use in the treatment of brain tumors. In this regard, oral doses of ribociclib inhibited RB phosphorylation, downregulated E2F target genes (CNE2, CCNA2, MKI67, TOP2A and PLK1) and decreased proliferation in group-3-MB mouse and human orthotopic PDX. Additionally, the combination of ribociclib and gemcitabine slowed tumor progression and metastatic spread and increased survival, warranting further investigation [862].

**Palbociclib (Ibrance®).** Also known as PD-0332991 (Ibrance®, Pfizer, New York, USA), this compound represents an ATP-competitor with selective potency against CDK4/6, approved by the FDA in 2015 [859]. The effects of this inhibitor have been assessed in several childhood tumors. In primary EPN cells, for example, it was able to reduce proliferation at 0.5  $\mu\text{M}$ , with G1 arrest and reduced expression of CDC6, MCM2, MAD2L1, CDK2, BRCA2 and RAD51 [863]. Similar results were observed in NB, where this inhibitor reduced proliferation, inhibited colony formation in a dose-dependent manner and affected cell differentiation, tumor progression and metastasis in a preclinical chick embryo model [863–866]. Palbociclib has also been shown to be a new option for targeted therapy in childhood sarcomas. Perez et al. (2015), by treating a panel of 10 low-passaged sarcoma cell lines generated directly from patient samples and two commercial cell lines of heterogeneous origin and different molecular karyotypes (including liposarcoma, leiomyosarcoma, EWS, RMS and myxofibrosarcoma), determined IC<sub>50</sub> values ranging from 8 to 26  $\mu\text{M}$  depending on their levels of CDK4 expression. Moreover, palbociclib was active in vivo against subcutaneously engrafted CDK4-expressing sarcomas, although responses were negative in tumors displaying low levels of CDK4 and high levels of p16ink4a [867]. Strong decreased cell proliferation and G0/G1-phase arrest with decreased S/G2 fractions were also observed in leiomyosarcomas by another group [868]. Most interestingly, this compound is capable of inhibiting growth in sarcomas with different translocation backgrounds. For example, Palbociclib (100 mg/kg) was able to reduce the volume of tumors originated

from an EWS sample with CDKN2A/B loss and FUS-ERG fusion implanted in the right chest wall of nude mice [869]. Additionally, satisfactory results were obtained after the treatment of a child with a refractory pediatric sarcoma harboring paracentric inversion on the short arm of chromosome X, resulting in the fusion of the BCOR and CCNB3 genes [870]. Regarding assays in OS, this inhibitor reduced proliferation and migration with doses of 0.04, 0.16, 0.625, 2.5 and 10  $\mu\text{M}$  [393]. Migration and invasion have also been hampered by palbociclib in glioma cell lines, both in vitro and in vivo, with doses ranging between 10 nM and 10  $\mu\text{M}$  [871–873]. Moreover, this inhibitor showed significant therapeutic benefit in mice after intracranial transplant of genetically relevant murine or human astrocytoma cells expressing BRAFV600E, and extended survival of animals when combined with PLX4720 (PubChem CID24180719) [874]. Similar results were also obtained in a DIPG with PDGF-B overexpression and Ink4a-ARF loss. Palbociclib induced cell cycle arrest in vitro and in vivo. However, in models engineered for PDGF-B expression with p53 deletion, the results were disappointing. Regarding survival, Palbociclib treatment prolonged animal survival by 12%, which was further increased by combinations with a previous single dose of 10 Gy radiation therapy [875].

**Abemaciclib (Verzenio®)**. This compound, under the name Verzenio® (Eli Lilly, Indianapolis, USA), is a highly selective CDKs 4/6 inhibitor that acts by competing for the ATP binding site. This inhibitor is the most different from its peers (palbociclib and ribociclib), being more lipophilic and able to cross the BBB and penetrate breast tissue [859]. In addition, this inhibitor has potent activity against recurrent ER+/HER2- breast cancers [876,877]. However, its clinical adverse effects are not well described [878–880]. Preclinical studies in pediatric tumors indicate effectiveness against EP, NB, EWS and OS [412,880–882]. Moreover, in gliomas, this inhibitor has been shown to be efficient in reducing cell migration and invasion, as well [883,884]. Finally, combining abemaciclib with other inhibitors, one of them being trametinib (MEK inhibitor), synergistically reduced the survival of the RAS-mutant RMS cell line RD. However, when PDX-bearing mice were treated with that combination, they exhibited progressive disease compared to the RMS standard-of-care regimen (irinotecan + vincristine) [885].

**AT-7519**. This is a potent pan-CDK inhibitor, acting on CDK1/2/4/6/9. Preclinical studies have shown a reduction in cell proliferation and induction of cell death in many cell lines, regardless of tumor origin [886–888]. In addition, this inhibitor showed promise for the treatment of MYC-amplified NB, evinced by apoptosis induction in vitro and dose-dependent growth inhibition in PDX, with improved survival and tumor regression in 86% of patients 7 days of treatment initiation [889].

#### 4.5. Polo-Like and Aurora Kinases Inhibitors

**BI-2536**. This is an ATP-competitor dihydropteridinone that has proved to be more than 1000 times more specific for PLK1 than for other kinases [471,890]. This compound has been tested in several tumor cells, although reports for pediatric tumors are more uncommon. Our group showed that this compound reduces proliferation in up to 64% of cases, causes G2/M arrest and induces apoptosis after 24 h of treatment in the SF188 cell line, and it exerts the strongest radiosensitizing effect among all the cell lines tested [891,892]. Anti-mitotic and sensitizing to ionizing radiation effects were also demonstrated by us in MB cells, even though the results were comparable to other PLK1 inhibitors [893]. Others also showed that this compound suppresses self-renewal of patient-derived primary cells with high PLK1 but not low PLK1 expression, and it did not affect the growth of normal neural stem cells. Finally, BI2536 extended survival in MB-bearing mice [487,488]. Sensitizing effects were also observed for hyperthermia in the RB cell lines Y79 and WERI-Rb-1 [894].

With  $\text{IC}_{50}$  lower than 100 nM, BI 3526 was also able to induce cell cycle arrest at the G2/M phase and cell apoptosis in NB cells [492,895,896]. It has recently been proposed that this drug induces cell death by regulating the expression of the minichromosome



maintenance complex components 2 and 10, which are involved in DNA replication and have been associated with poor outcome in other tumors [897].

Perturbation of normal mitotic progression by BI 2536 nanomolar concentrations (10, 50 and 100 nmol/L) also significantly decreased cell proliferation and clonogenic capacity, inducing mitotic arrest and aneuploidy in OS cell lines, resulting in caspase-independent mitotic catastrophe followed by necrosis [898]. Conversely, in another set of OS cell lines, apoptosis induction was validated through PARP cleavage and caspase activation. Irrespective of this, BI 2536-treated xenograft mouse models presented significantly smaller tumors compared with controls [899]. Moreover, in RMS, PLK1 inhibition by BI 2536 led to elevated ubiquitination and rapid proteasomal degradation of the PAX3-FOXO1 chimeric oncoprotein in vitro, whereas it reduced PAX3-FOXO1-mediated gene expression and elicited tumor regression in a xenograft mouse model [900]. Moreover, in this tumor type, this drug presented high antiproliferative activity when combined with Eribulin, a microtubule-interfering drug [901].

**NMS-1286937.** Also known as Onvansertib or NMS-P937, this novel PLK1-specific inhibitor has shown high potency at low nanomolar concentrations on a large number of cell lines, both from solid and hematologic tumors; in addition, differentially from other PLK1 inhibitors that compulsorily need intravenous administration, this small molecule can be administered orally [902]. Considering pediatric tumors, onvansertib has shown promising results in OS and MB. In the former, this drug proved to be highly active in both drug-sensitive and drug-resistant cell lines, except for cell lines overexpressing the multiple-drug-resistant transporter ABCB1 [903]. Results were also very promising in group-3 MB, which is characterized by PLK1 overexpression. In the study, treatment of D341, D425 and D458 cell lines resulted in reduced colony formation, cell proliferation, stem cell renewal and G2/M arrest. The half-maximal inhibitory concentrations varied from 4.9 to 6 nM. Other cell lines within the SHH subgroup needed 27.94 nM for comparable results. Moreover, onvansertib acted as a radiosensitizer, and showed marked time- and dose-dependent growth arrest of neurospheres and patient-derived short-term cultures. Most notably, onvansertib dramatically improved the median survival of orthotopic PDX models from 68 to 95 days [904].

**GSK-461364.** This compound is a second-generation and potent ATP-competitive thiophene amide PLK1 inhibitor. The anti-mitotic effects of this compound have been demonstrated in several tumors; however, it has been observed that its activity can be hampered by the overexpression of the multidrug resistance pump ABCB1 [905]. Preclinical findings in the pediatric setting include reduced viability after treatment in NB, MB and OS, in all of which it diminished growth and caused cell cycle arrest with massive apoptosis at a low-dose nanomolar range [906,907]. This compound also demonstrated a synergistic cytotoxic effect with paclitaxel, even though combination with methotrexate, cisplatin, vinblastine or doxorubicin was not that effective [907]. Conversely, this PLK1 inhibitor has been shown to be an effective radiosensitizer [908]. In vivo, GSK461364 treatment (50 mg/kg body weight intraperitoneally administered) strongly delayed the establishment of high-risk NB tumors in nude mice (by 22 days) irrespective of MYC status of the cell lines used, and significantly increased survival time in the treated group [906].

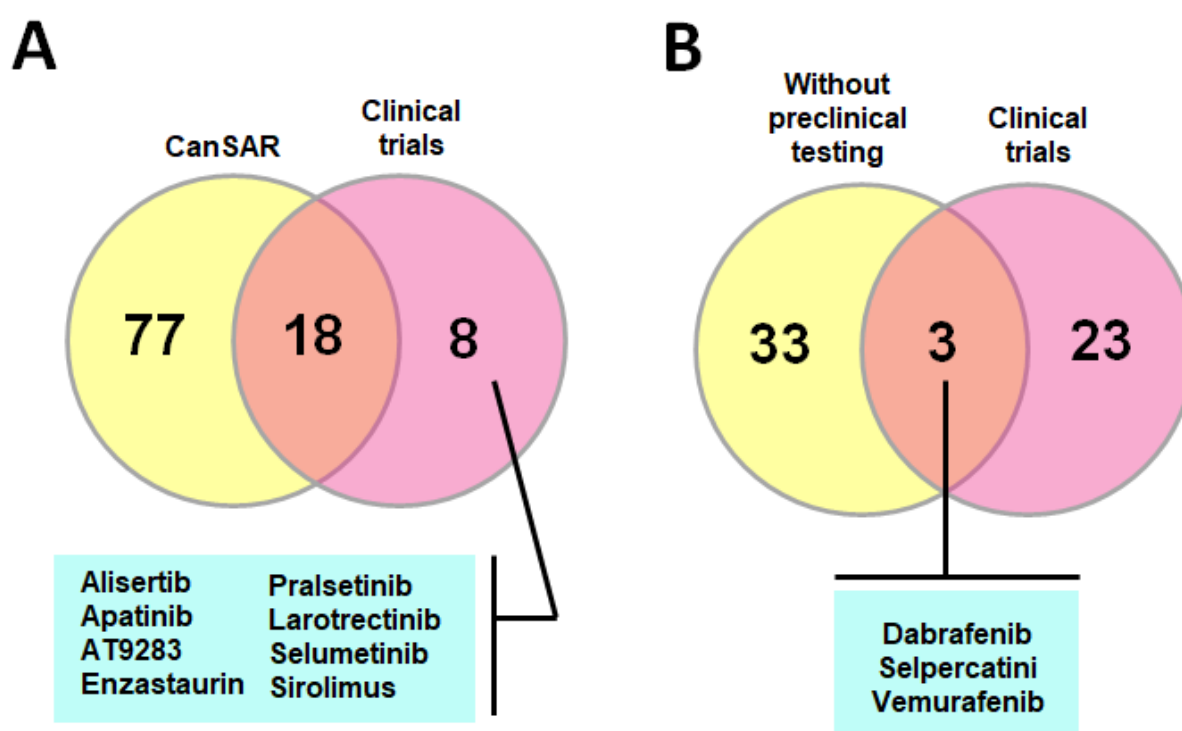
**Volasertib.** This drug, also known as BI 6727, is also a dihydropteridinone derivative that induces a distinct prometaphase arrest phenotype (polo-arrest) and subsequent apoptosis. Regarding pediatric tumors, in 2014, the NCI-supported PPTP Program published initial results about the use of BI 6727 in pediatric tumors. The systematic work presented in vitro results on 24 cell lines that included 4 RMS, 4 EWS, 1 GBM, 4 NB and several leukemias, concluding that the compound was effective without histotype selectivity. Then, the responsiveness of solid tumor xenografts (that also included OS and WT) using a dose of 30 mg/kg for 3 weeks was also tested. Volasertib was able to induce regression in only a minority of the models tested; however, significant differences in EFS distribution compared to control in 59% of the evaluable xenografts were observed, with better results for OS, WT and NB (the only one that showed objective responses) [909]. The results for OS

were later validated in vitro by our research group, where treatment with BI 6727 not only led to growth arrest, triggered apoptosis and radiosensitized cells, but also it seemed to be more efficient in sensitizing OS cells to standard cytotoxics compared with GSK461364 [908]. Anti-mitotic effects were also observed in MB cells [893].

**Aurora kinase inhibitors.** Over the past two decades, several small-molecule inhibitors of Aurora kinases have been developed, most of which primarily target Aurora B. Despite not being approved or with clinical promise by the CanSAR platform, some of these inhibitors have shown promising results. MLN8237, for instance, was evaluated against a panel of EWS ( $n = 11$ ) and NB ( $n = 17$ ) cell lines with acceptable results in vivo [910]. The drug also inhibited growth uniformly in the majority of the cell lines from the PPTP in vitro panel, with  $IC_{50}$  values ranging from 49 nM to 61 nM. In vivo, EFS was 80% higher in treated animals compared to controls, showing even more auspicious results than those obtained for Volasertib [911]. More recently another AURK inhibitor, designated as PHA-680626, disrupted the AURKA/N-Myc, presenting a new alternative for the treatment of high-risk NB [912,913]. Additionally, AMG-900 (pan-aurora inhibitor) blocked MB cell proliferation and increased apoptosis and acted synergistically with the histone deacetylase inhibitor SaHa [914].

### 5. Kinase Inhibitors in Clinical Trials

Clinical trials including children and using kinase inhibitors have been increasingly reported in recent decades. Among the 95 predicted compounds for our selected group of kinases retrieved through the CanSAR platform, for instance, 18 have already entered clinical trials (Figure 6A), most of which have focused on measuring cytotoxic effects on high-risk, refractory and recurrent tumors.



**Figure 6.** (A) Venn diagram showing the number of kinase inhibitors retrieved from the CanSAR platform versus those already tested in clinical trials. Around 30% of the compounds have been tested in patients. Of note, eight kinase inhibitors tested in clinical trials were not found in the CanSAR platform. (B) Venn diagram comparing kinase inhibitors found in the CanSAR platform without evidence of preclinical tests in pediatric cancer versus clinical trials. Three of the drugs are already tested in pediatric patients, without in vitro or in vivo evidence.

In order to gather an updated and comprehensive review on clinical data and outcome of pediatric tumors treated by these new TKIs compounds and already published to date, a PubMed search was performed (as per Oct 2022) using the following uniterms: ((cancer) AND (pediatric)) AND (kinase inhibitor). The following additional filters were set: clinical trial, meta-analysis, randomized controlled trial. In total, 233 articles were retrieved. Abstracts of the whole set of results were carefully read to exclude duplicated data/patients from the same clinical trial, articles where children and/or adolescents (<21 years of age) were not included or where clinical trials not testing kinase inhibitors were retrieved. A total of 53 articles met these inclusion/exclusion criteria, and were read and analyzed in full. Data on study design, study population, main clinical information and outcomes are summarized in Supplementary Table S5. The next part of this section does not review all studies on this subject, and also does not include clinical trials under development or in recruitment with unpublished data. Our main purpose was to gather and to discuss major sedimented data of clinical value and of clinical interest and applicability in this setting.

### 5.1. TRK—Tyrosine Receptor Kinases

#### 5.1.1. EGFR and VEGFR

Abnormal or disrupted angiogenesis is considered to be one of the hallmarks of cancer, and an increasing interest in targeting EGFR and VEGFR pathways has been observed in clinical oncology [915]. Unfortunately, most of these studies are focused on the adult population, and experience with these drugs in pediatric cancer is less robust. Yet, many of these compounds were tested in clinical trials that included children and/or adolescents with cancer, with variable results.

Regarding VEGFR inhibitors, at least three major molecule subtypes have been described. Type I VEGFR inhibitors exert their action as competing molecules to ATP [916]. Some examples of type I VEGFR inhibitors include pazopanib, axitinib, sunitinib, ponatinib and others. Type II inhibitors bind to the inactive “DFG-out” conformation adjacent to the ATP-binding site. Some examples of type II inhibitors include sorafenib and lenvatinib. Type III inhibitors lead to an irreversible binding of kinases at specific sites [917]. In addition, many of these VEGFR inhibitor molecules are under clinical evaluation in association with different inhibitors, particularly immunotherapy [918]. In addition, VEGFR2 inhibitors of dual action against other tumor-associated biomarkers are gaining much attention lately [919].

Inaba et al. (2011) evaluated the combination of sorafenib, a potent multikinase inhibitor, in association with cytarabine and clofarabine to treat relapsed or refractory childhood leukemia [916]. A total of 12 patients (<21 years of age; 11 with acute myeloid leukemia (AML) and one with early T-cell precursor leukemia) entered this phase I study. Of note, complete remission (CR) was obtained in 6 out of 12 patients, CR without complete blood recovery in 2 cases and partial remission (PR) in 1 case. Dermatologic, gastrointestinal (GI), metabolic and infectious adverse events were observed, and were more pronounced in the sorafenib higher-dose stratum. Grade 3 hand-foot skin reactions and/or rash were dose-limiting toxicities (DLTs). Sorafenib was also evaluated in a phase II trial, in association with everolimus to treat patients with progressive and unresectable high-grade osteosarcoma who failed standard treatment [920]. Although some encouraging initial results with sorafenib were observed earlier in this setting, a larger phase II study by Grignani et al. has shown some minor activity for selected cases, and the trial did not reach the 6-month progression-free survival target in at least 50% of patients [920]. Sorafenib was also evaluated in a phase I study that included refractory or relapsed hepatic tumors (hepatoblastoma or HCC) in children. The drug was used in association with irinotecan [921]. Six patients were evaluable for tumor response: two patients survived with no evidence of disease (NOD), one patient was alive with disease (AWD) and two patients DOD upon publication date. Radiation therapy and/or metastasectomies were offered after study protocol based on individual clinical needs. Increased grade 3 or 4 transaminase levels or neutropenia were reported.

Axitinib, a VEGFR1, 2 and 3 inhibitor, was also evaluated in a phase I study that included refractory solid tumors, as part of a Children's Oncology Group (COG) trial and a pilot consortium trial ADVL1315. Nineteen patients were evaluated, with ages ranging from 9 to 17 years. Five patients achieved stable disease (SD), and a PR was observed in one case (an alveolar soft tissue sarcoma). The maximum tolerated dose (MTD) of axitinib was set at 2.4 mg/m<sup>2</sup>/dose [922]. Lenvatinib, a multiple oral tyrosine-kinase inhibitor against VEGFRs 1 to 3, RET, KIT, FGFRs and PDGFR-alpha were evaluated in a phase I/II pediatric and young adult trial for osteosarcomas [631]. The phase I study observed SD (some lasting for 23 weeks) as the best response obtained with Lenvatinib; the phase II study depicted two patients with partial response and thirteen children with SD. Although this single agent showed some activity in osteosarcoma, future studies will focus on the association of Lenvatinib with chemotherapy, or different molecules. Recently (September 2021), the FDA approved the use of cabozantinib for the treatment of patients (>12 years of age) with metastatic or locally advanced differentiated thyroid cancer (DTC), not amenable to receive iodine therapy, and who have failed different TKIs therapies. This approval was mainly achieved as a result of clinical findings of the COSMIC-311 study that observed prolonged progression-free survival in the group receiving the drug compared to the control (placebo) group.

Anti-EGFR therapy to treat pediatric malignancies has been less frequently evaluated in clinical trials. The Children's Oncology Group (COG) evaluated gefitinib, an oral EGFR tyrosine kinase inhibitor, in children with refractory solid tumors [923]. Twenty-five patients were enrolled, and although the drug was well tolerated, only one patient showed partial tumor response in this study cohort. Gefitinib was also evaluated in concomitance to radiotherapy in newly diagnosed children with brainstem gliomas (DIPGs). Forty-three eligible patients entered this study. Although the vast majority of patients experienced rapid and fatal tumor progression, three patients remained free of tumor progression for more than 36 months, pointing to a possible benefit of this approach for a small subset of DIPGs [924].

#### 5.1.2. RET Inhibitors

Recently, RET-altered tumors were considered amenable to receive targeted therapy with RET inhibitors in a tissue-agnostic manner [925]. Vandetanib, a multi-TKI including RET inhibition, was evaluated in association with bortezomib in 22 patients (17 evaluable cases) with medullary thyroid cancer, with 27% showing partial responses [926]. Additionally, selpercatinib was evaluated in 42 patients with RET-fused tumors of different histologies other than lung and thyroid; durable antitumor activity across different tumor subtypes was observed, with only minor adverse effects [927]. Tissue-agnostic benefits of the use of RET inhibitors in patients with RET-fused tumors of different histologies were also confirmed with different drugs, such as Pralsetinib [928].

#### 5.1.3. ALK Inhibitors

A consortium phase I study coordinated by the COG evaluated the use of crizotinib for childhood cancer with refractory solid tumors or anaplastic large-cell lymphomas (ALCL). Seventy-nine children (aged 6 years or older) and adolescents were enrolled; tumor responses were more pronounced among patients with tumors with activating ALK aberrations [929]. In addition, Fukano et al. investigated the role of alectinib in primary refractory ALCL, or after relapsing, in a phase II study that included both children and adults. Eight out of ten enrolled patients responded to alectinib, with minor adverse effects described [930].

Moreover, Entrectinib, a potent CNS-penetrant inhibitor of TRKA/B/C, ROS1 and ALK, was also evaluated to treat children and young adults with solid or primary CNS tumors harboring NTRK, ROS1 or ALK aberrations [730]. In this phase I/II trial, the objective response rate (ORR) was 57.7% among 43 response-evaluable patients. Entrectinib



shows a suitable safety profile and is effective as an option to treat pediatric patients with solid tumors harboring NTRK1/2/3 or ROS1 fusions.

### 5.2. PI3K/AKT/mTOR Pathway

The use of mTOR inhibitors, particularly everolimus, may be considered an important hallmark to treat children (>3 years old) with subependymal giant cell astrocytomas (SEGAs) associated with tuberous sclerosis complex (TSC) and not amenable to surgical treatment. This indication is largely derived from a phase III study (EXIST-1) that evaluated 117 patients in a double-blind placebo controlled trial, showing 50% tumor reduction exclusively in the treatment arm. The most frequent adverse events were oral ulcer and pyrexia; however, there was no treatment discontinuation due to adverse events [931].

### 5.3. MAPK Pathway

Abnormal, disrupted or constitutively activated MAPK pathways are involved in many pediatric cancers, particularly in low-grade gliomas (LGG). Recently, BRAF V600E-mutated tumors in children were eligible for agnostic treatment with BRAF plus MEK inhibitors [932]. Patients were randomized to receive either dabrafenib plus trametinib or standard chemotherapy. Among 110 treated children, complete and partial responses were reached in 47% of patients receiving the targeted therapy versus 11% for patients receiving chemotherapy alone. Of note, the clinical benefit rate, defined as complete, partial and stable disease lasting for more than 24 weeks, was 86% for trametinib plus dabrafenib versus 46% for standard chemotherapy [933]. Monotherapy with the BRAF inhibitor Dabrafenib was also previously evaluated in refractory or relapsed BRAF V600-mutated LGGs in children in a phase I/II study [934]. Among 32 enrolled patients (aged 1 to <18 years), the ORR was 44% and the 1-year PFS was 85%. In addition, adverse events (AE) were described in 91% of the participants; the most frequent AEs were fatigue, skin rash, dry skin and fever.

Disrupted MAPK pathways are also observed in patients with NF-1, where germline pathogenic neurofibromin mutations lead to the abrogation of the repressive function of this protein, with consequent activation of the PI3-K/AKT and RAS/MAPK cell signaling [935]. Besides the augmented frequency of LGG in patients with NF-1, plexiform neurofibromas are also frequently diagnosed in these patients, sometimes with life-threatening clinical consequences. Selumetinib, an MEK inhibitor, was evaluated in patients with symptomatic and inoperable plexiform neurofibromas [936]. Fifty children were enrolled in this phase II study that showed sustained tumor reduction, associated with clinical benefits. Improvements were observed in reducing pain and recovering motor function. However, 5 out of 50 patients discontinued treatment due to adverse effects possibly related to selumetinib, and 6 patients experienced disease progression while receiving the medication.

### 5.4. Cell Cycle Kinases

Ribociclib, an oral CDK4/6 inhibitor with pre-clinical evidence of action in different types of pediatric cancer, was tested either alone or in combination with chemotherapy in phase I and I/II studies, respectively [331,937]. Stable disease was the best response observed for both trials. The most common AEs were hematologic, including neutropenia, anemia and lymphopenia. More recently, palbociclib, a different oral CDK4/6 inhibitor, was evaluated in a phase I study directed at children and adolescents with progressive brain tumors. MTD of palbociclib was set at 75 mg/m<sup>2</sup> (as monotherapy) for 21 days, followed by 7 days without medication. Neutropenia and thrombocytopenia were common AEs; no objective responses were observed among 35 enrolled patients [938].

In addition, different aurora-kinase inhibitors (AKIs) have undergone clinical trials in pediatric cancer. Thirty-seven patients were enrolled in a phase I COG study evaluating MLN8237, a selective AKI-A [939]. Myelosuppression, mucositis and hand-foot skin syndrome were common side effects. One PR and six prolonged SD were observed. AT9283, a different multitarget of AKIs A and B, was evaluated to treat pediatric patients with

different types of solid tumors [940]. Of twenty-three evaluable patients, the authors described one confirmed PR and nine cases of disease stabilization after two courses of AT9283. More recently, Alisertib, a potent AKI-A, was evaluated in the pediatric population with both recurred/refractory solid tumors or leukemia (phase I). Five objective responses were reported, including two complete responses out of one-hundred and thirty-seven evaluable participants [941].

## 6. Final Remarks

Despite improvements, cancer is still responsible for 8% of all disease-related deaths in the pediatric setting [942]. Conventional chemotherapy not only is often ineffective but can also cause long-term complications that hamper the patient's quality of life.

Over the past two decades, precision oncology and the advent of innovator small-molecule drugs or immunotherapy have revolutionized the treatment of many adult cancers, such as CML, GBM and certain types of breast carcinomas [12,943–947]. The tidal increase in genomic, epigenomic, transcriptomic, proteomic and biochemical data has enormously improved our understanding of the specific molecular signatures of pediatric solid tumors, as well as allowing the sub-classification of some tumor types, as is the case of MB and EPN [948], and the application of corresponding specific therapies. Indeed, we are currently in a transition state where the broadly applied decades-old and not always conclusively curative cytotoxic drugs are being gradually substituted by targeted ones, and in the near future, molecular technology will steer diagnosis and personalized treatment [949].

With over 500 kinases in the human genome regulating key biological processes, many members of this molecular family have gained scientific limelight in oncology and academic pharma. Herein, we provided an in-depth review of published data on the roles of the dysregulation of a selected group of kinases in tumor pathophysiology and corroborated their importance as therapeutic candidates in the context of pediatric solid tumors.

By 2020, the FDA had approved 52 small-molecule therapeutics that target nearly 20 different protein kinases (half of them are multikinase inhibitors and the majority target RTKs) [950]. Other drugs targeting an additional 15–20 protein kinases remain in clinical trials worldwide. Nevertheless, a total of 40 target kinases represents only 10% of the kinase superfamily, and most of those kinase-directed drugs have not been tested in pediatric patients. Still, critical challenges must be overcome in experimental oncology and the translation into clinical options. These challenges include the following:

- (1) **Selectivity:** Most inhibitors developed so far target the ATP-binding site, meaning that they may act on multiple targets simultaneously and open new opportunities for the treatment of different tumor histologies [951,952]. Imatinib, for example, which has led to a significant increase in CML survival rates by selectively targeting the tumor-specific protein BCR/ABL, was included for the treatment of gastrointestinal stromal tumors (GIST), which are characterized by KIT-activating mutations [953]. Nevertheless, most inhibitors discovered to date have faced several adversities limiting their clinical use. First of all, the high sequence similarity in the ATP-binding sites frequently results in poor selectivity (refer to Figure 5 and Supplementary Figure S3) that may lead to undesired side effects. Moreover, these small molecules must compete with high intracellular ATP levels, leading to differences in potency when measured *in vivo* by biochemical versus cellular assays. In fact, many compounds inhibit their enzymes at nanomolar concentrations when measured biochemically, but only inhibit tumor cell growth under 3-fold higher concentrations [954]. Nevertheless, the increasing number of recognized kinase-specific structural features has allowed the emergence of superior non-ATP competitive kinase inhibitors that target other allosteric sites, which mostly act by inducing a conformational shift in the target enzyme, depleting its function [951,955–957].
- (2) **Adverse effects:** Imatinib and dasatinib, for instance, are both licensed for the treatment of children with CML. Despite its undeniable benefits, and with the spectrum of

side effects being comparable to what has been reported in adults (i.e., gastrointestinal toxicity, skin rash and muscle cramps), in a growing organism, imatinib treatment impairs longitudinal growth through the disturbance of osseous remodeling and inhibition of growth hormone secretion, which raises concerns about its lifelong use [958]. Moreover, despite the wealth of compounds that emerge on a daily basis, showing selectivity, potency and favorable pharmacological profiles, the probabilities for the translation into effective patient treatment for the great majority of them are extremely low. In this regard, less than 30% of the compounds approved or with clinical promise retrieved from the CanSAR platform have entered clinical trials in the pediatric setting (refer to Figure 6A). PLK1 inhibitors, for instance, despite the robust results obtained *in vitro* and *in vivo*, have demonstrated poor applicability due to severe hematological toxicity [959].

- (3) **Mutational burden and lack of predictive biomarkers:** As stated before, the mutational identity may vary between adult and pediatric cancer, a feature that reflects in treatment response. Current treatments targeting ALK mutations in other cancers, for example, have not shown significant efficacy against NB. In this tumor, two hotspot mutations, at positions R1275Q and F1174L, occur in a high proportion of patients; tumors harboring the first are highly sensitive to crizotinib, while tumors bearing the second are resistant [960–962]. Moreover, as suggested by Bellantoni and Wagner (2021), childhood solid tumors may have fewer potentially targetable mutations, evinced by the inhibition of RTK for the treatment of OS, where it is necessary to target several relevant RTKs simultaneously to achieve desirable results [821]. Therefore, ground-breaking drugs for adult cancer may not be effective in the pediatric setting. In parallel, inhibitors are not effective if the target is not essential to drive tumor growth or does not represent a prognostic factor, as is the case of ROCK kinases. Even though these proteins have gained popularity and progressively been researched as targets for the development of novel anti-cancer drugs due to their association with metastasis and poorer patient survival in adult tumors, the influence of both isoforms on the prognosis of childhood cancer remains controversial [963].
- (4) **Intrinsic and acquired resistance:** Resistance to targeted therapies is considered a largely inevitable hurdle that has a substantial impact on patients. Refractoriness to chemotherapy due to acquired F1174S ALK mutation in NB has been reported [964]. Likewise, the location of EGFR mutations significantly changes the effectiveness of EGFR; several mutations conferring resistance to EGFR tyrosine kinase inhibitors (such as T790M, L833V, A839T, V851I, A871T and G873E) have been reported [938]. Other examples include inadequate response to imatinib due to BCR-ABL1 kinase domain mutations that impart varying degrees of drug insensitivity, observed as underlying mechanism in 5–10% of adults and children with CML, bypassing pathway activation [965,966]. Moreover, despite an initial benefit of the targeted drug in molecularly well-defined tumors, patients inevitably experience tumor progression due to the development of resistance (i.e., Crizotinib in ALK-rearranged NSCLC population and CNS relapses) [967].
- (5) **Lack of compounds designed specifically for childhood tumors:** In general, few pediatric patients with cancer are enrolled in clinical trials. The perception that adult studies can be generalized to children with similar diseases is a major obstacle. Consequently, most treatments are based on modifications of previously approved regimes for the adult population, and many compounds enter clinical trials without preclinical testing in pediatric oncology (refer to Figure 6B), which is mandatory to obtain a more accurate interpretation of its possible therapeutic potential in a certain cancer entity [968]. Moreover, pediatric cancer is rare, and even among patients with the same cancer type, there is often broad heterogeneity in terms of prognosis, molecular features or pathology. Therefore, few institutions have sufficient patients and the chances of every potential agent or combination being tested are reduced. Even so, priorities for funding are typically assessed according to the “burden of

illness” for diseases, which is traditionally determined by disease frequency and mortality rate, leading to reluctance to distribute limited research funding to pediatric trials [969–971].

Regardless of the above challenges, kinase-based drug discovery has attained dramatic growth in the past 20 years. Although kinase inhibition represents a young therapeutic strategy compared with other traditional tactics, the FDA has approved a median of almost two small-molecule kinase inhibitors per year [952]. Thus, increasing numbers of targeted therapies are being tested for pediatric cancers, and many have shown undeniable success. Besides, the inhibition of kinases in normal cells can be clinically tolerated, presenting a therapeutic window that allows the softening of the acute side effects that generally lead to refusal and abandonment of treatment [972].

Moreover, as research advances, it has become clear that kinase inhibitors do not have to be absolutely selective. Crizotinib, for instance, was initially developed as an MET inhibitor, but later it was found to be even more efficient in cancers with ALK rearrangements. Additionally, molecularly targeted therapies are proving to be more effective in combination regimes to completely shut down the dysregulated pathway. As an example, it has been shown that everolimus improves CNS retention of vandetanib, dasatinib and sorafenib, which may have a great impact on the treatment of CNS tumors or brain metastases [751,752]. In the same vein, third- or fourth-generation inhibitors are being developed to avoid resistance and improve other biopharmaceutical properties such as brain penetration. These inhibitors, coupled with the increased ability to characterize tumors on molecular and genomic levels, will not only enable treatment refinement by identifying which patients may benefit most, but in the near future may conquer many diseases that are currently incurable.

**Supplementary Materials:** The following supporting information can be downloaded at: <https://www.mdpi.com/article/10.3390/pharmaceutics15020664/s1>, Figure S1: Flowchart depicting the identification of databases used in the present study, describing the number originally identified, included and excluded, and the reasons for exclusions; Figure S2: Flowchart representing kinases of interest analysis in cell line dependency score and the identification of inhibitors, preclinical studies and clinical test describing the number originally identified, included and excluded, and the reasons for exclusions; Figure S3: Schematic illustrations of kinases druggability identified by the CanSAR database in the other kinase families, including the total number of compounds with predicted interaction capacity with each kinase, as well as FDA-approved drugs, and clinical candidates. Interaction networks of kinase inhibitors and associated binding proteins according to STITCH (‘search tool for interactions of chemicals’). Compounds are represented as pill-shaped nodes, while proteins are shown as spheres. Small nodes represent proteins of unknown 3D structures, while large nodes show proteins with known or predicted structures. Nodes that are associated to each other are linked by an edge: thicker lines represent stronger binding affinities. Networks were constructed considering a minimum required interaction score of 0.700, and based on associations reported in Curated Databases (gray lines), or on both Databases and Experimental/Biochemical Data (green lines). Purple lines represent functional links between proteins; Table S1: Differential expression of kinases (pediatric tumors vs normal tissue); Table S2: Kinase expression and clinical features; Table S3: DepMap scores; Table S4: preclinical data; Table S5: Clinical trials.

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## References

1. Steliarova-Foucher, E.; Stiller, C.; Lacour, B.; Kaatsch, P. International Classification of Childhood Cancer, third edition. *Cancer* **2005**, *103*, 1457–1467. [[CrossRef](#)] [[PubMed](#)]
2. Instituto Nacional de Câncer (Brazil). *Coordenação de Prevenção e Vigilância and Sociedade Brasileira de Oncologia Pediátrica, Câncer na Criança e no Adolescente no Brasil: Dados dos Registros de Base Populacional e de Mortalidade*; Ministério da Saúde, Instituto Nacional de Câncer–INCA: Brasília, Brazil, 2008.
3. Downing, J.R.; Wilson, R.K.; Zhang, J.; Mardis, E.R.; Pui, C.-H.; Ding, L.; Ley, T.J.; E Evans, W. The Pediatric Cancer Genome Project. *Nat. Genet.* **2012**, *44*, 619–622. [[CrossRef](#)] [[PubMed](#)]
4. Toren, A.; Rechavi, G.; Ramot, B. Pediatric Cancer: Environmental and Genetic Aspects. *Pediatr. Hematol. Oncol.* **1996**, *13*, 319–331. [[CrossRef](#)] [[PubMed](#)]
5. Verma, V.; Denniston, K.A.; Lin, C.; Lin, C. A Comparison of Pediatric vs. Adult Patients with the Ewing Sarcoma Family of Tumors. *Front. Oncol.* **2017**, *7*, 82. [[CrossRef](#)]
6. Sultan, I.; Qaddoumi, I.; Yaser, S.; Rodriguez-Galindo, C.; Ferrari, A. Comparing Adult and Pediatric Rhabdomyosarcoma in the Surveillance, Epidemiology and End Results Program, 1973 to 2005: An Analysis of 2,600 Patients. *J. Clin. Oncol.* **2009**, *27*, 3391–3397. [[CrossRef](#)]
7. Spector, L.; Brown, M.B.; Wantman, E.; Letterie, G.S.; Toner, J.P.; Doody, K.; Ginsburg, E.; Williams, M.; Koch, L.; Schymura, M.J.; et al. Association of In Vitro Fertilization With Childhood Cancer in the United States. *JAMA Pediatr.* **2019**, *173*, e190392. [[CrossRef](#)]
8. Rahal, Z.; Abdulhai, F.; Kadara, H.; Saab, R. Genomics of adult and pediatric solid tumors. *Am. J. Cancer Res.* **2018**, *8*, 1356–1386.
9. Gröbner, S.N.; Worst, B.C.; Weischenfeldt, J.; Buchhalter, I.; Kleinheinz, K.; Rudneva, V.A.; Johann, P.D.; Balasubramanian, G.P.; Segura-Wang, M.; Brabetz, S.; et al. The landscape of genomic alterations across childhood cancers. *Nature* **2018**, *555*, 321–327. [[CrossRef](#)]
10. Sweet-Cordero, E.A.; Biegel, J.A. The genomic landscape of pediatric cancers: Implications for diagnosis and treatment. *Science* **2019**, *363*, 1170–1175. [[CrossRef](#)]
11. Vellichirammal, N.N.; Chaturvedi, N.K.; Joshi, S.S.; Coulter, D.W.; Guda, C. Fusion genes as biomarkers in pediatric cancers: A review of the current state and applicability in diagnostics and personalized therapy. *Cancer Lett.* **2020**, *499*, 24–38. [[CrossRef](#)]
12. The Cancer Genome Atlas Research Network. Comprehensive genomic characterization defines human glioblastoma genes and core pathways. *Nature* **2008**, *455*, 1061–1068. [[CrossRef](#)]
13. Schwartzentruber, J.; Korshunov, A.; Liu, X.-Y.; Jones, D.T.W.; Pfaff, E.; Jacob, K.; Sturm, D.; Fontebasso, A.M.; Khuong-Quang, D.-A.; Tönjes, M.; et al. Driver mutations in histone H3.3 and chromatin remodelling genes in paediatric glioblastoma. *Nature* **2012**, *482*, 226–231. [[CrossRef](#)]
14. Paugh, B.S.; Qu, C.; Jones, C.; Liu, Z.; Adamowicz-Brice, M.; Zhang, J.; Bax, D.A.; Coyle, B.; Barrow, J.; Hargrave, D.; et al. Integrated Molecular Genetic Profiling of Pediatric High-Grade Gliomas Reveals Key Differences With the Adult Disease. *J. Clin. Oncol.* **2010**, *28*, 3061–3068. [[CrossRef](#)]
15. Appay, R.; Fina, F.; Macagno, N.; Padovani, L.; Colin, C.; Baretts, D.; Ordioni, J.; Scavarda, D.; Giangaspero, F.; Badiali, M.; et al. Duplications of KIAA1549 and BRAF screening by Droplet Digital PCR from formalin-fixed paraffin-embedded DNA is an accurate alternative for KIAA1549-BRAF fusion detection in pilocytic astrocytomas. *Mod. Pathol.* **2018**, *31*, 1490–1501. [[CrossRef](#)]
16. Fukuoka, K.; on behalf of the Japan Pediatric Molecular Neuro-Oncology Group (JPMNG); Kanemura, Y.; Shofuda, T.; Fukushima, S.; Yamashita, S.; Narushima, D.; Kato, M.; Honda-Kitahara, M.; Ichikawa, H.; et al. Significance of molecular classification of ependymomas: C11orf95-RELA fusion-negative supratentorial ependymomas are a heterogeneous group of tumors. *Acta Neuropathol. Commun.* **2018**, *6*, 134. [[CrossRef](#)]
17. Wachtel, M.; Schäfer, B.W. PAX3-FOXO1: Zooming in on an “undruggable” target. *Semin. Cancer Biol.* **2018**, *50*, 115–123. [[CrossRef](#)]
18. Giovannini, M.; Biegel, J.A.; Serra, M.; Wang, J.Y.; Wei, Y.H.; Nycum, L.; Emanuel, B.S.; Evans, G.A. EWS-erg and EWS-Flil1 fusion transcripts in Ewing’s sarcoma and primitive neuroectodermal tumors with variant translocations. *J. Clin. Investig.* **1994**, *94*, 489–496. [[CrossRef](#)]
19. Jemal, A.; Murray, T.; Samuels, A.; Ghafoor, A.; Ward, E.; Thun, M.J. Cancer Statistics, 2003. *CA A Cancer J. Clin.* **2003**, *53*, 5–26. [[CrossRef](#)]
20. Pui, C.-H. Recent Research Advances in Childhood Acute Lymphoblastic Leukemia. *J. Formos. Med. Assoc.* **2010**, *109*, 777–787. [[CrossRef](#)]
21. Pezuk, J.A.; Valera, E.T.; Brassesco, M.S. PLK1 Inhibition: Prospective Role for the Treatment of Pediatric Tumors. *Curr. Drug Targets* **2016**, *17*, 1661–1672. [[CrossRef](#)]
22. Manning, G.; Whyte, D.B.; Martinez, R.; Hunter, T.; Sudarsanam, S. The Protein Kinase Complement of the Human Genome. *Science* **2002**, *298*, 1912–1934. [[CrossRef](#)] [[PubMed](#)]
23. Theivendren, P.; Kunjiappan, S.; Hegde, Y.M.; Vellaichamy, S.; Gopal, M.; Dhramalingam, S.R.; Kumar, S. Importance of Protein Kinase and Its Inhibitor: A Review. *Protein Kinases-Promis Targets Anticance. Drug Res. IntechOpen Ser. Biochem.* **2021**, *24*, 75–100. [[CrossRef](#)]

24. Alberts, B.; Johnson, A.; Lewis, J.; Raff, M.; Roberts, K.; Walter, P. *Bruce Alberts, Molecular Biology of the Cell*, 4th ed.; Garland Science: New York, NY, USA, 2002.
25. Turdo, A.; D'Accardo, C.; Glaviano, A.; Porcelli, G.; Colarossi, C.; Colarossi, L.; Mare, M.; Faldetta, N.; Modica, C.; Pistone, G.; et al. Targeting Phosphatases and Kinases: How to Checkmate Cancer. *Front. Cell Dev. Biol.* **2021**, *9*, 690306. [[CrossRef](#)] [[PubMed](#)]
26. Giamas, G.; Man, Y.L.; Hirner, H.; Bischof, J.; Kramer, K.; Khan, K.; Ahmed, S.S.L.; Stebbing, J.; Knippschild, U. Kinases as targets in the treatment of solid tumors. *Cell Signal.* **2010**, *22*, 984–1002. [[CrossRef](#)]
27. Armstrong, H.; Bording-Jorgensen, M.; Dijk, S.; Wine, E. The Complex Interplay between Chronic Inflammation, the Microbiome, and Cancer: Understanding Disease Progression and What We Can Do to Prevent It. *Cancers* **2018**, *10*, 83. [[CrossRef](#)]
28. Das, S.; Bhattacharya, B.; Das, B.; Sinha, B.; Jamatia, T.; Paul, K. Etiologic Role of Kinases in the Progression of Human Cancers and Its Targeting Strategies. *Indian J. Surg. Oncol.* **2019**, *12*, 34–45. [[CrossRef](#)]
29. McKay, M.M.; Morrison, D.K. Integrating signals from RTKs to ERK/MAPK. *Oncogene* **2007**, *26*, 3113–3121. [[CrossRef](#)]
30. Hubbard, S.R.; Miller, W.T. Receptor tyrosine kinases: Mechanisms of activation and signaling. *Curr. Opin. Cell Biol.* **2007**, *19*, 117–123. [[CrossRef](#)]
31. Lemmon, M.A.; Schlessinger, J. Cell Signaling by Receptor Tyrosine Kinases. *Cell* **2010**, *141*, 1117–1134. [[CrossRef](#)]
32. Ullrich, A.; Schlessinger, J. Signal transduction by receptors with tyrosine kinase activity. *Cell* **1990**, *61*, 203–212. [[CrossRef](#)]
33. Popovic, N.; Wilson, E. Cell Surface Receptors. In *Comprehensive Toxicology*; Elsevier: Amsterdam, The Netherlands, 2010; pp. 81–91. [[CrossRef](#)]
34. Paul, M.K.; Mukhopadhyay, A.K. Tyrosine kinase—Role and significance in Cancer. *Int. J. Med. Sci.* **2004**, *1*, 101–115. [[CrossRef](#)]
35. Schmidt-Arras, D.; Böhmer, F.-D. Mislocalisation of Activated Receptor Tyrosine Kinases—Challenges for Cancer Therapy. *Trends Mol. Med.* **2020**, *26*, 833–847. [[CrossRef](#)]
36. Bhargava, R.; Gerald, W.L.; Li, A.R.; Pan, Q.; Lal, P.; Ladanyi, M.; Chen, B. EGFR gene amplification in breast cancer: Correlation with epidermal growth factor receptor mRNA and protein expression and HER-2 status and absence of EGFR-activating mutations. *Mod. Pathol.* **2005**, *18*, 1027–1033. [[CrossRef](#)]
37. Drilon, A.; Cappuzzo, F.; Ou, S.-H.I.; Camidge, D.R. Targeting MET in Lung Cancer: Will Expectations Finally Be MET? *J. Thorac. Oncol.* **2017**, *12*, 15–26. [[CrossRef](#)]
38. Katoh, M. Fibroblast growth factor receptors as treatment targets in clinical oncology. *Nat. Rev. Clin. Oncol.* **2018**, *16*, 105–122. [[CrossRef](#)]
39. Helsten, T.; Elkin, S.; Arthur, E.; Tomson, B.N.; Carter, J.; Kurzrock, R. The FGFR Landscape in Cancer: Analysis of 4,853 Tumors by Next-Generation Sequencing. *Clin. Cancer Res.* **2016**, *22*, 259–267. [[CrossRef](#)]
40. Liu, G.; Chen, T.; Ding, Z.; Wang, Y.; Wei, Y.; Wei, X. Inhibition of FGF-FGFR and VEGF-VEGFR signalling in cancer treatment. *Cell Prolif.* **2021**, *54*, e13009. [[CrossRef](#)]
41. Ryall, S.; Tabori, U.; Hawkins, C. Pediatric low-grade glioma in the era of molecular diagnostics. *Acta Neuropathol. Commun.* **2020**, *8*, 30. [[CrossRef](#)]
42. Rivera, B.; Gayden, T.; Carrot-Zhang, J.; Nadaf, J.; Boshari, T.; Faury, D.; Zeinieh, M.; Blanc, R.; Burk, D.L.; Fahiminiya, S.; et al. Germline and somatic FGFR1 abnormalities in dysembryoplastic neuroepithelial tumors. *Acta Neuropathol.* **2016**, *131*, 847–863. [[CrossRef](#)]
43. Valera, E.T.; McConechy, M.K.; Gayden, T.; Rivera, B.; Jones, D.T.W.; Wittmann, A.; Han, H.; Bareke, E.; Nikbakht, H.; Mikael, L.; et al. Methylation analysis and whole-exome sequencing reveal that brain tumors associated with encephalocraniocutaneous lipomatoses are midline pilocytic astrocytomas. *Acta Neuropathol.* **2018**, *136*, 657–660. [[CrossRef](#)]
44. Amary, M.F.; Ye, H.; Berisha, F.; Khatri, B.; Forbes, G.; Lehovsky, K.; Frezza, A.M.; Behjati, S.; Tarpey, P.; Pillay, N.; et al. Fibroblastic growth factor receptor 1 amplification in osteosarcoma is associated with poor response to neo-adjuvant chemotherapy. *Cancer Med.* **2014**, *3*, 980–987. [[CrossRef](#)] [[PubMed](#)]
45. Kim, J.-A.; Berlow, N.E.; Lathara, M.; Bharathy, N.; Martin, L.R.; Purohit, R.; Cleary, M.M.; Liu, Q.; Michalek, J.E.; Srinivasa, G.; et al. Sensitization of osteosarcoma to irradiation by targeting nuclear FGFR1. *Biochem. Biophys. Res. Commun.* **2022**, *621*, 101–108. [[CrossRef](#)] [[PubMed](#)]
46. Ogura, K.; Elkrif, A.; Bowman, A.S.; Koche, R.P.; de Stanchina, E.; Benayed, R.; Mauguen, A.; Mattar, M.S.; Khodos, I.; Meyers, P.A.; et al. Prospective Clinical Genomic Profiling of Ewing Sarcoma: *ERF* and *FGFR1* Mutations as Recurrent Secondary Alterations of Potential Biologic and Therapeutic Relevance. *JCO Precis. Oncol.* **2022**, *6*, e2200048. [[CrossRef](#)] [[PubMed](#)]
47. Agelopoulos, K.; Richter, G.H.; Schmidt, E.; Dirksen, U.; von Heyking, K.; Moser, B.; Klein, H.-U.; Kontny, U.; Dugas, M.; Poos, K.; et al. Deep Sequencing in Conjunction with Expression and Functional Analyses Reveals Activation of FGFR1 in Ewing Sarcoma. *Clin. Cancer Res.* **2015**, *21*, 4935–4946. [[CrossRef](#)]
48. Rakheja, D.; Park, J.Y.; Yang, M.S.; Martinez, D.P.; Koduru, P.; Wilson, K.S.; Garcia, R.; Uddin, N. Rhabdomyosarcoma With Epithelioid Features And *NSD3::FOXO1* Fusion: Evidence For Reconsideration Of Previously Reported *FOXO1::FGFR1* Fusion. *Int. J. Surg. Pathol.* **2022**. [[CrossRef](#)]
49. Goldstein, M.; Meller, I.; Orr-Urtreger, A. FGFR1 over-expression in primary rhabdomyosarcoma tumors is associated with hypomethylation of a 5' CpG Island and abnormal expression of the AKT1, NOG, and BMP4 genes. *Genes Chromosom. Cancer* **2007**, *46*, 1028–1038. [[CrossRef](#)]

50. Gasparini, P.; Fortunato, O.; De Cecco, L.; Casanova, M.; Iannó, M.F.; Carenzo, A.; Centonze, G.; Milione, M.; Collini, P.; Boeri, M.; et al. Age-Related Alterations in Immune Contexture Are Associated with Aggressiveness in Rhabdomyosarcoma. *Cancers* **2019**, *11*, 1380. [[CrossRef](#)]
51. Missiaglia, E.; Selfe, J.; Pritchard-Jones, K.; Kool, M.; Shipley, J.; Hamdi, M.; Williamson, D.; Schaaf, G.; Fang, C.; Koster, J.; et al. Genomic imbalances in rhabdomyosarcoma cell lines affect expression of genes frequently altered in primary tumors: An approach to identify candidate genes involved in tumor development. *Genes Chromosom. Cancer* **2009**, *48*, 455–467. [[CrossRef](#)]
52. Lehtinen, B.; Raita, A.; Kesseli, J.; Annala, M.; Nordfors, K.; Yli-Harja, O.; Zhang, W.; Visakorpi, T.; Nykter, M.; Haapasalo, H.; et al. Clinical association analysis of ependymomas and pilocytic astrocytomas reveals elevated FGFR3 and FGFR1 expression in aggressive ependymomas. *BMC Cancer* **2017**, *17*, 310. [[CrossRef](#)]
53. Cimmino, F.; Montella, A.; Tirelli, M.; Avitabile, M.; Lasorsa, V.A.; Visconte, F.; Cantalupo, S.; Maiorino, T.; De Angelis, B.; Morini, M.; et al. FGFR1 is a potential therapeutic target in neuroblastoma. *Cancer Cell Int.* **2022**, *22*, 174. [[CrossRef](#)]
54. Schmelz, K.; Toedling, J.; Huska, M.; Cwikla, M.C.; Kruetzfeldt, L.-M.; Proba, J.; Ambros, P.F.; Ambros, I.M.; Boral, S.; Lodrini, M.; et al. Spatial and temporal intratumour heterogeneity has potential consequences for single biopsy-based neuroblastoma treatment decisions. *Nat. Commun.* **2021**, *12*, 6804. [[CrossRef](#)] [[PubMed](#)]
55. Nobusawa, S.; Hirato, J.; Yokoo, H. Molecular genetics of ependymomas and pediatric diffuse gliomas: A short review. *Brain Tumor Pathol.* **2014**, *31*, 229–233. [[CrossRef](#)] [[PubMed](#)]
56. Park, S.-H.; Won, J.; Kim, S.-I.; Lee, Y.; Park, C.-K.; Kim, S.-K.; Choi, S.-H. Molecular Testing of Brain Tumor. *J. Pathol. Transl. Med.* **2017**, *51*, 205–223. [[CrossRef](#)] [[PubMed](#)]
57. Georgiou, V.; Gkretsi, V. The role of fibroblast growth factors and their receptors in gliomas: The mutations involved. *Rev. Neurosci.* **2018**, *30*, 543–554. [[CrossRef](#)]
58. Vega, J.E.V.; Brat, D.J. Incorporating Advances in Molecular Pathology Into Brain Tumor Diagnostics. *Adv. Anat. Pathol.* **2018**, *25*, 143–171. [[CrossRef](#)]
59. Jimenez-Pascual, A.; Siebzehnruhl, F.A. Fibroblast Growth Factor Receptor Functions in Glioblastoma. *Cells* **2019**, *8*, 715. [[CrossRef](#)]
60. Ohashi, R.; Matsuda, Y.; Ishiwata, T.; Naito, Z. Downregulation of fibroblast growth factor receptor 2 and its isoforms correlates with a high proliferation rate and poor prognosis in high-grade glioma. *Oncol. Rep.* **2014**, *32*, 1163–1169. [[CrossRef](#)]
61. Yan, Y.; Li, Z.; Zeng, S.; Wang, X.; Gong, Z.; Xu, Z. FGFR2-mediated phosphorylation of PTEN at tyrosine 240 contributes to the radioresistance of glioma. *J. Cell Commun. Signal.* **2019**, *13*, 279–280. [[CrossRef](#)]
62. Salm, F.; Cwiek, P.; Ghosal, A.; Buccarello, A.L.; Largey, F.; Wotzkow, C.; Höland, K.; Styp-Rekowska, B.; Djonov, V.; Zlobec, I.; et al. RNA interference screening identifies a novel role for autocrine fibroblast growth factor signaling in neuroblastoma chemoresistance. *Oncogene* **2012**, *32*, 3944–3953. [[CrossRef](#)]
63. Kumar, K.S.; Neve, A.; Stucklin, A.S.G.; Kuzan-Fischer, C.M.; Rushing, E.J.; Taylor, M.D.; Tripolitsioti, D.; Behrmann, L.; Kirschenbaum, D.; Grotzer, M.; et al. TGF- $\beta$  Determines the Pro-migratory Potential of bFGF Signaling in Medulloblastoma. *Cell Rep.* **2018**, *23*, 3798–3812.e8. [[CrossRef](#)]
64. Vignovich, J.; Becker, D. Expression of BFGF and differential expression of FGF receptors in normal human myoblasts and rhabdomyosarcomas. *Int. J. Oncol.* **1993**, *2*, 637–642. [[CrossRef](#)]
65. Hirotsu, M.; Setoguchi, T.; Matsunoshita, Y.; Sasaki, H.; Nagao, H.; Gao, H.; Sugimura, K.; Komiya, S. Tumour formation by single fibroblast growth factor receptor 3-positive rhabdomyosarcoma-initiating cells. *Br. J. Cancer* **2009**, *101*, 2030–2037. [[CrossRef](#)]
66. Sahu, D.K.; Singh, N.; Das, M.; Rawat, J.; Gupta, D.K. Differential expression profiling of onco and tumor-suppressor genes from major-signaling pathways in Wilms' tumor. *Pediatr. Surg. Int.* **2022**, *38*, 1601–1617. [[CrossRef](#)]
67. Kostopoulou, O.N.; Holzhauser, S.; Lange, B.; Ohmayer, A.; Andonova, T.; Bersani, C.; Dalianis, T. Analyses of FGFR3 and PIK3CA mutations in neuroblastomas and the effects of the corresponding inhibitors on neuroblastoma cell lines. *Int. J. Oncol.* **2019**, *55*, 1372–1384. [[CrossRef](#)]
68. Ahrendsen, J.T.; Sinai, C.; Meredith, D.M.; Malinowski, S.W.; Cooney, T.M.; Bandopadhyay, P.; Ligon, K.L.; Alexandrescu, S. Molecular Alterations in Pediatric Low-Grade Gliomas That Led to Death. *J. Neuropathol. Exp. Neurol.* **2021**, *80*, 1052–1059. [[CrossRef](#)]
69. Johnson, A.; Severson, E.; Gay, L.; Vergilio, J.-A.; Elvin, J.; Suh, J.; Daniel, S.; Covert, M.; Frampton, G.M.; Hsu, S.; et al. Comprehensive Genomic Profiling of 282 Pediatric Low- and High-Grade Gliomas Reveals Genomic Drivers, Tumor Mutational Burden, and Hypermutation Signatures. *Oncologist* **2017**, *22*, 1478–1490. [[CrossRef](#)]
70. Frattini, V.; Pagnotta, S.M.; Tala, J.J.; Russo, M.V.; Lee, S.B.; Garofano, L.; Zhang, J.; Shi, P.; Lewis, G.; et al. A metabolic function of FGFR3-TACC3 gene fusions in cancer. *Nature* **2018**, *553*, 222–227. [[CrossRef](#)]
71. Kamura, S.; Matsumoto, Y.; Fukushi, J.-I.; Fujiwara, T.; Iida, K.; Okada, Y.; Iwamoto, Y. Basic fibroblast growth factor in the bone microenvironment enhances cell motility and invasion of Ewing's sarcoma family of tumours by activating the FGFR1-PI3K-Rac1 pathway. *Br. J. Cancer* **2010**, *103*, 370–381. [[CrossRef](#)]
72. Lee, S.; Lim, S.; Cho, D. Personalized genomic analysis based on circulating tumor cells of extra-skeletal Ewing sarcoma of the uterus: A case report of a 16-year-old Korean female. *Exp. Ther. Med.* **2018**, *16*, 1343–1349. [[CrossRef](#)]
73. Li, Z.; Dou, P.; Liu, T.; He, S. Application of Long Noncoding RNAs in Osteosarcoma: Biomarkers and Therapeutic Targets. *Cell. Physiol. Biochem.* **2017**, *42*, 1407–1419. [[CrossRef](#)]

74. Ren, T.; Qing, Y.; Dai, N.; Li, M.; Qian, C.; Yang, Y.; Cheng, Y.; Li, Z.; Zhang, S.; Zhong, Z.; et al. Apurinic/aprimidinic endonuclease 1 induced upregulation of fibroblast growth factor 2 and its receptor 3 induces angiogenesis in human osteosarcoma cells. *Cancer Sci.* **2014**, *105*, 186–194. [[CrossRef](#)] [[PubMed](#)]
75. Bi, Y.; Jing, Y.; Cao, Y. Overexpression of miR-100 inhibits growth of osteosarcoma through FGFR3. *Tumor Biol.* **2015**, *36*, 8405–8411. [[CrossRef](#)]
76. Gabler, L.; Jaunecker, C.N.; Katz, S.; van Schoonhoven, S.; Englinger, B.; Pirker, C.; Mohr, T.; Vician, P.; Stojanovic, M.; Woitzuck, V.; et al. Fibroblast growth factor receptor 4 promotes glioblastoma progression: A central role of integrin-mediated cell invasiveness. *Acta Neuropathol. Commun.* **2022**, *10*, 65. [[CrossRef](#)] [[PubMed](#)]
77. Hao, X.; Guo, Z.; Sun, H.; Liu, X.; Zhang, Y.; Zhang, L.; Sun, W.; Tian, Y. Urinary protein biomarkers for pediatric medulloblastoma. *J. Proteom.* **2020**, *225*, 103832. [[CrossRef](#)] [[PubMed](#)]
78. El Demellawy, D.; McGowan-Jordan, J.; de Nanassy, J.; Chernetsova, E.; Nasr, A. Update on molecular findings in rhabdomyosarcoma. *Pathology* **2017**, *49*, 238–246. [[CrossRef](#)] [[PubMed](#)]
79. Taylor Vi, J.G.T.; Cheuk, A.T.; Tsang, P.S.; Chung, J.-Y.; Song, Y.K.; Desai, K.; Yu, Y.; Chen, Q.-R.; Shah, K.; Youngblood, V.; et al. Identification of FGFR4-activating mutations in human rhabdomyosarcomas that promote metastasis in xenotransplanted models. *J. Clin. Investig.* **2009**, *119*, 3395–3407. [[CrossRef](#)]
80. Wesche, J.; Haglund, K.; Haugsten, E.M. Fibroblast growth factors and their receptors in cancer. *Biochem. J.* **2011**, *437*, 199–213. [[CrossRef](#)]
81. Baird, K.; Davis, S.; Antonescu, C.R.; Harper, U.L.; Walker, R.L.; Chen, Y.; Glatfelter, A.A.; Duray, P.H.; Meltzer, P.S. Gene Expression Profiling of Human Sarcomas: Insights into Sarcoma Biology. *Cancer Res* **2005**, *65*, 9226–9235. [[CrossRef](#)]
82. Cao, L.; Yu, Y.; Bilke, S.; Walker, R.L.; Mayeenuddin, L.H.; Azorsa, D.O.; Yang, F.; Pineda, M.; Helman, L.J.; Meltzer, P.S. Genome-Wide Identification of PAX3-FKHR Binding Sites in Rhabdomyosarcoma Reveals Candidate Target Genes Important for Development and Cancer. *Cancer Res* **2010**, *70*, 6497–6508. [[CrossRef](#)]
83. Shern, J.F.; Chen, L.; Chmielecki, J.; Wei, J.S.; Patidar, R.; Rosenberg, M.; Ambrogio, L.; Auclair, D.; Wang, J.; Song, Y.K.; et al. Comprehensive genomic analysis of rhabdomyosarcoma reveals a landscape of alterations affecting a common genetic axis in fusion-positive and fusion-negative tumors. *Cancer Discov.* **2014**, *4*, 216–231. [[CrossRef](#)]
84. Ramadan, F.; Fahs, A.; Ghayad, S.E.; Saab, R. Signaling pathways in Rhabdomyosarcoma invasion and metastasis. *Cancer Metastasis Rev.* **2020**, *39*, 287–301. [[CrossRef](#)]
85. Whittle, S.B.; Reyes, S.; Du, M.; Gireud-Goss, M.; Zhang, L.; Woodfield, S.E.; Ittmann, M.; Scheurer, M.; Bean, A.J.; Zage, P.E. A Polymorphism in the FGFR4 Gene Is Associated With Risk of Neuroblastoma and Altered Receptor Degradation. *J. Pediatr. Hematol.* **2016**, *38*, 131–138. [[CrossRef](#)]
86. Sugiyama, N.; Varjosalo, M.; Meller, P.; Lohi, J.; Chan, K.M.; Zhou, Z.; Alitalo, K.; Taipale, J.; Keski-Oja, J.; Lehti, K. FGF receptor-4 (FGFR4) polymorphism acts as an activity switch of a membrane type 1 matrix metalloproteinase–FGFR4 complex. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 15786–15791. [[CrossRef](#)]
87. Hajjo, R.; Sweidan, K. Review on Epidermal Growth Factor Receptor (EGFR) Structure, Signaling Pathways, Interactions, and Recent Updates of EGFR Inhibitors. *Curr. Top. Med. Chem.* **2020**, *20*, 815–834. [[CrossRef](#)]
88. Nicholson, R.; Gee, J.; Harper, M. EGFR and cancer prognosis. *Eur. J. Cancer* **2001**, *37*, 9–15. [[CrossRef](#)]
89. Spano, J.-P.; Lagorce, C.; Atlan, D.; Milano, G.; Domont, J.; Benamouzig, R.; Attar, A.; Benichou, J.; Martin, A.; Morere, J.-F.; et al. Impact of EGFR expression on colorectal cancer patient prognosis and survival. *Ann. Oncol.* **2005**, *16*, 102–108. [[CrossRef](#)]
90. Zarghooni, M.; Bartels, U.; Lee, E.; Buczkowicz, P.; Morrison, A.; Huang, A.; Bouffet, E.; Hawkins, C. Whole-Genome Profiling of Pediatric Diffuse Intrinsic Pontine Gliomas Highlights Platelet-Derived Growth Factor Receptor  $\alpha$  and Poly (ADP-ribose) Polymerase As Potential Therapeutic Targets. *J. Clin. Oncol.* **2010**, *28*, 1337–1344. [[CrossRef](#)]
91. Suri, V.; Das, P.; Jain, A.; Sharma, M.C.; Borkar, S.A.; Suri, A.; Gupta, D.; Sarkar, C. Pediatric glioblastomas: A histopathological and molecular genetic study. *Neuro-Oncology* **2009**, *11*, 274–280. [[CrossRef](#)]
92. Wong, A.J.; Bigner, S.H.; Bigner, D.D.; Kinzler, K.W.; Hamilton, S.R.; Vogelstein, B. Increased expression of the epidermal growth factor receptor gene in malignant gliomas is invariably associated with gene amplification. *Proc. Natl. Acad. Sci. USA* **1987**, *84*, 6899–6903. [[CrossRef](#)]
93. Kraus, J.A.; Felsberg, J.; Tonn, J.C.; Reifenberger, G.; Pietsch, T. Molecular genetic analysis of the TP53, PTEN, CDKN2A, EGFR, CDK4 and MDM2 tumour-associated genes in supratentorial primitive neuroectodermal tumours and glioblastomas of childhood. *Neuropathol. Appl. Neurobiol.* **2002**, *28*, 325–333. [[CrossRef](#)]
94. Pollack, I.F.; Hamilton, R.L.; James, C.D.; Finkelstein, S.D.; Burnham, J.; Yates, A.J.; Holmes, E.J.; Zhou, T.; Finlay, J.L. Rarity of PTEN deletions and EGFR amplification in malignant gliomas of childhood: Results from the Children’s Cancer Group 945 cohort. *J. Neurosurgery Pediatr.* **2006**, *105*, 418–424. [[CrossRef](#)] [[PubMed](#)]
95. Hatanpaa, K.J.; Burma, S.; Zhao, D.; Habib, A.A. Epidermal Growth Factor Receptor in Glioma: Signal Transduction, Neuropathology, Imaging, and Radioresistance. *Neoplasia* **2010**, *12*, 675–684. [[CrossRef](#)] [[PubMed](#)]
96. Ganigi, P.; Santosh, V.; Anandh, B.; Chandramouli, B.; Kolluri, V.S. Expression of p53, EGFR, pRb and bcl-2 Proteins in Pediatric Glioblastoma Multiforme: A Study of 54 Patients. *Pediatr. Neurosurg.* **2005**, *41*, 292–299. [[CrossRef](#)] [[PubMed](#)]
97. Gilbertson, R.J.; Hill, D.A. ERBB1 is amplified and overexpressed in high-grade diffusely infiltrative pediatric brain stem glioma. *Clin. Cancer Res.* **2003**, *9 Pt 1*, 3620–3624. [[PubMed](#)]



98. Korshunov, A.; Golanov, A.; Timirgazi, V. Immunohistochemical markers for intracranial ependymoma recurrence: An analysis of 88 cases. *J. Neurol. Sci.* **2000**, *177*, 72–82. [[CrossRef](#)] [[PubMed](#)]
99. Mendrzyk, F.; Korshunov, A.; Benner, A.; Toedt, G.; Pfister, S.; Radlwimmer, B.; Lichter, P. Identification of Gains on 1q and Epidermal Growth Factor Receptor Overexpression as Independent Prognostic Markers in Intracranial Ependymoma. *Clin. Cancer Res.* **2006**, *12*, 2070–2079. [[CrossRef](#)]
100. Massimino, M.; Buttarelli, F.R.; Antonelli, M.; Gandola, L.; Modena, P.; Giangaspero, F. Intracranial ependymoma: Factors affecting outcome. *Futur. Oncol.* **2009**, *5*, 207–216. [[CrossRef](#)]
101. Gilbertson, R.J.; Perry, R.H.; Kelly, P.J.; Pearson, A.D.; Lunec, J. Prognostic significance of HER2 and HER4 coexpression in childhood medulloblastoma. *Cancer Res.* **1997**, *57*, 3272–3280.
102. Bodey, B.; E Kaiser, H.; E Siegel, S. Epidermal growth factor receptor (EGFR) expression in childhood brain tumors. *Vivo* **2005**, *19*, 931–941.
103. Layfield, L.J.; Thompson, J.K.; Ms, R.K.D.; Kerns, B.-J. Prognostic indicators for neuroblastoma: Stage, grade, DNA ploidy, MIB-1-proliferation index, p53, HER-2/neu and EGFR—a survival study. *J. Surg. Oncol.* **1995**, *59*, 21–27. [[CrossRef](#)]
104. Izycka-Swieszewska, E.; Wozniak, A.; Drozyska, E.; Kot, J.; Grajkowska, W.; Klepacka, T.; Perek, D.; Koltan, S.; Bien, E.; Limon, J. Expression and significance of HER family receptors in neuroblastic tumors. *Clin. Exp. Metastasis* **2011**, *28*, 271–282. [[CrossRef](#)]
105. Izycka-Swieszewska, E.; Wozniak, A.; Kot, J.; Grajkowska, W.; Balcerska, A.; Perek, D.; Dembowska-Baginska, B.; Klepacka, T.; Drozyska, E. Prognostic significance of HER2 expression in neuroblastic tumors. *Mod. Pathol.* **2010**, *23*, 1261–1268. [[CrossRef](#)]
106. Wang, H.; Yang, Q.; Fu, Z.; Zuo, D.; Hua, Y.; Cai, Z. ErbB Receptors as Prognostic and Therapeutic Drug Targets in Bone and Soft Tissue Sarcomas. *Cancer Investig.* **2014**, *32*, 533–542. [[CrossRef](#)]
107. Wen, Y.H.; Koeppen, H.; Garcia, R.; Chiriboga, L.; Tarlow, B.D.; Peters, B.A.; Eigenbrot, C.; Yee, H.; Steiner, G.; Greco, M.A. Epidermal growth factor receptor in osteosarcoma: Expression and mutational analysis. *Hum. Pathol.* **2007**, *38*, 1184–1191. [[CrossRef](#)]
108. Liu, L.; Xiao, C.; Sun, Q. MiRNA-375 inhibits retinoblastoma progression through targeting ERBB2 and inhibiting MAPK1/MAPK3 signalling pathway. *Cutan. Ocul. Toxicol.* **2021**, *41*, 1–10. [[CrossRef](#)]
109. Vasei, M.; Modjtahedi, H.; Ale-Booyeh, O.; Mosallaei, A.; Kajbafzadeh, A.M.; Shahriari, M.; Ghaderi, A.A.; Soleymannpour, H.; Kosari, F.; Moch, H.; et al. Amplification and expression of EGFR and ERBB2 in Wilms tumor. *Cancer Genet. Cytogenet.* **2009**, *194*, 88–95. [[CrossRef](#)]
110. Little, S.E.; Bax, D.A.; Rodriguez-Pinilla, M.; Natrajan, R.; Messahel, B.; Pritchard-Jones, K.; Vujanic, G.M.; Reis-Filho, J.S.; Jones, C. Multifaceted Dysregulation of the Epidermal Growth Factor Receptor Pathway in Clear Cell Sarcoma of the Kidney. *Clin. Cancer Res.* **2007**, *13*, 4360–4364. [[CrossRef](#)]
111. Armistead, P.M.; Salganick, J.; Roh, J.S.; Steinert, D.M.; Patel, S.; Munsell, M.; El-Naggar, A.K.; Benjamin, R.S.; Zhang, W.; Trent, J.C. Expression of receptor tyrosine kinases and apoptotic molecules in rhabdomyosarcoma: Correlation with overall survival in 105 patients. *Cancer* **2007**, *110*, 2293–2303. [[CrossRef](#)]
112. Shibuya, M. Vascular endothelial growth factor and its receptor system: Physiological functions in angiogenesis and pathological roles in various diseases. *J. Biochem.* **2012**, *153*, 13–19. [[CrossRef](#)]
113. Risau, W. Mechanisms of angiogenesis. *Nature* **1997**, *386*, 671–674. [[CrossRef](#)]
114. Simons, M.; Gordon, E.; Claesson-Welsh, L. Mechanisms and regulation of endothelial VEGF receptor signalling. *Nat. Rev. Mol. Cell Biol.* **2016**, *17*, 611–625. [[CrossRef](#)] [[PubMed](#)]
115. Tamura, R.; Sato, M.; Morimoto, Y.; Ohara, K.; Kosugi, K.; Oishi, Y.; Kuranari, Y.; Murase, M.; Yoshida, K.; Toda, M. Quantitative assessment and clinical relevance of VEGFRs-positive tumor cells in refractory brain tumors. *Exp. Mol. Pathol.* **2020**, *114*, 104408. [[CrossRef](#)] [[PubMed](#)]
116. Slongo, M.L.; Molena, B.; Brunati, A.M.; Frasson, M.; Gardiman, M.; Carli, M.; Perilongo, G.; Rosolen, A.; Onisto, M. Functional VEGF and VEGF receptors are expressed in human medulloblastomas. *Neuro-Oncology* **2007**, *9*, 384–392. [[CrossRef](#)] [[PubMed](#)]
117. Gesundheit, B.; Klement, G.; Senger, C.; Kerbel, R.; Kieran, M.; Baruchel, S.; Becker, L. Differences in vasculature between pilocytic and anaplastic astrocytomas of childhood. *Med. Pediatr. Oncol.* **2003**, *41*, 516–526. [[CrossRef](#)]
118. Farschtschi, S.; Merker, V.L.; Wolf, D.; Schuhmann, M.; Blakeley, J.; Plotkin, S.R.; Hagel, C.; Mautner, V.F. Bevacizumab treatment for symptomatic spinal ependymomas in neurofibromatosis type 2. *Acta Neurol. Scand.* **2015**, *133*, 475–480. [[CrossRef](#)]
119. Virág, J.; Kenessey, I.; Haberler, C.; Piurko, V.; Bálint, K.; Döme, B.; Tímár, J.; Garami, M.; Hegedűs, B. Angiogenesis and Angiogenic Tyrosine Kinase Receptor Expression in Pediatric Brain Tumors. *Pathol. Oncol. Res.* **2013**, *20*, 417–426. [[CrossRef](#)]
120. Zhou, Q.; Yan, X.; Zhu, H.; Xin, Z.; Zhao, J.; Shen, W.; Yin, W.; Guo, Y.; Xu, H.; Zhao, M.; et al. Identification of three tumor antigens and immune subtypes for mRNA vaccine development in diffuse glioma. *Theranostics* **2021**, *11*, 9775–9790. [[CrossRef](#)]
121. Fakhari, M.; Pullirsch, D.; Abraham, D.; Paya, K.; Hofbauer, R.; Holzfeind, P.; Hofmann, M.; Aharinejad, S. Selective upregulation of vascular endothelial growth factor receptors neuropilin-1 and -2 in human neuroblastoma. *Cancer* **2001**, *94*, 258–263. [[CrossRef](#)]
122. Czapiewski, P.; Kunc, M.; Haybaeck, J. Genetic and molecular alterations in olfactory neuroblastoma: Implications for pathogenesis, prognosis and treatment. *Oncotarget* **2016**, *7*, 52584–52596. [[CrossRef](#)]
123. Behjati, S.; Tarpey, P.S.; Haase, K.; Ye, H.; Young, M.D.; Alexandrov, L.B.; Farnon, S.J.; Collord, G.; Wedge, D.C.; Martincorena, I.; et al. Recurrent mutation of IGF signalling genes and distinct patterns of genomic rearrangement in osteosarcoma. *Nat. Commun.* **2017**, *8*, 15936. [[CrossRef](#)]

124. Joseph, C.G.; Hwang, H.; Jiao, Y.; Wood, L.D.; Kinde, I.; Wu, J.; Mandahl, N.; Luo, J.; Hruban, R.H.; Diaz, L.; et al. Exomic analysis of myxoid liposarcomas, synovial sarcomas, and osteosarcomas. *Genes Chromosom. Cancer* **2013**, *53*, 15–24. [[CrossRef](#)]
125. Perry, J.A.; Kiezun, A.; Tonzi, P.; Van Allen, E.M.; Carter, S.L.; Baca, S.C.; Cowley, G.S.; Bhatt, A.S.; Rheinbay, E.; Pedamallu, C.S.; et al. Complementary genomic approaches highlight the PI3K/mTOR pathway as a common vulnerability in osteosarcoma. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, E5564–E5573. [[CrossRef](#)]
126. Negri, G.L.; Grande, B.M.; Delaidelli, A.; El-Naggar, A.; Cochrane, D.; Lau, C.C.; Triche, T.J.; Moore, R.A.; Jones, S.J.M.; Montpetit, A.; et al. Integrative genomic analysis of matched primary and metastatic pediatric osteosarcoma. *J. Pathol.* **2019**, *249*, 319–331. [[CrossRef](#)]
127. Subbiah, V.; Cote, G.J. Advances in Targeting RET-Dependent Cancers. *Cancer Discov.* **2020**, *10*, 498–505. [[CrossRef](#)]
128. Takahashi, M.; Kawai, K.; Asai, N. Roles of the RET Proto-oncogene in Cancer and Development. *JMA J.* **2020**, *3*, 175–181. [[CrossRef](#)]
129. Li, A.Y.; McCusker, M.G.; Russo, A.; Scilla, K.A.; Gittens, A.; Arensmeyer, K.; Mehra, R.; Adamo, V.; Rolfo, C. RET fusions in solid tumors. *Cancer Treat. Rev.* **2019**, *81*, 101911. [[CrossRef](#)]
130. Adashek, J.J.; Desai, A.P.; Andreev-Drakhlina, A.Y.; Roszik, J.; Cote, G.J.; Subbiah, V. Hallmarks of RET and Co-occurring Genomic Alterations in RET-aberrant Cancers. *Mol. Cancer Ther.* **2021**, *20*, 1769–1776. [[CrossRef](#)]
131. Elisei, R.; Romei, C.; Vorontsova, T.; Cosci, B.; Veremeychik, V.; Kuchinskaya, E.; Basolo, F.; Demidchik, E.P.; Miccoli, P.; Pinchera, A.; et al. RET/PTC Rearrangements in Thyroid Nodules: Studies in Irradiated and Not Irradiated, Malignant and Benign Thyroid Lesions in Children and Adults. *J. Clin. Endocrinol. Metab.* **2001**, *86*, 3211–3216. [[CrossRef](#)]
132. Rabes, H.M.; Klugbauer, S. Molecular genetics of childhood papillary thyroid carcinomas after irradiation: High prevalence of RET rearrangement. *Genes Environ. Cancer* **1998**, *154*, 248–264. [[CrossRef](#)]
133. Zimmerman, D. Thyroid neoplasia in children. *Curr. Opin. Pediatr.* **1997**, *9*, 413–418. [[CrossRef](#)]
134. Ortiz, M.V.; Gerdemann, U.; Raju, S.G.; Henry, D.; Smith, S.; Rothenberg, S.M.; Cox, M.C.; Proust, S.; Bender, J.G.; Frazier, A.L.; et al. Activity of the Highly Specific RET Inhibitor Selpercatinib (LOXO-292) in Pediatric Patients With Tumors Harboring RET Gene Alterations. *JCO Precis. Oncol.* **2020**, *4*, 341–347. [[CrossRef](#)] [[PubMed](#)]
135. Kovac, M.; Woolley, C.; Ribic, S.; Blattmann, C.; Roth, E.; Morini, M.; Kovacova, M.; Ameline, B.; Kulozik, A.; Bielack, S.; et al. Germline RET variants underlie a subset of paediatric osteosarcoma. *J. Med. Genet.* **2020**, *58*, 20–24. [[CrossRef](#)] [[PubMed](#)]
136. Greenfield, E.M.; Collier, C.D.; Getty, P.J. Receptor Tyrosine Kinases in Osteosarcoma: 2019 Update. *Adv. Exp. Med. Biol.* **2020**, *1258*, 141–155. [[CrossRef](#)] [[PubMed](#)]
137. Luo, J.; Xia, Y.; Yin, Y.; Luo, J.; Liu, M.; Zhang, C.; Zhao, Y.; Yang, L.; Kong, L. ATF4 destabilizes RET through nonclassical GRP78 inhibition to enhance chemosensitivity to bortezomib in human osteosarcoma. *Theranostics* **2019**, *9*, 6334–6353. [[CrossRef](#)] [[PubMed](#)]
138. Rettew, A.N.; Young, E.D.; Lev, D.C.; Kleinerman, E.S.; Abdulkarim, F.W.; Getty, P.J.; Greenfield, E.M. Multiple receptor tyrosine kinases promote the in vitro phenotype of metastatic human osteosarcoma cell lines. *Oncogenesis* **2012**, *1*, e34. [[CrossRef](#)]
139. Rettew, A.N.; Getty, P.J.; Greenfield, E.M. Receptor Tyrosine Kinases in Osteosarcoma: Not Just the Usual Suspects. *Curr. Adv. Osteosarcoma* **2014**, *804*, 47–66. [[CrossRef](#)]
140. Dabir, S.; Babakoohi, S.; Kluge, A.; Morrow, J.J.; Kresak, A.; Yang, M.; MacPherson, D.; Wildey, G.; Dowlati, A. RET Mutation and Expression in Small-Cell Lung Cancer. *J. Thorac. Oncol.* **2014**, *9*, 1316–1323. [[CrossRef](#)]
141. Cockburn, J.G.; Richardson, D.S.; Gujral, T.S.; Mulligan, L.M. RET-Mediated Cell Adhesion and Migration Require Multiple Integrin Subunits. *J. Clin. Endocrinol. Metab.* **2010**, *95*, E342–E346. [[CrossRef](#)]
142. Trusolino, L.; Bertotti, A.; Comoglio, P.M. MET signalling: Principles and functions in development, organ regeneration and cancer. *Nat. Rev. Mol. Cell Biol.* **2010**, *11*, 834–848. [[CrossRef](#)]
143. Park, K.C.; Richardson, D.R. The c-MET oncoprotein: Function, mechanisms of degradation and its targeting by novel anti-cancer agents. *Biochim. Biophys. Acta (BBA)-Gen. Subj.* **2020**, *1864*, 129650. [[CrossRef](#)]
144. Marona, P.; Górka, J.; Kotlinowski, J.; Majka, M.; Jura, J.; Miekus, K. C-Met as a Key Factor Responsible for Sustaining Undifferentiated Phenotype and Therapy Resistance in Renal Carcinomas. *Cells* **2019**, *8*, 272. [[CrossRef](#)]
145. Zambelli, A.; Biamonti, G.; Amato, A. HGF/c-Met Signalling in the Tumor Microenvironment. *TumorMicroenviron. Signal. Pathw. Part B* **2020**, *1270*, 31–44. [[CrossRef](#)]
146. Cooper, C.S.; Park, M.; Blair, D.G.; Tainsky, M.A.; Huebner, K.; Croce, C.M.; Vande Woude, G.F. Molecular cloning of a new transforming gene from a chemically transformed human cell line. *Nature* **1984**, *311*, 29–33. [[CrossRef](#)]
147. Grundy, M.; Narendran, A. The hepatocyte growth factor/mesenchymal epithelial transition factor axis in high-risk pediatric solid tumors and the anti-tumor activity of targeted therapeutic agents. *Front. Pediatr.* **2022**, *10*, 910268. [[CrossRef](#)]
148. Stucklin, A.S.G.; Ryall, S.; Fukuoka, K.; Zapotocky, M.; Lassaletta, A.; Li, C.; Bridge, T.; Kim, B.; Arnoldo, A.; Kowalski, P.E.; et al. Alterations in ALK/ROS1/NTRK/MET drive a group of infantile hemispheric gliomas. *Nat. Commun.* **2019**, *10*, 4343. [[CrossRef](#)]
149. Kunkel, P.; Müller, S.; Schirmacher, P.; Stavrou, D.; Fillbrandt, R.; Westphal, M.; Lamszus, K. Expression and localization of scatter factor/hepatocyte growth factor in human astrocytomas. *Neuro-Oncology* **2001**, *3*, 82–88. [[CrossRef](#)]
150. Li, Y.; Lal, B.; Kwon, S.; Fan, X.; Saldanha, U.; Reznik, T.E.; Kuchner, E.B.; Eberhart, C.; Laterra, J.; Abounader, R. The Scatter Factor/Hepatocyte Growth Factor: C-Met Pathway in Human Embryonal Central Nervous System Tumor Malignancy. *Cancer Res* **2005**, *65*, 9355–9362. [[CrossRef](#)]

151. Provençal, M.; Labbé, D.; Veitch, R.; Boivin, D.; Rivard, G.; Sartelet, H.; Robitaille, Y.; Gingras, D.; Béliveau, R. c-Met activation in medulloblastoma induces tissue factor expression and activity: Effects on cell migration. *Carcinog.* **2009**, *30*, 1089–1096. [[CrossRef](#)]
152. E Crosswell, H.; Dasgupta, A.; Alvarado, C.S.; Watt, T.; Christensen, J.G.; De, P.; Durden, D.L.; Findley, H.W. PHA665752, a small-molecule inhibitor of c-Met, inhibits hepatocyte growth factor-stimulated migration and proliferation of c-Met-positive neuroblastoma cells. *BMC Cancer* **2009**, *9*, 411. [[CrossRef](#)]
153. Hecht, M.; Papoutsi, M.; Tran, H.D.; Wilting, J.; Schweigerer, L. Hepatocyte Growth Factor/c-Met Signaling Promotes the Progression of Experimental Human Neuroblastomas. *Cancer Res* **2004**, *64*, 6109–6118. [[CrossRef](#)]
154. Alami, J.; Williams, B.R.G.; Yeager, H. Expression and localization of HGF and met in Wilms' tumours. *J. Pathol.* **2001**, *196*, 76–84. [[CrossRef](#)]
155. Cao, X.; Liu, D.-H.; Zhou, Y.; Yan, X.-M.; Yuan, L.-Q.; Pan, J.; Fu, M.-C.; Zhang, T.; Wang, J. Histone deacetylase 5 promotes Wilms' tumor cell proliferation through the upregulation of c-Met. *Mol. Med. Rep.* **2016**, *13*, 2745–2750. [[CrossRef](#)]
156. Nair, R.M.; Prabhu, V.; Manukonda, R.; Mishra, D.K.; Kaliki, S.; Vemuganti, G.K. Overexpression of metastasis-associated in colon cancer 1 in retinoblastoma. *Tumor Biol.* **2020**, *42*, 1010428320975973. [[CrossRef](#)]
157. Patanè, S.; Avnet, S.; Coltella, N.; Costa, B.; Sponza, S.; Olivero, M.; Vigna, E.; Naldini, L.; Baldini, N.; Ferracini, R.; et al. MET Overexpression Turns Human Primary Osteoblasts into Osteosarcomas. *Cancer Res* **2006**, *66*, 4750–4757. [[CrossRef](#)]
158. Chen, W.; Wu, S.; Huang, Y.; Zhang, T.; Dong, H.; Zheng, X.; Chen, T.; Gong, X.; Liu, G.; Zhao, X. A c-Met Inhibitor Suppresses Osteosarcoma Progression via the ERK1/2 Pathway in Human Osteosarcoma Cells. *Oncotargets Ther.* **2021**, *14*, 4791–4804. [[CrossRef](#)]
159. Entz-Werle, N.; Lavaux, T.; Metzger, N.; Stoetzel, C.; Lasthaus, C.; Marec, P.; Kalita, C.; Brugieres, L.; Pacquement, H.; Schmitt, C.; et al. Involvement of MET/TWIST/APC Combination or the Potential Role of Ossification Factors in Pediatric High-Grade Osteosarcoma Oncogenesis. *Neoplasia* **2007**, *9*, 678–688. [[CrossRef](#)]
160. Fleuren, E.D.; Roeffen, M.H.; Leenders, W.P.; Flucke, U.E.; Vletterie, M.; Schreuder, H.W.; Boerman, O.C.; van der Graaf, W.T.; Versleijen-Jonkers, Y.M. Expression and clinical relevance of MET and ALK in Ewing sarcomas. *Int. J. Cancer* **2013**, *133*, 427–436. [[CrossRef](#)]
161. Yan, D.; Da Dong, X.; Chen, X.; Wang, L.; Lu, C.; Wang, J.; Qu, J.; Tu, L. MicroRNA-1/206 Targets c-Met and Inhibits Rhabdomyosarcoma Development. *J. Biol. Chem.* **2009**, *284*, 29596–29604. [[CrossRef](#)]
162. Du, J.; Wang, Y.; Meng, L.; Liu, Y.; Pang, Y.; Cui, W.; Zhang, L.; Li, Z.; Liu, Q.; Shang, H.; et al. c-MET expression potentially contributes to the poor prognosis of rhabdomyosarcoma. *Int. J. Clin. Exp. Pathol.* **2018**, *11*, 4083–4092.
163. Otabe, O.; Kikuchi, K.; Tsuchiya, K.; Katsumi, Y.; Yagyū, S.; Miyachi, M.; Iehara, T.; Hosoi, H. MET/ERK2 pathway regulates the motility of human alveolar rhabdomyosarcoma cells. *Oncol. Rep.* **2016**, *37*, 98–104. [[CrossRef](#)]
164. Taulli, R.; Scuoppo, C.; Bersani, F.; Accornero, P.; Forni, P.E.; Miretti, S.; Grinza, A.; Allegra, P.; Schmitt-Ney, M.; Crepaldi, T.; et al. Validation of Met as a Therapeutic Target in Alveolar and Embryonal Rhabdomyosarcoma. *Cancer Res* **2006**, *66*, 4742–4749. [[CrossRef](#)]
165. Morris, S.W.; Kirstein, M.N.; Valentine, M.B.; Dittmer, K.G.; Shapiro, D.N.; Saltman, D.L.; Look, A.T. Fusion of a Kinase Gene, ALK, to a Nucleolar Protein Gene, NPM, in Non-Hodgkin's Lymphoma. *Science* **1994**, *263*, 1281–1284. [[CrossRef](#)]
166. Hallberg, B.; Palmer, R. The role of the ALK receptor in cancer biology. *Ann. Oncol.* **2016**, *27*, iii4–iii15. [[CrossRef](#)]
167. Takita, J. The role of anaplastic lymphoma kinase in pediatric cancers. *Cancer Sci.* **2017**, *108*, 1913–1920. [[CrossRef](#)]
168. Peron, M.; Lovisa, F.; Poli, E.; Basso, G.; Bonvini, P. Understanding the Interplay between Expression, Mutation and Activity of ALK Receptor in Rhabdomyosarcoma Cells for Clinical Application of Small-Molecule Inhibitors. *PLoS ONE* **2015**, *10*, e0132330. [[CrossRef](#)]
169. Felkai, L.; Bánusz, R.; Kovalszky, I.; Sápi, Z.; Garami, M.; Papp, G.; Karászi, K.; Varga, E.; Csóka, M. The Presence of ALK Alterations and Clinical Relevance of Crizotinib Treatment in Pediatric Solid Tumors. *Pathol. Oncol. Res.* **2017**, *25*, 217–224. [[CrossRef](#)]
170. Aygun, N. Biological and Genetic Features of Neuroblastoma and Their Clinical Importance. *Curr. Pediatr. Rev.* **2018**, *14*, 73–90. [[CrossRef](#)]
171. Pastor, E.R.; Mousa, S.A. Current management of neuroblastoma and future direction. *Crit. Rev. Oncol.* **2019**, *138*, 38–43. [[CrossRef](#)]
172. Pacenta, H.L.; E Macy, M. Entrectinib and other ALK/TRK inhibitors for the treatment of neuroblastoma. *Drug Des. Dev. Ther.* **2018**, *12*, 3549–3561. [[CrossRef](#)]
173. Berry, T.; Luther, W.; Bhatnagar, N.; Jamin, Y.; Poon, E.; Sanda, T.; Pei, D.; Sharma, B.; Vetharoy, W.R.; Hallsworth, A.; et al. The ALKF1174L Mutation Potentiates the Oncogenic Activity of MYCN in Neuroblastoma. *Cancer Cell* **2012**, *22*, 117–130. [[CrossRef](#)]
174. Valera, E.T.; Nader, L.; Queiroz, R.G.; Santos, A.C.; Sousa, G.R.; Oliveira, R.S.; Santos, M.V.; Machado, H.R.; Tone, L.G. Perinatal complex low- and high-grade glial tumor harboring a novel GIGYF2-ALK fusion. *Pediatr. Blood Cancer* **2019**, *67*, e28015. [[CrossRef](#)]
175. Argani, P.; Lian, D.W.; Agaimy, A.; Metzler, M.; Wobker, S.E.; Matoso, A.; Epstein, J.I.; Sung, Y.-S.; Zhang, L.; Antonescu, C.R. Pediatric Mesothelioma With ALK Fusions. *Am. J. Surg. Pathol.* **2021**, *45*, 653–661. [[CrossRef](#)] [[PubMed](#)]
176. Olsen, T.K.; Panagopoulos, I.; Meling, T.R.; Micci, F.; Gorunova, L.; Thorsen, J.; Due-Tønnessen, B.; Scheie, D.; Lund-Iversen, M.; Krossnes, B.; et al. Fusion genes with ALK as recurrent partner in ependymoma-like gliomas: A new brain tumor entity? *Neuro-Oncology* **2015**, *17*, 1365–1373. [[CrossRef](#)] [[PubMed](#)]



177. Łastowska, M.; Trubicka, J.; Niemira, M.; Paczkowska-Abdulsalam, M.; Karkucińska-Więckowska, A.; Kaleta, M.; Drogosiewicz, M.; Tarasińska, M.; Perek-Polnik, M.; Krętowski, A.; et al. ALK Expression Is a Novel Marker for the WNT-activated Type of Pediatric Medulloblastoma and an Indicator of Good Prognosis for Patients. *Am. J. Surg. Pathol.* **2017**, *41*, 781–787. [[CrossRef](#)] [[PubMed](#)]
178. Porta, C.; Paglino, C.; Mosca, A. Targeting PI3K/Akt/mTOR Signaling in Cancer. *Front. Oncol.* **2014**, *4*, 64. [[CrossRef](#)]
179. Castellano, E.; Downward, J. RAS Interaction with PI3K: More than Just another Effector Pathway. *Genes Cancer* **2011**, *2*, 261–274. [[CrossRef](#)]
180. Revathidevi, S.; Munirajan, A.K. Akt in cancer: Mediator and more. *Semin. Cancer Biol.* **2019**, *59*, 80–91. [[CrossRef](#)]
181. Barrett, D.; Brown, V.I.; Grupp, S.A.; Teachey, D.T. Targeting the PI3K/AKT/mTOR Signaling Axis in Children with Hematologic Malignancies. *Pediatr. Drugs* **2012**, *14*, 299–316. [[CrossRef](#)]
182. Zoncu, R.; Efeyan, A.; Sabatini, D.M. mTOR: From growth signal integration to cancer, diabetes and ageing. *Nat. Rev. Mol. Cell Biol.* **2011**, *12*, 21–35. [[CrossRef](#)]
183. Aoki, M.; Fujishita, T. Oncogenic Roles of the PI3K/AKT/mTOR Axis. *Curr. Top. Microbiol. Immunol.* **2017**, *407*, 153–189. [[CrossRef](#)]
184. Shorning, B.; Dass, M.; Smalley, M.; Pearson, H. The PI3K-AKT-mTOR Pathway and Prostate Cancer: At the Crossroads of AR, MAPK, and WNT Signaling. *Int. J. Mol. Sci.* **2020**, *21*, 4507. [[CrossRef](#)]
185. Narayanankutty, A. PI3K/ Akt/ mTOR Pathway as a Therapeutic Target for Colorectal Cancer: A Review of Preclinical and Clinical Evidence. *Curr. Drug Targets* **2019**, *20*, 1217–1226. [[CrossRef](#)]
186. Fattahi, S.; Amjadi-Moheb, F.; Tabaripour, R.; Ashrafi, G.H.; Akhavan-Niaki, H. PI3K/AKT/mTOR signaling in gastric cancer: Epigenetics and beyond. *Life Sci.* **2020**, *262*, 118513. [[CrossRef](#)]
187. Bertacchini, J.; Heidari, N.; Mediani, L.; Capitani, S.; Shahjahani, M.; Ahmadzadeh, A.; Saki, N. Targeting PI3K/AKT/mTOR network for treatment of leukemia. *Cell. Mol. Life Sci.* **2015**, *72*, 2337–2347. [[CrossRef](#)]
188. Miricescu, D.; Totan, A.; Stanescu-Spinu, I.-I.; Badoiu, S.; Stefani, C.; Greabu, M. PI3K/AKT/mTOR Signaling Pathway in Breast Cancer: From Molecular Landscape to Clinical Aspects. *Int. J. Mol. Sci.* **2020**, *22*, 173. [[CrossRef](#)]
189. Gann, C.-N.; Morsli, N.; Chen, X.; Barrueco, J. Response to ‘Dai W et al. Am J Cancer Res 2015;5(10):3270-3275’ from the makers of nintedanib. *Am. J. Cancer Res.* **2016**, *6*, 1547–1548.
190. Remke, M.; Pfister, S.; Kox, C.; Toedt, G.; Becker, N.; Benner, A.; Werft, W.; Breit, S.; Liu, S.; Engel, F.; et al. High-resolution genomic profiling of childhood T-ALL reveals frequent copy-number alterations affecting the TGF- $\beta$  and PI3K-AKT pathways and deletions at 6q15-16.1 as a genomic marker for unfavorable early treatment response. *Blood* **2009**, *114*, 1053–1062. [[CrossRef](#)]
191. Armengol, G.; Canellas, A.; Álvarez, Y.; Bastida, P.; De Toledo, J.S.; Pérez-Iribarne, M.D.M.; Camós, M.; Tuset, E.; Estella, J.; Coll, M.D.; et al. Genetic changes including gene copy number alterations and their relation to prognosis in childhood acute myeloid leukemia. *Leuk. Lymphoma* **2009**, *51*, 114–124. [[CrossRef](#)]
192. Knobbe, C.B.; Reifenberger, G. Genetic Alterations and Aberrant Expression of Genes Related to the Phosphatidylinositol-3'-Kinase/Protein Kinase B (Akt) Signal Transduction Pathway in Glioblastomas. *Brain Pathol.* **2006**, *13*, 507–518. [[CrossRef](#)]
193. Engelman, J.A.; Luo, J.; Cantley, L.C. The evolution of phosphatidylinositol 3-kinases as regulators of growth and metabolism. *Nat. Rev. Genet.* **2006**, *7*, 606–619. [[CrossRef](#)]
194. Okkenhaug, K.; Ahmadi, K.; White, S.; Timms, J.; Waterfield, M.D. Cellular Function of Phosphoinositide 3-Kinases: Implications for Development, Immunity, Homeostasis, and Cancer. *Annu. Rev. Cell Dev. Biol.* **2001**, *17*, 615–675. [[CrossRef](#)]
195. Zhao, L.; Vogt, P.K. Helical domain and kinase domain mutations in p110 of phosphatidylinositol 3-kinase induce gain of function by different mechanisms. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 2652–2657. [[CrossRef](#)]
196. Samuels, Y.; Wang, Z.; Bardelli, A.; Silliman, N.; Ptak, J.; Szabo, S.; Yan, H.; Gazdar, A.; Powell, S.M.; Riggins, G.J.; et al. High Frequency of Mutations of the PIK3CA Gene in Human Cancers. *Science* **2004**, *304*, 554. [[CrossRef](#)] [[PubMed](#)]
197. Jiang, W.; He, T.; Liu, S.; Zheng, Y.; Xiang, L.; Pei, X.; Wang, Z.; Yang, H. The PIK3CA E542K and E545K mutations promote glycolysis and proliferation via induction of the  $\beta$ -catenin/SIRT3 signaling pathway in cervical cancer. *J. Hematol. Oncol.* **2018**, *11*, 139. [[CrossRef](#)] [[PubMed](#)]
198. Ikenoue, T.; Kanai, F.; Hikiba, Y.; Obata, T.; Tanaka, Y.; Imamura, J.; Ohta, M.; Jazag, A.; Guleng, B.; Tateishi, K.; et al. Functional Analysis of PIK3CA Gene Mutations in Human Colorectal Cancer. *Cancer Res* **2005**, *65*, 4562–4567. [[CrossRef](#)] [[PubMed](#)]
199. Murat, C.; Braga, P.; Fortes, M.; Bronstein, M.; Correa-Giannella, M.L.; Giorgi, R. Mutation and genomic amplification of the PIK3CA proto-oncogene in pituitary adenomas. *Braz. J. Med. Biol. Res.* **2012**, *45*, 851–855. [[CrossRef](#)]
200. Shi, J.; Yao, D.; Liu, W.; Wang, N.; Lv, H.; Zhang, G.; Ji, M.; Xu, L.; He, N.; Shi, B.; et al. Highly frequent PIK3CA amplification is associated with poor prognosis in gastric cancer. *BMC Cancer* **2012**, *12*, 50. [[CrossRef](#)]
201. Holst, F.; Werner, H.M.; Mjøs, S.; Hoivik, E.A.; Kusonmano, K.; Wik, E.; Berg, A.; Birkeland, E.; Gibson, W.J.; Halle, M.K.; et al. PIK3CA Amplification Associates with Aggressive Phenotype but Not Markers of AKT-MTOR Signaling in Endometrial Carcinoma. *Clin. Cancer Res.* **2019**, *25*, 334–345. [[CrossRef](#)]
202. Huw, L.-Y.; O'Brien, C.; Pandita, A.; Mohan, S.; Spoerke, J.M.; Lu, S.; Wang, Y.; Hampton, G.M.; Wilson, T.R.; Lackner, M.R. Acquired PIK3CA amplification causes resistance to selective phosphoinositide 3-kinase inhibitors in breast cancer. *Oncogenesis* **2013**, *2*, e83. [[CrossRef](#)]



203. Salm, F.; Dimitrova, V.; Von Bueren, A.O.; Ćwiek, P.; Rehrauer, H.; Djonov, V.; Anderle, P.; Arcaro, A. The Phosphoinositide 3-Kinase p110 $\alpha$  Isoform Regulates Leukemia Inhibitory Factor Receptor Expression via c-Myc and miR-125b to Promote Cell Proliferation in Medulloblastoma. *PLoS ONE* **2015**, *10*, e0123958. [[CrossRef](#)]
204. Guerreiro, A.S.; Fattet, S.; Fischer, B.; Shalaby, T.; Jackson, S.P.; Schoenwaelder, S.M.; Grotzer, M.A.; Delattre, O.; Arcaro, A. Targeting the PI3K p110 $\alpha$  Isoform Inhibits Medulloblastoma Proliferation, Chemoresistance, and Migration. *Clin. Cancer Res.* **2008**, *14*, 6761–6769. [[CrossRef](#)]
205. Guerreiro, A.S.; Fattet, S.; Kulesza, D.W.; Atamer, A.; Elsing, A.N.; Shalaby, T.; Jackson, S.P.; Schoenwaelder, S.M.; Grotzer, M.A.; Delattre, O.; et al. A Sensitized RNA Interference Screen Identifies a Novel Role for the PI3K p110 $\gamma$  Isoform in Medulloblastoma Cell Proliferation and Chemoresistance. *Mol. Cancer Res.* **2011**, *9*, 925–935. [[CrossRef](#)]
206. Luk, S.K.; Piekorz, R.P.; Nürnberg, B.; To, S.-S.T. The catalytic phosphoinositid 3-kinase isoform p110 $\delta$  is required for glioma cell migration and invasion. *Eur. J. Cancer* **2012**, *48*, 149–157. [[CrossRef](#)]
207. Boller, D.; Schramm, A.; Doepfner, K.T.; Shalaby, T.; von Bueren, A.O.; Eggert, A.; Grotzer, M.A.; Arcaro, A. Targeting the Phosphoinositide 3-Kinase Isoform p110 $\delta$  Impairs Growth and Survival in Neuroblastoma Cells. *Clin. Cancer Res.* **2008**, *14*, 1172–1181. [[CrossRef](#)]
208. Fransson, S.; Martinsson, T.; Ejeskär, K. Neuroblastoma tumors with favorable and unfavorable outcomes: Significant differences in mRNA expression of genes mapped at 1p36.2. *Genes Chromosom. Cancer* **2006**, *46*, 45–52. [[CrossRef](#)]
209. Wang, Q.; Diskin, S.; Rappaport, E.; Attiyeh, E.; Mosse, Y.; Shue, D.; Seiser, E.; Jagannathan, J.; Shusterman, S.; Bansal, M.; et al. Integrative Genomics Identifies Distinct Molecular Classes of Neuroblastoma and Shows That Multiple Genes Are Targeted by Regional Alterations in DNA Copy Number. *Cancer Res* **2006**, *66*, 6050–6062. [[CrossRef](#)]
210. Yoon, C.; Lu, J.; Ryeom, S.W.; Simon, M.C.; Yoon, S.S. PIK3R3, part of the regulatory domain of PI3K, is upregulated in sarcoma stem-like cells and promotes invasion, migration, and chemotherapy resistance. *Cell Death Dis.* **2021**, *12*, 749. [[CrossRef](#)]
211. Staff, T.P.O. Correction: Variable expression of PIK3R3 and PTEN in Ewing sarcoma impacts oncogenic phenotypes. *PLoS ONE* **2015**, *10*, e0120830. [[CrossRef](#)]
212. Bellacosa, A.; Franke, T.F.; E Gonzalez-Portal, M.; Datta, K.; Taguchi, T.; Gardner, J.; Cheng, J.Q.; Testa, J.R.; Tsichlis, P.N. Structure, expression and chromosomal mapping of c-akt: Relationship to v-akt and its implications. *Oncogene* **1993**, *8*, 745–754.
213. Basu, A.; Lambring, C. Akt Isoforms: A Family Affair in Breast Cancer. *Cancers* **2021**, *13*, 3445. [[CrossRef](#)]
214. Meier, R.; Alessi, D.R.; Cron, P.; Andjelković, M.; Hemmings, B.A. Mitogenic Activation, Phosphorylation, and Nuclear Translocation of Protein Kinase B $\beta$ . *J. Biol. Chem.* **1997**, *272*, 30491–30497. [[CrossRef](#)]
215. Hinz, N.; Jücker, M. Distinct functions of AKT isoforms in breast cancer: A comprehensive review. *Cell Commun. Signal.* **2019**, *17*, 1–29. [[CrossRef](#)]
216. Chen, H.; Zhou, L.; Wu, X.; Li, R.; Wen, J.; Sha, J.; Wen, X. The PI3K AKT pathway in the pathogenesis of prostate cancer. *Front. Biosci.* **2016**, *21*, 1084–1091. [[CrossRef](#)]
217. Carpten, J.D.; Faber, A.L.; Horn, C.; Donoho, G.P.; Briggs, S.L.; Robbins, C.M.; Hostetter, G.; Boguslawski, S.; Moses, T.Y.; Savage, S.; et al. A transforming mutation in the pleckstrin homology domain of AKT1 in cancer. *Nature* **2007**, *448*, 439–444. [[CrossRef](#)]
218. Schüller, U.; Rüter, M.; Herms, J.; Kretzschmar, H.A.; Grasbon-Frodl, E. Absence of mutations in the AKT1 oncogene in glioblastomas and medulloblastomas. *Acta Neuropathol.* **2008**, *115*, 367–368. [[CrossRef](#)]
219. Hartmann, W.; Digon-Söntgerath, B.; Koch, A.; Waha, A.; Endl, E.; Dani, I.; Denkhäus, D.; Goodyer, C.G.; Sörensen, N.; Wiestler, O.D.; et al. Phosphatidylinositol 3'-Kinase/AKT Signaling Is Activated in Medulloblastoma Cell Proliferation and Is Associated with Reduced Expression of PTEN. *Clin. Cancer Res.* **2006**, *12*, 3019–3027. [[CrossRef](#)]
220. Granados, V.A.; Avirmeni-Vadlamudi, U.; Dalal, P.; Scarborough, S.R.; Galindo, K.A.; Mahajan, P.; Galindo, R.L. Selective Targeting of Myoblast Fusogenic Signaling and Differentiation-Arrest Antagonizes Rhabdomyosarcoma Cells. *Cancer Res* **2019**, *79*, 4585–4591. [[CrossRef](#)] [[PubMed](#)]
221. Hotfilder, M.; Sondermann, P.; Senß, A.; van Valen, F.; Jürgens, H.; Vormoor, J. PI3K/AKT is involved in mediating survival signals that rescue Ewing tumour cells from fibroblast growth factor 2-induced cell death. *Br. J. Cancer* **2005**, *92*, 705–710. [[CrossRef](#)]
222. Ren, C.; Pan, R.; Hou, L.; Wu, H.; Sun, J.; Zhang, W.; Tian, X.; Chen, H. Suppression of CLEC3A inhibits osteosarcoma cell proliferation and promotes their chemosensitivity through the AKT1/mTOR/HIF1 $\alpha$  signaling pathway. *Mol. Med. Rep.* **2020**, *21*, 1739–1748. [[CrossRef](#)]
223. Kuijjer, M.L.; Akker, B.E.W.M.V.D.; Hilhorst, R.; Mommersteeg, M.; Buddingh, E.; Serra, M.; Bürger, H.; Hogendoorn, P.C.W.; Cleton-Jansen, A.-M. Kinome and mRNA expression profiling of high-grade osteosarcoma cell lines implies Akt signaling as possible target for therapy. *BMC Med. Genom.* **2014**, *7*, 4. [[CrossRef](#)]
224. Zhu, Y.; Zhou, J.; Ji, Y.; Yu, B. Elevated expression of AKT2 correlates with disease severity and poor prognosis in human osteosarcoma. *Mol. Med. Rep.* **2014**, *10*, 737–742. [[CrossRef](#)] [[PubMed](#)]
225. Liu, W.; Zhou, Z.; Zhang, Q.; Rong, Y.; Li, L.; Luo, Y.; Wang, J.; Yin, G.; Lv, C.; Cai, W. Overexpression of miR-1258 inhibits cell proliferation by targeting AKT3 in osteosarcoma. *Biochem. Biophys. Res. Commun.* **2019**, *510*, 479–486. [[CrossRef](#)] [[PubMed](#)]
226. Qiao, J.; Lee, S.; Paul, P.; Qiao, L.; Taylor, C.J.; Schlegel, C.; Colon, N.C.; Chung, D.H. Akt2 Regulates Metastatic Potential in Neuroblastoma. *PLoS ONE* **2013**, *8*, e56382. [[CrossRef](#)] [[PubMed](#)]
227. Saxton, R.A.; Sabatini, D.M. mTOR Signaling in Growth, Metabolism, and Disease. *Cell* **2017**, *168*, 960–976, Erratum in *Cell* **2017**, *169*, 361–371. [[CrossRef](#)] [[PubMed](#)]
228. Hemmings, B.A.; Restuccia, D.F. PI3K-PKB/Akt Pathway. *Cold Spring Harb. Perspect. Biol.* **2012**, *4*, a011189. [[CrossRef](#)]

229. Zarogoulidis, P.; Lampaki, S.; Turner, J.F.; Huang, H.; Kakolyris, S.; Syrigos, K.; Zarogoulidis, K. mTOR pathway: A current, up-to-date mini-review (Review). *Oncol. Lett.* **2014**, *8*, 2367–2370. [[CrossRef](#)]
230. Zou, Z.; Tao, T.; Li, H.; Zhu, X. mTOR signaling pathway and mTOR inhibitors in cancer: Progress and challenges. *Cell Biosci.* **2020**, *10*, 31. [[CrossRef](#)]
231. Grabiner, B.C.; Nardi, V.; Birsoy, K.; Possemato, R.; Shen, K.; Sinha, S.; Jordan, A.; Beck, A.H.; Sabatini, D.M. A Diverse Array of Cancer-Associated MTOR Mutations Are Hyperactivating and Can Predict Rapamycin Sensitivity. *Cancer Discov.* **2014**, *4*, 554–563. [[CrossRef](#)]
232. Culjkovic, B.; Topisirovic, I.; Skrabanek, L.; Ruiz-Gutierrez, M.; Borden, K.L. eIF4E is a central node of an RNA regulon that governs cellular proliferation. *J. Cell Biol.* **2006**, *175*, 415–426. [[CrossRef](#)]
233. Mamane, Y.; Petroulakis, E.; Rong, L.; Yoshida, K.; Ler, L.W.; Sonenberg, N. eIF4E—From translation to transformation. *Oncogene* **2004**, *23*, 3172–3179. [[CrossRef](#)]
234. Wang, G.; Jia, Y.; Ye, Y.; Kang, E.; Chen, H.; Wang, J.; He, X. Identification of key methylation differentially expressed genes in posterior fossa ependymoma based on epigenomic and transcriptome analysis. *J. Transl. Med.* **2021**, *19*, 1–14. [[CrossRef](#)] [[PubMed](#)]
235. Machado, L.E.; Alvarenga, A.W.; Da Silva, F.F.; Roffé, M.; Begnami, M.D.; Torres, L.F.B.; Da Cunha, I.W.; Martins, V.R.; Hajj, G.N.M. Overexpression of mTOR and p(240–244)S6 in IDH1 Wild-Type Human Glioblastomas Is Predictive of Low Survival. *J. Histochem. Cytochem.* **2018**, *66*, 403–414. [[CrossRef](#)]
236. Shi, J.; Zhang, P.; Su, H.; Cai, L.; Zhao, L.; Zhou, H. Bioinformatics Analysis of Neuroblastoma miRNA Based on GEO Data. *Pharmacogenomics Pers. Med.* **2021**, *14*, 849–858. [[CrossRef](#)]
237. Pócza, T.; Sebestyén, A.; Turányi, E.; Krénacs, T.; Márk, A.; Sticz, T.B.; Jakab, Z.; Hauser, P. mTOR Pathway As a Potential Target In a Subset of Human Medulloblastoma. *Pathol. Oncol. Res.* **2014**, *20*, 893–900. [[CrossRef](#)]
238. Kaid, C.; Assoni, A.; Marçola, M.; Semedo-Kuriki, P.; Bortolin, R.H.; Carvalho, V.M.; Okamoto, O.K. Proteome and miRNome profiling of microvesicles derived from medulloblastoma cell lines with stem-like properties reveals biomarkers of poor prognosis. *Brain Res.* **2020**, *1730*, 146646. [[CrossRef](#)]
239. Chakraborty, S.; Khare, S.; Dorairaj, S.K.; Prabhakaran, V.C.; Prakash, D.R.; Kumar, A. Identification of genes associated with tumorigenesis of retinoblastoma by microarray analysis. *Genomics* **2007**, *90*, 344–353. [[CrossRef](#)]
240. Subbiah, V.; Brown, R.E.; Jiang, Y.; Buryanek, J.; Hayes-Jordan, A.; Kurzrock, R.; Anderson, P.M. Morphoproteomic Profiling of the Mammalian Target of Rapamycin (mTOR) Signaling Pathway in Desmoplastic Small Round Cell Tumor (EWS/WT1), Ewing's Sarcoma (EWS/FLI1) and Wilms' Tumor (WT1). *PLoS ONE* **2013**, *8*, e68985. [[CrossRef](#)]
241. Ahmed, A.A.; Sherman, A.K.; Pawel, B.R. Expression of therapeutic targets in Ewing sarcoma family tumors. *Hum. Pathol.* **2012**, *43*, 1077–1083. [[CrossRef](#)]
242. Dobashi, Y.; Suzuki, S.; Sato, E.; Hamada, Y.; Yanagawa, T.; Ooi, A. EGFR-dependent and independent activation of Akt/mTOR cascade in bone and soft tissue tumors. *Mod. Pathol.* **2009**, *22*, 1328–1340. [[CrossRef](#)]
243. Krishnan, K.; Bruce, B.; Hewitt, S.; Thomas, D.; Khanna, C.; Helman, L.J. Ezrin mediates growth and survival in Ewing's sarcoma through the AKT/mTOR, but not the MAPK, signaling pathway. *Clin. Exp. Metastasis* **2006**, *23*, 227–236. [[CrossRef](#)]
244. Hu, K.; Dai, H.-B.; Qiu, Z.-L. mTOR signaling in osteosarcoma: Oncogenesis and therapeutic aspects (Review). *Oncol. Rep.* **2016**, *36*, 1219–1225. [[CrossRef](#)] [[PubMed](#)]
245. Egas-Bejar, D.; Anderson, P.M.; Agarwal, R.; Corrales-Medina, F.; Devarajan, E.; Huh, W.W.; Brown, R.E.; Subbiah, V. Theranostic profiling for actionable aberrations in advanced high risk osteosarcoma with aggressive biology reveals high molecular diversity: The human fingerprint hypothesis. *Oncoscience* **2014**, *1*, 167–179. [[CrossRef](#)] [[PubMed](#)]
246. Petricoin, E.F.; Espina, V.; Araujo, R.P.; Midura, B.; Yeung, C.; Wan, X.; Eichler, G.S.; Johann, D.J.; Qualman, S.; Tsokos, M.; et al. Phosphoprotein Pathway Mapping: Akt/Mammalian Target of Rapamycin Activation Is Negatively Associated with Childhood Rhabdomyosarcoma Survival. *Cancer Res* **2007**, *67*, 3431–3440. [[CrossRef](#)]
247. Doble, B.W.; Woodgett, J.R. GSK-3: Tricks of the trade for a multi-tasking kinase. *J. Cell Sci.* **2003**, *116*, 1175–1186. [[CrossRef](#)]
248. Kockeritz, L.; Doble, B.; Patel, S.; Woodgett, J. Glycogen Synthase Kinase-3—An Overview of An Over-Achieving Protein Kinase. *Curr. Drug Targets* **2006**, *7*, 1377–1388. [[CrossRef](#)]
249. Sutherland, C. What Are the bona fide GSK3 Substrates? *Int. J. Alzheimer's Dis.* **2011**, *2011*, 505607. [[CrossRef](#)]
250. McCubrey, J.A.; Davis, N.M.; Abrams, S.L.; Montalto, G.; Cervello, M.; Basecke, J.; Libra, M.; Nicoletti, F.; Cocco, L.; Martelli, A.M.; et al. Diverse roles of GSK-3: Tumor promoter–tumor suppressor, target in cancer therapy. *Adv. Biol. Regul.* **2014**, *54*, 176–196. [[CrossRef](#)]
251. Patel, S.; Woodgett, J. Glycogen Synthase Kinase-3 and Cancer: Good Cop, Bad Cop? *Cancer Cell* **2008**, *14*, 351–353. [[CrossRef](#)]
252. Mancinelli, R.; Carpino, G.; Petrunaro, S.; Mammola, C.L.; Tomaipitina, L.; Filippini, A.; Facchiano, A.; Ziparo, E.; Giampietri, C. Multifaceted Roles of GSK-3 in Cancer and Autophagy-Related Diseases. *Oxidative Med. Cell. Longev.* **2017**, *2017*, 1–14. [[CrossRef](#)]
253. Luo, J. Glycogen synthase kinase 3 $\beta$  (GSK3 $\beta$ ) in tumorigenesis and cancer chemotherapy. *Cancer Lett.* **2008**, *273*, 194–200. [[CrossRef](#)]
254. Banerji, V.; Frumm, S.M.; Ross, K.N.; Li, L.S.; Schinzel, A.C.; Hahn, C.K.; Kakoza, R.M.; Chow, K.T.; Ross, L.; Alexe, G.; et al. The intersection of genetic and chemical genomic screens identifies GSK-3 $\alpha$  as a target in human acute myeloid leukemia. *J. Clin. Investig.* **2012**, *122*, 935–947. [[CrossRef](#)] [[PubMed](#)]

255. Farago, M.; Dominguez, I.; Landesman-Bollag, E.; Xu, X.; Rosner, A.; Cardiff, R.D.; Seldin, D.C. Kinase-Inactive Glycogen Synthase Kinase 3 $\beta$  Promotes Wnt Signaling and Mammary Tumorigenesis. *Cancer Res* **2005**, *65*, 5792–5801. [[CrossRef](#)]
256. Ougolkov, A.V.; Fernandez-Zapico, M.E.; Savoy, D.N.; Urrutia, R.A.; Billadeau, D.D. Glycogen Synthase Kinase-3 $\beta$  Participates in Nuclear Factor  $\kappa$ B-Mediated Gene Transcription and Cell Survival in Pancreatic Cancer Cells. *Cancer Res* **2005**, *65*, 2076–2081. [[CrossRef](#)] [[PubMed](#)]
257. Wang, Z.; Smith, K.S.; Murphy, M.; Piloto, O.; Somervaille, T.C.P.; Cleary, M.L. Glycogen synthase kinase 3 in MLL leukaemia maintenance and targeted therapy. *Nature* **2008**, *455*, 1205–1209. [[CrossRef](#)] [[PubMed](#)]
258. Brassesco, M.S.; Valera, E.T.; Meyer, C.; Marschalek, R.; Lopes, B.A.; Queiroz, R.G.D.P.; Calado, R.D.T.; Scrideli, C.; Tone, L.G. A new complex rearrangement in infant ALL: T(X;11;17)(p11.2;q23;q12). *Cancer Genet.* **2018**, *228–229*, 110–114. [[CrossRef](#)]
259. Meyer, C.; Burmeister, T.; Gröger, D.; Tsaur, G.; Fehina, L.; Renneville, A.; Sutton, R.; Venn, N.C.; Emerenciano, M.; Pombo-De-Oliveira, M.S.; et al. The MLL recombinome of acute leukemias in 2017. *Leukemia* **2018**, *32*, 273–284. [[CrossRef](#)]
260. Ocasio, J.K.; Bates, R.D.P.; Rapp, C.D.; Gershon, T.R. GSK-3 modulates SHH-driven proliferation in postnatal cerebellar neurogenesis and medulloblastoma. *Development* **2019**, *146*, dev177550. [[CrossRef](#)]
261. Silva-Evangelista, C.; Barret, E.; Ménez, V.; Merlevede, J.; Kergrohen, T.; Saccasyn, A.; Oberlin, E.; Puget, S.; Beccaria, K.; Grill, J.; et al. A kinome-wide shRNA screen uncovers vaccinia-related kinase 3 (VRK3) as an essential gene for diffuse intrinsic pontine glioma survival. *Oncogene* **2019**, *38*, 6479–6490. [[CrossRef](#)]
262. Lenz, J.E.; Riestler, R.; Schleicher, S.B.; Handgretinger, R.; Boehme, K.A.; Traub, F. Interaction of arsenic trioxide and etoposide in Ewing sarcoma cell lines. *Oncol. Rep.* **2019**, *43*, 337–345. [[CrossRef](#)]
263. Machado, I.; López-Guerrero, J.A.; Navarro, S.; Alberghini, M.; Scotlandi, K.; Picci, P.; Llombart-Bosch, A. Epithelial cell adhesion molecules and epithelial mesenchymal transition (EMT) markers in Ewing’s sarcoma family of tumors (ESFTs). Do they offer any prognostic significance? *Virchows Arch.* **2012**, *461*, 333–337. [[CrossRef](#)]
264. Ma, C.; Bower, K.A.; Chen, G.; Shi, X.; Ke, Z.-J.; Luo, J. Interaction between ERK and GSK3 $\beta$  Mediates Basic Fibroblast Growth Factor-induced Apoptosis in SK-N-MC Neuroblastoma Cells. *J. Biol. Chem.* **2008**, *283*, 9248–9256. [[CrossRef](#)]
265. Woodgett, J.R. Can a two-faced kinase be exploited for osteosarcoma? *Gynecol. Oncol.* **2012**, *104*, 722–723. [[CrossRef](#)]
266. Le Guellec, S.; Moyal, E.C.-J.; Filleron, T.; Delisle, M.-B.; Chevreau, C.; Rubie, H.; Castex, M.-P.; De Gauzy, J.S.; Bonneville, P.; Gomez-Brouchet, A. The  $\beta$ 5/focal adhesion kinase/glycogen synthase kinase 3 $\beta$  integrin pathway in high-grade osteosarcoma: A protein expression profile predictive of response to neoadjuvant chemotherapy. *Hum. Pathol.* **2013**, *44*, 2149–2158. [[CrossRef](#)]
267. Zeng, F.-Y.; Dong, H.; Cui, J.; Liu, L.; Chen, T. Glycogen synthase kinase 3 regulates PAX3-FKHR-mediated cell proliferation in human alveolar rhabdomyosarcoma cells. *Biochem. Biophys. Res. Commun.* **2010**, *391*, 1049–1055. [[CrossRef](#)]
268. Dionyssiou, M.G.; Ehyai, S.; Avrutin, E.; Connor, M.K.; McDermott, J.C. Glycogen synthase kinase 3 $\beta$  represses MYOGENIN function in alveolar rhabdomyosarcoma. *Cell Death Dis.* **2014**, *5*, e1094. [[CrossRef](#)]
269. Belyea, B.; Kephart, J.G.; Blum, J.; Kirsch, D.G.; Linaudic, C.M. Embryonic Signaling Pathways and Rhabdomyosarcoma: Contributions to Cancer Development and Opportunities for Therapeutic Targeting. *Sarcoma* **2012**, *2012*, 1–13. [[CrossRef](#)]
270. Ugolokov, A.V.; Bondarenko, G.I.; Dubrovskiy, O.; Berbegall, A.P.; Navarro, S.; Noguera, R.; O’Halloran, T.V.; Hendrix, M.J.; Giles, F.J.; Mazar, A.P. 9-ING-41, a small-molecule glycogen synthase kinase-3 inhibitor, is active in neuroblastoma. *Anti-Cancer Drugs* **2018**, *29*, 717–724. [[CrossRef](#)]
271. Li, Z.; Tan, F.; Thiele, C.J. Inactivation of glycogen synthase kinase-3 $\beta$  contributes to brain-derived neurotrophic factor/TrkB-induced resistance to chemotherapy in neuroblastoma cells. *Mol. Cancer Ther.* **2007**, *6*, 3113–3121. [[CrossRef](#)]
272. Duffy, D.J.; Krstic, A.; Schwarzl, T.; Higgins, D.G.; Kolch, W. GSK3 Inhibitors Regulate MYCN mRNA Levels and Reduce Neuroblastoma Cell Viability through Multiple Mechanisms, Including p53 and Wnt Signaling. *Mol. Cancer Ther.* **2014**, *13*, 454–467. [[CrossRef](#)]
273. Dickey, A.; Schleicher, S.; Leahy, K.; Hu, R.; Hallahan, D.; Thotala, D.K. GSK-3 $\beta$  inhibition promotes cell death, apoptosis, and in vivo tumor growth delay in neuroblastoma Neuro-2A cell line. *J. Neuro-Oncology* **2010**, *104*, 145–153. [[CrossRef](#)]
274. Katoh, Y.; Katoh, M. Hedgehog Target Genes: Mechanisms of Carcinogenesis Induced by Aberrant Hedgehog Signaling Activation. *Curr. Mol. Med.* **2009**, *9*, 873–886. [[CrossRef](#)] [[PubMed](#)]
275. Urbanska, K.; Trojanek, J.; Del Valle, L.; Eldeen, M.B.; Hofmann, F.; Garcia-Echeverria, C.; Khalili, K.; Reiss, K. Inhibition of IGF-I receptor in anchorage-independence attenuates GSK-3 $\beta$  constitutive phosphorylation and compromises growth and survival of medulloblastoma cell lines. *Oncogene* **2006**, *26*, 2308–2317. [[CrossRef](#)] [[PubMed](#)]
276. Atkins, R.; Stylli, S.; Luwor, R.; Kaye, A.; Hovens, C. Glycogen synthase kinase-3 $\beta$  (GSK-3 $\beta$ ) and its dysregulation in glioblastoma multiforme. *J. Clin. Neurosci.* **2013**, *20*, 1185–1192. [[CrossRef](#)] [[PubMed](#)]
277. Domoto, T.; Pyko, I.V.; Furuta, T.; Miyashita, K.; Uehara, M.; Shimasaki, T.; Nakada, M.; Minamoto, T. Glycogen synthase kinase-3 $\beta$  is a pivotal mediator of cancer invasion and resistance to therapy. *Cancer Sci.* **2016**, *107*, 1363–1372. [[CrossRef](#)] [[PubMed](#)]
278. Peti, W.; Page, R. Molecular basis of MAP kinase regulation. *Protein Sci.* **2013**, *22*, 1698–1710. [[CrossRef](#)]
279. Schaeffer, H.J.; Weber, M.J. Mitogen-Activated Protein Kinases: Specific Messages from Ubiquitous Messengers. *Mol. Cell. Biol.* **1999**, *19*, 2435–2444. [[CrossRef](#)]
280. Kim, E.K.; Choi, E.-J. Compromised MAPK signaling in human diseases: An update. *Arch. Toxicol.* **2015**, *89*, 867–882. [[CrossRef](#)]
281. Cuadrado, A.; Nebreda, A.R. Mechanisms and functions of p38 MAPK signalling. *Biochem. J.* **2010**, *429*, 403–417. [[CrossRef](#)]



282. Kim, E.K.; Choi, E.-J. Pathological roles of MAPK signaling pathways in human diseases. *Biochim. Biophys. Acta (BBA)-Mol. Basis Dis.* **2010**, *1802*, 396–405. [[CrossRef](#)]
283. Cicenás, J.; Zalyte, E.; Rimkus, A.; Dapkus, D.; Noreika, R.; Urbonavicius, S. JNK, p38, ERK, and SGK1 Inhibitors in Cancer. *Cancers* **2017**, *10*, 1. [[CrossRef](#)]
284. Mishima, K.; Inoue, K.; Hayashi, Y. Overexpression of extracellular-signal regulated kinases on oral squamous cell carcinoma. *Oral Oncol.* **2002**, *38*, 468–474. [[CrossRef](#)]
285. Kudaravalli, S.; Hollander, P.D.; Mani, S.A. Role of p38 MAP kinase in cancer stem cells and metastasis. *Oncogene* **2022**, *41*, 3177–3185. [[CrossRef](#)]
286. Tournier, C. The 2 Faces of JNK Signaling in Cancer. *Genes Cancer* **2013**, *4*, 397–400. [[CrossRef](#)]
287. Martínez-Limón, A.; Joaquin, M.; Caballero, M.; Posas, F.; de Nadal, E. The p38 Pathway: From Biology to Cancer Therapy. *Int. J. Mol. Sci.* **2020**, *21*, 1913. [[CrossRef](#)]
288. Pandey, P.; Raingeaud, J.; Kaneki, M.; Weichselbaum, R.; Davis, R.J.; Kufe, D.; Kharbanda, S. Activation of p38 Mitogen-activated Protein Kinase by c-Abl-dependent and -independent Mechanisms. *J. Biol. Chem.* **1996**, *271*, 23775–23779. [[CrossRef](#)]
289. Sui, X.; Kong, N.; Ye, L.; Han, W.; Zhou, J.; Zhang, Q.; He, C.; Pan, H. p38 and JNK MAPK pathways control the balance of apoptosis and autophagy in response to chemotherapeutic agents. *Cancer Lett.* **2014**, *344*, 174–179. [[CrossRef](#)]
290. Grossi, V.; Peserico, A.; Tezil, T.; Simone, C. p38 $\alpha$  MAPK pathway: A key factor in colorectal cancer therapy and chemoresistance. *World J. Gastroenterol.* **2014**, *20*, 9744–9758. [[CrossRef](#)]
291. Masliah-Planchon, J.; Garinet, S.; Pasmant, E. RAS-MAPK pathway epigenetic activation in cancer: miRNAs in action. *Oncotarget* **2015**, *7*, 38892–38907. [[CrossRef](#)]
292. Schubbert, S.; Shannon, K.; Bollag, G. Hyperactive Ras in developmental disorders and cancer. *Nat. Rev. Cancer* **2007**, *7*, 295–308. [[CrossRef](#)]
293. Liu, F.; Yang, X.; Geng, M.; Huang, M. Targeting ERK, an Achilles' Heel of the MAPK pathway, in cancer therapy. *Acta Pharm. Sin. B* **2018**, *8*, 552–562. [[CrossRef](#)]
294. Malumbres, M.; Barbacid, M. RAS oncogenes: The first 30 years. *Nat. Rev. Cancer* **2003**, *3*, 459–465. [[CrossRef](#)] [[PubMed](#)]
295. Hobbs, G.A.; Der, C.J.; Rossman, K.L. RAS isoforms and mutations in cancer at a glance. *J. Cell Sci.* **2016**, *129*, 1287–1292. [[CrossRef](#)]
296. Yaeger, R.; Corcoran, R.B. Targeting Alterations in the RAF–MEK Pathway. *Cancer Discov.* **2019**, *9*, 329–341. [[CrossRef](#)] [[PubMed](#)]
297. Burotto, M.; Chiou, V.L.; Lee, J.-M.; Kohn, E.C. The MAPK pathway across different malignancies: A new perspective. *Cancer* **2014**, *120*, 3446–3456. [[CrossRef](#)] [[PubMed](#)]
298. Stefan, E.; Bister, K. MYC and RAF: Key Effectors in Cellular Signaling and Major Drivers in Human Cancer. *Poxviruses* **2017**, *407*, 117–151. [[CrossRef](#)]
299. Maurer, G.; Tarkowski, B.; Baccharini, M. Raf kinases in cancer—roles and therapeutic opportunities. *Oncogene* **2011**, *30*, 3477–3488. [[CrossRef](#)]
300. Lito, P.; Rosen, N.; Solit, D.B. Tumor adaptation and resistance to RAF inhibitors. *Nat. Med.* **2013**, *19*, 1401–1409. [[CrossRef](#)]
301. The Cancer Genome Atlas Network. Genomic Classification of Cutaneous Melanoma. *Cell* **2015**, *161*, 1681–1696. [[CrossRef](#)]
302. Samatar, A.A.; Poulikakos, P.I. Targeting RAS–ERK signalling in cancer: Promises and challenges. *Nat. Rev. Drug Discov.* **2014**, *13*, 928–942. [[CrossRef](#)]
303. Pfister, S.; Janzarik, W.G.; Remke, M.; Ernst, A.; Werft, W.; Becker, N.; Toedt, G.; Wittmann, A.; Kratz, C.; Olbrich, H.; et al. BRAF gene duplication constitutes a mechanism of MAPK pathway activation in low-grade astrocytomas. *J. Clin. Investig.* **2008**, *118*, 1739–1749. [[CrossRef](#)]
304. Bar, E.E.; Lin, A.; Tihan, T.; Burger, P.C.; Eberhart, C.G. Frequent Gains at Chromosome 7q34 Involving BRAF in Pilocytic Astrocytoma. *J. Neuropathol. Exp. Neurol.* **2008**, *67*, 878–887. [[CrossRef](#)]
305. Reis, G.F.; Bloomer, M.M.; Perry, A.; Phillips, J.J.; Grenert, J.P.; Karnezis, A.N.; Tihan, T. Pilocytic astrocytomas of the optic nerve and their relation to pilocytic astrocytomas elsewhere in the central nervous system. *Mod. Pathol.* **2013**, *26*, 1279–1287. [[CrossRef](#)]
306. Anagnostopoulos, A.K.; Dimas, K.S.; Papathanassiou, C.; Braoudaki, M.; Anastasiadou, E.; Vougas, K.; Karamolegou, K.; Kontos, H.; Prodromou, N.; Tzortzatou-Stathopoulou, F.; et al. Proteomics Studies of Childhood Pilocytic Astrocytoma. *J. Proteome Res.* **2011**, *10*, 2555–2565. [[CrossRef](#)]
307. MacDonald, T.J.; Brown, K.M.; LaFleur, B.; Peterson, K.; Lawlor, C.; Chen, Y.; Packer, R.J.; Cogen, P.; Stephan, D.A. Expression profiling of medulloblastoma: PDGFRA and the RAS/MAPK pathway as therapeutic targets for metastatic disease. *Nat. Genet.* **2001**, *29*, 143–152. [[CrossRef](#)]
308. Badodi, S.; Pomella, N.; Lim, Y.M.; Brandner, S.; Morrison, G.; Pollard, S.M.; Zhang, X.; Zabet, N.R.; Marino, S. Combination of BMI1 and MAPK/ERK inhibitors is effective in medulloblastoma. *Neuro-Oncology* **2022**, *24*, 1273–1285. [[CrossRef](#)]
309. Tsumura, H.; Yoshida, T.; Saito, H.; Imanaka-Yoshida, K.; Suzuki, N. Cooperation of oncogenic K-ras and p53 deficiency in pleomorphic rhabdomyosarcoma development in adult mice. *Oncogene* **2006**, *25*, 7673–7679. [[CrossRef](#)]
310. Na, K.Y.; Kim, Y.W.; Park, Y.-K. Mitogen-activated protein kinase pathway in osteosarcoma. *Pathology* **2012**, *44*, 540–546. [[CrossRef](#)]
311. Wu, J.; Zhang, C.; Chen, L. MiR-511 mimic transfection inhibits the proliferation, invasion of osteosarcoma cells and reduces metastatic osteosarcoma tumor burden in nude mice via targeting MAPK1. *Cancer Biomarkers* **2019**, *26*, 343–351. [[CrossRef](#)]
312. Guo, H.; Zhang, H.-Y.; Wang, S.-L.; Ye, L.; Yang, G.-H.; Bu, H. Smad4 and ERK2 stimulated by transforming growth factor beta1 in rhabdomyosarcoma. *Chin. Med. J.* **2007**, *120*, 515–521. [[CrossRef](#)]



313. Lynch, J.; Fay, J.; Meehan, M.; Bryan, K.; Watters, K.M.; Murphy, D.M.; Stallings, R.L. MiRNA-335 suppresses neuroblastoma cell invasiveness by direct targeting of multiple genes from the non-canonical TGF- $\beta$  signalling pathway. *Carcinog.* **2012**, *33*, 976–985. [[CrossRef](#)]
314. Tabatabaei, S.N.; Derbali, R.M.; Yang, C.; Superstein, R.; Hamel, P.; Chain, J.L.; Hardy, P. Co-delivery of miR-181a and melphalan by lipid nanoparticles for treatment of seeded retinoblastoma. *J. Control. Release* **2019**, *298*, 177–185. [[CrossRef](#)] [[PubMed](#)]
315. Poon, R.Y.C. Cell cycle control: A system of interlinking oscillators. *Methods Mol. Biol.* **2016**, *1342*, 3–19. [[PubMed](#)]
316. Saka, Y.; Giuraniuc, C.V.; Ohkura, H. Accurate chromosome segregation by probabilistic self-organisation. *BMC Biol.* **2015**, *13*, 1–10. [[CrossRef](#)] [[PubMed](#)]
317. Gao, S.-W.; Liu, F. Novel insights into cell cycle regulation of cell fate determination. *J. Zhejiang Univ. B* **2019**, *20*, 467–475. [[CrossRef](#)]
318. Ding, L.; Cao, J.; Lin, W.; Chen, H.; Xiong, X.; Ao, H.; Yu, M.; Lin, J.; Cui, Q. The Roles of Cyclin-Dependent Kinases in Cell-Cycle Progression and Therapeutic Strategies in Human Breast Cancer. *Int. J. Mol. Sci.* **2020**, *21*, 1960. [[CrossRef](#)]
319. Pines, J. The cell cycle kinases. *Semin. Cancer Biol.* **1994**, *5*, 305–313.
320. Bruyère, C.; Meijer, L. Targeting cyclin-dependent kinases in anti-neoplastic therapy. *Curr. Opin. Cell Biol.* **2013**, *25*, 772–779. [[CrossRef](#)]
321. Chilà, R.; Guffanti, F.; Damia, G. Role and therapeutic potential of CDK12 in human cancers. *Cancer Treat. Rev.* **2016**, *50*, 83–88. [[CrossRef](#)]
322. Sherr, C.J. Cancer Cell Cycles. *Science* **1996**, *274*, 1672–1677. [[CrossRef](#)]
323. Hall, M.; Peters, G. Genetic Alterations of Cyclins, Cyclin-Dependent Kinases, and Cdk Inhibitors in Human Cancer. *Adv. Cancer Res.* **1996**, *68*, 67–108. [[CrossRef](#)]
324. Malumbres, M. Cyclins and related kinases in cancer cells. *J. BUON.* **2007**, *12* (Suppl. S1), S45–S52.
325. Malumbres, M.; Barbacid, M. Cell cycle, CDKs and cancer: A changing paradigm. *Nat. Rev. Cancer* **2009**, *9*, 153–166. [[CrossRef](#)]
326. Niwa, T.; Akaike, Y.; Watanabe, K.; Chibazakura, T. Hyperactivation of cyclin A-CDK induces centrosome overduplication and chromosome tetraploidization in mouse cells. *Biochem. Biophys. Res. Commun.* **2021**, *549*, 91–97. [[CrossRef](#)]
327. Viotto, D.; Russo, F.; Anania, I.; Segatto, I.; Vinciguerra, G.L.R.; Dall’Acqua, A.; Bomben, R.; Perin, T.; Cusan, M.; Schiappacassi, M.; et al. *CDKN1B* mutation and copy number variation are associated with tumor aggressiveness in luminal breast cancer. *J. Pathol.* **2020**, *253*, 234–245. [[CrossRef](#)]
328. Lu, Y.; Leow, A.; O Madu, C. The Role of Cyclin-Dependent Kinases on the Metastasis of Breast Cancer. *Nov. Approaches Cancer Study* **2020**, *4*, 377–385. [[CrossRef](#)]
329. Lam, Y.; E. di Tomaso, H.-K.; Ng, J.C.S.; Pang, M.F.; Roussel, N.M.; Hjelm, P. Expression of p19 INK4d, CDK4, CDK6 in glioblastoma multiforme. *Br. J. Neurosurg.* **2000**, *14*, 28–32. [[CrossRef](#)]
330. Richardson, S.; Hill, R.M.; Kui, C.; Lindsey, J.C.; Grabovksa, Y.; Keeling, C.; Pease, L.; Bashton, M.; Crosier, S.; Vinci, M.; et al. Emergence and maintenance of actionable genetic drivers at medulloblastoma relapse. *Neuro-Oncology* **2021**, *24*, 153–165. [[CrossRef](#)]
331. Wood, A.C.; Krytska, K.; Ryles, H.T.; Infarinato, N.R.; Sano, R.; Hansel, T.D.; Hart, L.S.; King, F.J.; Smith, T.R.; Ainscow, E.; et al. Dual *ALK* and *CDK4/6* Inhibition Demonstrates Synergy against Neuroblastoma. *Clin. Cancer Res.* **2017**, *23*, 2856–2868. [[CrossRef](#)]
332. Iolascon, A.; Faienza, M.F.; Coppola, B.; Rosolen, A.; Basso, G.; Ragione, F.D.; Schettini, F. Analysis of cyclin-dependent kinase inhibitor genes (*CDKN2A*, *CDKN2B*, and *CDKN2C*) in childhood rhabdomyosarcoma. *Genes Chromosomes Cancer* **1996**, *15*, 217–222. [[CrossRef](#)]
333. Komuro, H.; Valentine, M.B.; Rubnitz, J.E.; Saito, M.; Raimondi, S.C.; Carroll, A.J.; Yi, T.; Sherr, C.J.; Look, A.T. p27KIP1 Deletions in Childhood Acute Lymphoblastic Leukemia. *Neoplasia* **1999**, *1*, 253–261. [[CrossRef](#)]
334. Martinez-Soria, N.; McKenzie, L.; Draper, J.; Ptasinska, A.; Issa, H.; Potluri, S.; Blair, H.J.; Pickin, A.; Isa, A.; Chin, P.S.; et al. The Oncogenic Transcription Factor *RUNX1/ETO* Corrupts Cell Cycle Regulation to Drive Leukemic Transformation. *Cancer Cell* **2018**, *34*, 626–642.e8, Erratum in **2019**, *35*, P705. [[CrossRef](#)] [[PubMed](#)]
335. Diril, M.K.; Ratnacaram, C.K.; Padmakumar, V.C.; Du, T.; Wasser, M.; Coppola, V.; Tessarollo, L.; Kaldis, P. Cyclin-dependent kinase 1 (Cdk1) is essential for cell division and suppression of DNA re-replication but not for liver regeneration. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 3826–3831. [[CrossRef](#)] [[PubMed](#)]
336. Liu, X.; Wu, H.; Liu, Z. An Integrative Human Pan-Cancer Analysis of Cyclin-Dependent Kinase 1 (CDK1). *Cancers* **2022**, *14*, 2658. [[CrossRef](#)] [[PubMed](#)]
337. Ying, X.; Che, X.; Wang, J.; Zou, G.; Yu, Q.; Zhang, X. CDK1 serves as a novel therapeutic target for endometrioid endometrial cancer. *J. Cancer* **2021**, *12*, 2206–2215. [[CrossRef](#)]
338. Li, M.; He, F.; Zhang, Z.; Xiang, Z.; Hu, D. CDK1 serves as a potential prognostic biomarker and target for lung cancer. *J. Int. Med. Res.* **2020**, *48*. [[CrossRef](#)]
339. Kim, S.J.; Nakayama, S.; Miyoshi, Y.; Taguchi, T.; Tamaki, Y.; Matsushima, T.; Torikoshi, Y.; Tanaka, S.; Yoshida, T.; Ishihara, H.; et al. Determination of the specific activity of CDK1 and CDK2 as a novel prognostic indicator for early breast cancer. *Ann. Oncol.* **2007**, *19*, 68–72. [[CrossRef](#)]
340. Wang, C.; Xie, X.; Li, W.; Jiang, D. Expression of *KIF2A*, *NDC80*, *CDK1*, and *CCNB1* in breast cancer patients: Their interaction and linkage with tumor features and prognosis. *J. Clin. Lab. Anal.* **2022**, *36*, e24647. [[CrossRef](#)]

341. Xing, Z.; Wang, X.; Liu, J.; Zhang, M.; Feng, K.; Wang, X. Expression and prognostic value of CDK1, CCNA2, and CCNB1 gene clusters in human breast cancer. *J. Int. Med. Res.* **2021**, *49*. [\[CrossRef\]](#)
342. Huang, Z.; Shen, G.; Gao, J. CDK1 promotes the stemness of lung cancer cells through interacting with Sox2. *Clin. Transl. Oncol.* **2021**, *23*, 1743–1751. [\[CrossRef\]](#)
343. Ravindran Menon, D.; Luo, Y.; Arcaroli, J.J.; Liu, S.; Krishnankutty, L.N.; Osborne, D.G.; Li, Y.; Samson, J.M.; Bagby, S.; Tan, A.C.; et al. CDK1 Interacts with Sox2 and Promotes Tumor Initiation in Human Melanoma. *Cancer Res.* **2018**, *78*, 6561–6574. [\[CrossRef\]](#)
344. Zhong, S.; Yan, Q.; Ge, J.; Dou, G.; Zhao, G. Identification of driver genes and key pathways of ependymoma. *Turk. Neurosurg.* **2018**. [\[CrossRef\]](#)
345. Pérez-Ramírez, M.; Hernández-Jiménez, A.J.; Guerrero-Guerrero, A.; Benadón-Darszon, E.; Pérezpeña-Díazconti, M.; Siordia-Reyes, A.G.; García-Méndez, A.; de León, F.C.-P.; Salamanca-Gómez, F.A.; García-Hernández, N. Genomics and epigenetics: A study of ependymomas in pediatric patients. *Clin. Neurol. Neurosurg.* **2016**, *144*, 53–58. [\[CrossRef\]](#)
346. Li, Q.; Zhang, L.; Jiang, J.; Zhang, Y.; Wang, X.; Zhang, Q.; Wang, Y.; Liu, C.; Li, F. CDK1 and CCNB1 as potential diagnostic markers of rhabdomyosarcoma: Validation following bioinformatics analysis. *BMC Med. Genom.* **2019**, *12*, 198. [\[CrossRef\]](#)
347. Lu, S.; Sun, C.; Chen, H.; Zhang, C.; Li, W.; Wu, L.; Zhu, J.; Sun, F.; Huang, J.; Wang, J.; et al. Bioinformatics Analysis and Validation Identify CDK1 and MAD2L1 as Prognostic Markers of Rhabdomyosarcoma. *Cancer Manag. Res.* **2020**, *12*, 12123–12136. [\[CrossRef\]](#)
348. Schwermer, M.; Lee, S.; Köster, J.; van Maerken, T.; Stephan, H.; Eggert, A.; Morik, K.; Schulte, J.H.; Schramm, A. Sensitivity to cdk1-inhibition is modulated by p53 status in preclinical models of embryonal tumors. *Oncotarget* **2015**, *6*, 15425–15435. [\[CrossRef\]](#)
349. Shi, K.; Zhu, X.; Wu, J.; Chen, Y.; Zhang, J.; Sun, X. Centromere protein E as a novel biomarker and potential therapeutic target for retinoblastoma. *Bioengineered* **2021**, *12*, 5950–5970. [\[CrossRef\]](#)
350. Liu, J.; Wu, S.; Xie, X.; Wang, Z.; Lei, Q. Identification of potential crucial genes and key pathways in osteosarcoma. *Hereditas* **2020**, *157*, 29. [\[CrossRef\]](#)
351. Liu, L.; Xu, Y.; Reiter, R.J. Melatonin inhibits the proliferation of human osteosarcoma cell line MG-63. *Bone* **2013**, *55*, 432–438. [\[CrossRef\]](#)
352. Zhang, J.; Zhu, X.; Li, H.; Li, B.; Sun, L.; Xie, T.; Zhu, T.; Zhou, H.; Ye, Z. Piperine inhibits proliferation of human osteosarcoma cells via G2/M phase arrest and metastasis by suppressing MMP-2/-9 expression. *Int. Immunopharmacol.* **2015**, *24*, 50–58. [\[CrossRef\]](#)
353. Cai, D.; Latham, V.M.; Zhang, X.; Shapiro, G.I. Combined Depletion of Cell Cycle and Transcriptional Cyclin-Dependent Kinase Activities Induces Apoptosis in Cancer Cells. *Cancer Res* **2006**, *66*, 9270–9280. [\[CrossRef\]](#)
354. Chen, H.; Huang, Q.; Zhai, D.; Dong, J.; Wang, A.; Lan, Q. CDK1 expression and effects of CDK1 silencing on the malignant phenotype of glioma cells. *Zhonghua Zhong Liu Za Zhi* **2007**, *29*, 484–488.
355. Zhou, Y.; Yang, L.; Zhang, X.; Chen, R.; Chen, X.; Tang, W.; Zhang, M. Identification of Potential Biomarkers in Glioblastoma through Bioinformatic Analysis and Evaluating Their Prognostic Value. *BioMed Res. Int.* **2019**, *2019*, 1–13. [\[CrossRef\]](#) [\[PubMed\]](#)
356. McCurdy, S.R.; Pacal, M.; Ahmad, M.; Bremner, R. A CDK2 activity signature predicts outcome in CDK2-low cancers. *Oncogene* **2016**, *36*, 2491–2502. [\[CrossRef\]](#) [\[PubMed\]](#)
357. Santamaría, D.; Barrière, C.; Cerqueira, A.; Hunt, S.; Tardy, C.; Newton, K.; Cáceres, J.F.; Dubus, P.; Malumbres, M.; Barbacid, M. Cdk1 is sufficient to drive the mammalian cell cycle. *Nature* **2007**, *448*, 811–815. [\[CrossRef\]](#) [\[PubMed\]](#)
358. Tadesse, S.; Caldon, E.C.; Tilley, W.; Wang, S. Cyclin-Dependent Kinase 2 Inhibitors in Cancer Therapy: An Update. *J. Med. Chem.* **2018**, *62*, 4233–4251. [\[CrossRef\]](#)
359. Teixeira, L.K.; Wang, X.; Li, Y.; Ekholm-Reed, S.; Wu, X.; Wang, P.; Reed, S.I. Cyclin E Deregulation Promotes Loss of Specific Genomic Regions. *Curr. Biol.* **2015**, *25*, 1327–1333. [\[CrossRef\]](#)
360. Tadesse, S.; Anshabo, A.T.; Portman, N.; Lim, E.; Tilley, W.; Caldon, C.E.; Wang, S. Targeting CDK2 in cancer: Challenges and opportunities for therapy. *Drug Discov. Today* **2019**, *25*, 406–413. [\[CrossRef\]](#)
361. Campaner, S.; Doni, M.; Hydrbring, P.; Verrecchia, A.; Bianchi, L.; Sardella, D.; Schleker, T.; Perna, D.; Tronnersjö, S.; Murga, M.; et al. Cdk2 suppresses cellular senescence induced by the c-myc oncogene. *Nature* **2009**, *12*, 54–59. [\[CrossRef\]](#)
362. Chen, Z.; Wang, Z.; Pang, J.; Yu, Y.; Bieerkehazhi, S.; Lu, J.; Hu, T.; Zhao, Y.; Xu, X.; Zhang, H.; et al. Multiple CDK inhibitor dinaciclib suppresses neuroblastoma growth via inhibiting CDK2 and CDK9 activity. *Sci. Rep.* **2016**, *6*, 29090. [\[CrossRef\]](#)
363. Bo, L.; Wei, B.; Wang, Z.; Kong, D.; Gao, Z.; Miao, Z. Bioinformatics analysis of the CDK2 functions in neuroblastoma. *Mol. Med. Rep.* **2017**, *17*, 3951–3959. [\[CrossRef\]](#)
364. Bolin, S.; Borgenvik, A.; Persson, C.; Rosén, G.; Sundström, A.; Qi, J.; Bradner, J.E.; Weiss, W.A.; Cho, Y.-J.; Weishaupt, H.; et al. Abstract 2473: Combined BET-bromodomain and CDK2 inhibition in MYC-driven medulloblastoma. *Cancer Res* **2016**, *76*, 2473. [\[CrossRef\]](#)
365. Liu, H.; Weng, J. A comprehensive bioinformatic analysis of cyclin-dependent kinase 2 (CDK2) in glioma. *Gene* **2022**, *822*, 146325. [\[CrossRef\]](#)
366. Zhang, Y.; Xue, C.; Zhu, X.; Xian, H.; Huang, Z. Suppression of microRNA-125a-5p upregulates the TAZ-EGFR signaling pathway and promotes retinoblastoma proliferation. *Cell Signal.* **2016**, *28*, 850–860. [\[CrossRef\]](#)
367. Zhang, Y.; Xue, C.; Cui, H.; Huang, Z. High expression of TAZ indicates a poor prognosis in retinoblastoma. *Diagn. Pathol.* **2015**, *10*, 187. [\[CrossRef\]](#)
368. Knudsen, E.S.; Pazzagli, C.; Born, T.L.; Bertolaet, B.L.; Knudsen, K.; Arden, K.C.; Henry, R.R.; Feramisco, J.R. Elevated cyclins and cyclin-dependent kinase activity in the rhabdomyosarcoma cell line RD. *Cancer Res* **1998**, *58*, 2042–2049.

369. Fu, W.; Ma, L.; Chu, B.; Wang, X.; Bagui, T.K.; Bui, M.M.; Gemmer, J.; Altiok, S.; Letson, D.G.; Pledger, W.J. Abstract 3596: SCH727965, a cyclin-dependent kinases inhibitor, induces apoptosis in sarcoma cells through caspase 3- dependent pathway. *Cancer Res* **2011**, *71*, 3596. [[CrossRef](#)]
370. Musa, J.; Cidre-Aranaz, F.; Aynaud, M.-M.; Orth, M.F.; Knott, M.M.L.; Mirabeau, O.; Mazor, G.; Varon, M.; Hölting, T.L.B.; Grossetête, S.; et al. Cooperation of cancer drivers with regulatory germline variants shapes clinical outcomes. *Nat. Commun.* **2019**, *10*, 4128. [[CrossRef](#)]
371. Ohali, A.; Avigad, S.; Zaizov, R.; Ophir, R.; Horn-Saban, S.; Cohen, I.J.; Meller, I.; Kollender, Y.; Issakov, J.; Yaniv, I. Prediction of high risk Ewing's sarcoma by gene expression profiling. *Oncogene* **2004**, *23*, 8997–9006. [[CrossRef](#)]
372. Gao, X.; Leone, G.W.; Wang, H. Cyclin D-CDK4/6 functions in cancer. *Adv. Cancer Res.* **2020**, *148*, 147–169.
373. Dobashi, Y.; Goto, A.; Fukayama, M.; Abe, A.; Ooi, A. Overexpression of cdk4/cyclin D1, a possible mediator of apoptosis and an indicator of prognosis in human primary lung carcinoma. *Int. J. Cancer* **2004**, *110*, 532–541. [[CrossRef](#)]
374. Nadal, A.; Jares, P.; Pinyol, M.; Conde, L.; Romeu, C.; Fernández, P.L.; Campo, E.; Cardesa, A. Association of CDK4 and CCND1 mRNA overexpression in laryngeal squamous cell carcinomas occurs without CDK4 amplification. *Virchows Arch.* **2006**, *450*, 161–167. [[CrossRef](#)] [[PubMed](#)]
375. Hashimoto, H.; Kaku-Ito, Y.; Oda, Y.; Ito, T. CDK4: A Novel Therapeutic Target for Extramammary Paget's Disease. *Front. Oncol.* **2021**, *11*, 710378. [[CrossRef](#)] [[PubMed](#)]
376. Dong, Y.; Sui, L.; Sugimoto, K.; Tai, Y.; Tokuda, M. Cyclin D1-CDK4 complex, a possible critical factor for cell proliferation and prognosis in laryngeal squamous cell carcinomas. *Int. J. Cancer* **2001**, *95*, 209–215. [[CrossRef](#)] [[PubMed](#)]
377. Chen, T.-J.; Lee, S.-W.; Lin, L.-C.; Lin, C.-Y.; Chang, K.-Y.; Li, C.-F. Cyclin-dependent kinase 4 overexpression is mostly independent of gene amplification and constitutes an independent prognosticator for nasopharyngeal carcinoma. *Tumor Biol.* **2014**, *35*, 7209–7216. [[CrossRef](#)]
378. Wu, A.; Wu, B.; Guo, J.; Luo, W.; Wu, D.; Yang, H.; Zhen, Y.; Yu, X.; Wang, H.; Zhou, Y.; et al. Elevated expression of CDK4 in lung cancer. *J. Transl. Med.* **2011**, *9*, 38. [[CrossRef](#)]
379. Lu, J.-W.; Lin, Y.-M.; Chang, J.-G.; Yeh, K.-T.; Chen, R.-M.; Tsai, J.J.P.; Su, W.-W.; Hu, R.-M. Clinical implications of deregulated CDK4 and Cyclin D1 expression in patients with human hepatocellular carcinoma. *Med. Oncol.* **2013**, *30*, 379. [[CrossRef](#)]
380. An, H.-X.; Beckmann, M.W.; Reifemberger, G.; Bender, H.G.; Niederacher, D. Gene Amplification and Overexpression of CDK4 in Sporadic Breast Carcinomas Is Associated with High Tumor Cell Proliferation. *Am. J. Pathol.* **1999**, *154*, 113–118. [[CrossRef](#)]
381. Yang, C.; Li, Z.; Bhatt, T.; Dickler, M.; Giri, D.; Scaltriti, M.; Baselga, J.; Rosen, N.; Chandralapaty, S. Acquired CDK6 amplification promotes breast cancer resistance to CDK4/6 inhibitors and loss of ER signaling and dependence. *Oncogene* **2016**, *36*, 2255–2264. [[CrossRef](#)]
382. Wölfel, T.; Hauer, M.; Schneider, J.; Serrano, M.; Wölfel, C.; Klehmann-Hieb, E.; De Plaen, E.; Hankeln, T.; Büschenfelde, K.-H.M.Z.; Beach, D. A p16<sup>INK4a</sup>-insensitive CDK4 mutant targeted by cytolytic T lymphocytes in a human melanoma. *Science* **1995**, *269*, 1281–1284. [[CrossRef](#)]
383. Kim, H.; Ham, E.K.; Kim, Y.I.; Chi, J.G.; Lee, H.S.; Park, S.H.; Jung, Y.M.; Myung, N.K.; Lee, M.J.; Jang, J.-J. Overexpression of cyclin D1 and cdk4 in tumorigenesis of sporadic hepatoblastomas. *Cancer Lett.* **1998**, *131*, 177–183. [[CrossRef](#)]
384. Nebenfuhr, S.; Kollmann, K.; Sexl, V. The role of CDK6 in cancer. *Int. J. Cancer* **2020**, *147*, 2988–2995. [[CrossRef](#)]
385. Jena, N.; Sheng, J.; Hu, J.K.; Li, W.; Zhou, W.; Lee, G.; Tschlis, N.; A Pathak, A.; Brown, N.; A Deshpande, A.; et al. CDK6-mediated repression of CD25 is required for induction and maintenance of Notch1-induced T-cell acute lymphoblastic leukemia. *Leukemia* **2015**, *30*, 1033–1043. [[CrossRef](#)]
386. Kollmann, K.; Sexl, V. CDK6 and p16INK4A in lymphoid malignancies. *Oncotarget* **2013**, *4*, 1858–1859. [[CrossRef](#)]
387. Placke, T.; Faber, K.; Nonami, A.; Putwain, S.L.; Salih, H.R.; Heidel, F.H.; Krämer, A.; Root, D.E.; Barbie, D.A.; Krivtsov, A.V.; et al. Requirement for CDK6 in MLL-rearranged acute myeloid leukemia. *Blood* **2014**, *124*, 13–23. [[CrossRef](#)]
388. Van der Linden, M.; Willekes, M.; van Roon, E.; Seslija, L.; Schneider, P.; Pieters, R.; Stam, R. MLL fusion-driven activation of CDK6 potentiates proliferation in MLL-rearranged infant ALL. *Cell Cycle* **2014**, *13*, 834–844. [[CrossRef](#)]
389. Faussillon, M.; Monnier, L.; Junien, C.; Jeanpierre, C. Frequent overexpression of cyclin D2/cyclin-dependent kinase 4 in Wilms' tumor. *Cancer Lett.* **2005**, *221*, 67–75. [[CrossRef](#)]
390. Haruta, M.; Arai, Y.; Okita, H.; Tanaka, Y.; Takimoto, T.; Sugino, R.P.; Yamada, Y.; Kamijo, T.; Oue, T.; Fukuzawa, M.; et al. Combined Genetic and Chromosomal Characterization of Wilms Tumors Identifies Chromosome 12 Gain as a Potential New Marker Predicting a Favorable Outcome. *Neoplasia* **2018**, *21*, 117–131. [[CrossRef](#)]
391. Schubert, N.A.; Chen, C.Y.; Rodriguez, A.; Koster, J.; Dowless, M.; Pfister, S.M.; Shields, D.J.; Stancato, L.F.; Vassal, G.; Caron, H.N.; et al. Target actionability review to evaluate CDK4/6 as a therapeutic target in paediatric solid and brain tumours. *Eur. J. Cancer* **2022**, *170*, 196–208. [[CrossRef](#)]
392. Iwata, S.; Tatsumi, Y.; Yonemoto, T.; Araki, A.; Itami, M.; Kamoda, H.; Tsukanishi, T.; Hagiwara, Y.; Kinoshita, H.; Ishii, T.; et al. CDK4 overexpression is a predictive biomarker for resistance to conventional chemotherapy in patients with osteosarcoma. *Oncol. Rep.* **2021**, *46*, 1–11. [[CrossRef](#)]
393. Zhou, Y.; Shen, J.K.; Yu, Z.; Hornicek, F.J.; Kan, Q.; Duan, Z. Expression and therapeutic implications of cyclin-dependent kinase 4 (CDK4) in osteosarcoma. *Biochim. Biophys. Acta (BBA)-Mol. Basis Dis.* **2018**, *1864*, 1573–1582. [[CrossRef](#)]
394. Wunder, J.S.; Eppert, K.; Burrow, S.R.; Gogkoz, N.; Bell, R.S.; Andrulis, I.L. Co-amplification and overexpression of CDK4, SAS and MDM2 occurs frequently in human parosteal osteosarcomas. *Oncogene* **1999**, *18*, 783–788. [[CrossRef](#)] [[PubMed](#)]



395. Suehara, Y.; Alex, D.; Bowman, A.; Middha, S.; Zehir, A.; Chakravarty, D.; Wang, L.; Jour, G.; Nafa, K.; Hayashi, T.; et al. Clinical Genomic Sequencing of Pediatric and Adult Osteosarcoma Reveals Distinct Molecular Subsets with Potentially Targetable Alterations. *Clin. Cancer Res.* **2019**, *25*, 6346–6356. [[CrossRef](#)] [[PubMed](#)]
396. Guimarães, G.; Tesser-Gamba, F.; Petrilli, A.; Donato-Macedo, C.; Alves, M.; de Lima, F.; Garcia-Filho, R.; Oliveira, R.; Toledo, S. Molecular profiling of osteosarcoma in children and adolescents from different age groups using a next-generation sequencing panel. *Cancer Genet.* **2021**, *258–259*, 85–92. [[CrossRef](#)] [[PubMed](#)]
397. Hettmer, S.; Linardic, C.M.; Kelsey, A.; Rudzinski, E.R.; Vokuhl, C.; Selfe, J.; Ruhen, O.; Shern, J.F.; Khan, J.; Kovach, A.R.; et al. Molecular testing of rhabdomyosarcoma in clinical trials to improve risk stratification and outcome: A consensus view from European paediatric Soft tissue sarcoma Study Group, Children’s Oncology Group and Cooperative Weichteilsarkom-Studiengruppe. *Eur. J. Cancer* **2022**, *172*, 367–386. [[CrossRef](#)]
398. Barr, F.G.; Duan, F.; Smith, L.M.; Gustafson, D.; Pitts, M.; Hammond, S.; Gastier-Foster, J.M. Genomic and clinical analyses of 2p24 and 12q13-q14 amplification in alveolar rhabdomyosarcoma: A report from the Children’s Oncology Group. *Genes Chromosom. Cancer* **2009**, *48*, 661–672. [[CrossRef](#)]
399. de Andrade, C.R.; Jr, A.T.; Nishimoto, I.N.; Kowalski, L.P.; Lopes, M.A. Rhabdomyosarcoma of the head and neck: A clinicopathological and immunohistochemical analysis of 29 cases. *Braz. Dent. J.* **2010**, *21*, 68–73. [[CrossRef](#)]
400. Ragazzini, P.; Gamberi, G.; Pazzaglia, L.; Serra, M.; Magagnoli, G. Amplification of CDK4, MDM2, SAS and GLI genes in leiomyosarcoma, alveolar and embryonal rhabdomyosarcoma. *Histol. Histopathol.* **2004**, 401–411. [[CrossRef](#)]
401. Saab, R.; Bills, J.L.; Miceli, A.P.; Anderson, C.M.; Khoury, J.D.; Fry, D.W.; Navid, F.; Houghton, P.J.; Skapek, S.X. Pharmacologic inhibition of cyclin-dependent kinase 4/6 activity arrests proliferation in myoblasts and rhabdomyosarcoma-derived cells. *Mol. Cancer Ther.* **2006**, *5*, 1299–1308. [[CrossRef](#)]
402. Barghi, F.; Shannon, H.E.; Saadatzadeh, M.R.; Bailey, B.J.; Riyahi, N.; Bijangi-Vishehsaraei, K.; Just, M.; Ferguson, M.J.; Pandya, P.H.; Pollok, K.E. Precision Medicine Highlights Dysregulation of the CDK4/6 Cell Cycle Regulatory Pathway in Pediatric, Adolescents and Young Adult Sarcomas. *Cancers* **2022**, *14*, 3611. [[CrossRef](#)]
403. Kennedy, A.L.; Vallurupalli, M.; Chen, L.; Crompton, B.; Cowley, G.; Vazquez, F.; Weir, B.A.; Tsherniak, A.; Parasuraman, S.; Kim, S.; et al. Functional, chemical genomic, and super-enhancer screening identify sensitivity to cyclin D1/CDK4 pathway inhibition in Ewing sarcoma. *Oncotarget* **2015**, *6*, 30178–30193. [[CrossRef](#)]
404. Molenaar, J.J.; Ebus, M.E.; Koster, J.; van Sluis, P.; van Noesel, C.J.; Versteeg, R.; Caron, H.N. Cyclin D1 and CDK4 Activity Contribute to the Undifferentiated Phenotype in Neuroblastoma. *Cancer Res* **2008**, *68*, 2599–2609. [[CrossRef](#)]
405. Rader, J.; Russell, M.R.; Hart, L.S.; Nakazawa, M.S.; Belcastro, L.T.; Martinez, D.; Li, Y.; Carpenter, E.L.; Attiyeh, E.F.; Diskin, S.J.; et al. Dual CDK4/CDK6 Inhibition Induces Cell-Cycle Arrest and Senescence in Neuroblastoma. *Clin. Cancer Res.* **2013**, *19*, 6173–6182. [[CrossRef](#)]
406. Amoroso, L.; Ognibene, M.; Morini, M.; Conte, M.; Di Cataldo, A.; Tondo, A.; D’Angelo, P.; Castellano, A.; Garaventa, A.; Lasorsa, V.A.; et al. Genomic coamplification of CDK4/MDM2/FRS2 is associated with very poor prognosis and atypical clinical features in neuroblastoma patients. *Genes Chromosom. Cancer* **2019**, *59*, 277–285. [[CrossRef](#)]
407. Huang, W.; Hao, Z.; Mao, F.; Guo, D. Small Molecule Inhibitors in Adult High-Grade Glioma: From the Past to the Future. *Front. Oncol.* **2022**, *12*, 911876. [[CrossRef](#)]
408. Liu, A.; Zhao, H.; Sun, B.; Han, X.; Zhou, D.; Cui, Z.; Ma, X.; Zhang, J.; Yuan, L. A predictive analysis approach for paediatric and adult high-grade glioma: miRNAs and network insight. *Ann. Transl. Med.* **2020**, *8*, 242. [[CrossRef](#)]
409. Rallis, K.S.; George, A.M.; Wozniak, A.M.; Bigogno, C.M.; Chow, B.; Hanrahan, J.G.; Sideris, M. Molecular Genetics and Targeted Therapies for Paediatric High-grade Glioma. *Cancer Genom.-Proteom.* **2022**, *19*, 390–414. [[CrossRef](#)]
410. Sepúlveda-Sánchez, J.M.; Gil-Gil, M.; Alonso-García, M.; Salgado, M.V.; Vicente, E.; Barroso, C.M.; Sánchez, R.; Durán, G.; Peñas, R.D.L.; Muñoz-Langa, J.; et al. Phase II Trial of Palbociclib in Recurrent Retinoblastoma-Positive Anaplastic Oligodendroglioma: A Study from the Spanish Group for Research in Neuro-Oncology (GEINO). *Target. Oncol.* **2020**, *15*, 613–622. [[CrossRef](#)]
411. Zangen, I.L.; Kneitz, S.; Monoranu, C.-M.; Rutkowski, S.; Hinkes, B.; Vince, G.H.; Huang, B.; Roggendorf, W. Ependymoma gene expression profiles associated with histological subtype, proliferation, and patient survival. *Acta Neuropathol.* **2007**, *113*, 325–337. [[CrossRef](#)]
412. Liang, M.-L.; Chen, C.-H.; Liu, Y.-R.; Huang, M.-H.; Lin, Y.-C.; Wong, T.-T.; Lin, S.-E.; Chu, S.-S.; Ding, Y.-H.; Hsieh, T.-H. Abemaciclib, A Selective CDK4/6 Inhibitor, Restricts the Growth of Pediatric Ependymomas. *Cancers* **2020**, *12*, 3597. [[CrossRef](#)]
413. Magalhães, T.D.A.; Cruzeiro, G.A.V.; de Sousa, G.R.; da Silva, K.R.; Lira, R.C.P.; Scrideli, C.A.; Tone, L.G.; Valera, E.T.; Borges, K.S. Notch pathway in ependymoma RELA-fused subgroup: Upregulation and association with cancer stem cells markers expression. *Cancer Gene Ther.* **2019**, *27*, 509–512. [[CrossRef](#)]
414. Lummus, S.C.; Donson, A.M.; Gowan, K.; Jones, K.L.; Vibhakar, R.; Foreman, N.K.; Kleinschmidt-DeMasters, B.K. p16Loss and E2F/cell cycle deregulation in infant posterior fossa ependymoma. *Pediatr. Blood Cancer* **2017**, *64*, e26656. [[CrossRef](#)] [[PubMed](#)]
415. Shupp, A.; Casimiro, M.C.; Pestell, R.G. Biological functions of CDK5 and potential CDK5 targeted clinical treatments. *Oncotarget* **2017**, *8*, 17373–17382. [[CrossRef](#)] [[PubMed](#)]
416. Sharma, S.; Sicinski, P. A kinase of many talents: Non-neuronal functions of CDK5 in development and disease. *Open Biol.* **2020**, *10*, 190287. [[CrossRef](#)] [[PubMed](#)]
417. Shah, K.; Rossie, S. Tale of the Good and the Bad Cdk5: Remodeling of the Actin Cytoskeleton in the Brain. *Mol. Neurobiol.* **2017**, *55*, 3426–3438. [[CrossRef](#)]



418. Liu, W.; Li, J.; Song, Y.-S.; Li, Y.; Jia, Y.-H.; Zhao, H.-D. Cdk5 links with DNA damage response and cancer. *Mol. Cancer* **2017**, *16*, 60. [[CrossRef](#)]
419. Liu, S.-L.; Wang, C.; Jiang, T.; Tan, L.; Xing, A.; Yu, J.-T. The Role of Cdk5 in Alzheimer's Disease. *Mol. Neurobiol.* **2015**, *53*, 4328–4342. [[CrossRef](#)]
420. Liang, Q.; Li, L.; Zhang, J.; Lei, Y.; Wang, L.; Liu, D.-X.; Feng, J.; Hou, P.; Yao, R.; Zhang, Y.; et al. CDK5 is essential for TGF- $\beta$ 1-induced epithelial-mesenchymal transition and breast cancer progression. *Sci. Rep.* **2013**, *3*, 2932. [[CrossRef](#)]
421. Zeng, J.; Xie, S.; Liu, Y.; Shen, C.; Song, X.; Zhou, G.-L.; Wang, C. CDK5 Functions as a Tumor Promoter in Human Lung Cancer. *J. Cancer* **2018**, *9*, 3950–3961. [[CrossRef](#)]
422. Lin, H.; Lin, T.-Y.; Juang, J.-L. Abl deregulates Cdk5 kinase activity and subcellular localization in *Drosophila* neurodegeneration. *Cell Death Differ.* **2006**, *14*, 607–615. [[CrossRef](#)]
423. de Porras, V.R.; Bystrup, S.; Heras, S.C.-D.L.; Musulén, E.; Palomero, L.; Alonso, M.H.; Nieto, R.; Arango, D.; Moreno, V.; Queralt, C.; et al. Tumor Expression of Cyclin-Dependent Kinase 5 (Cdk5) Is a Prognostic Biomarker and Predicts Outcome of Oxaliplatin-Treated Metastatic Colorectal Cancer Patients. *Cancers* **2019**, *11*, 1540. [[CrossRef](#)]
424. Zhou, Y.; Wang, X.; Lv, P.; Yu, H.; Jiang, X. CDK5 Knockdown inhibits proliferation and induces apoptosis and Cell Cycle Arrest in Human Glioblastoma. *J. Cancer* **2021**, *12*, 3958–3966. [[CrossRef](#)]
425. Lin, H.; Chen, M.-C.; Chiu, C.-Y.; Song, Y.-M.; Lin, S.-Y. Cdk5 Regulates STAT3 Activation and Cell Proliferation in Medullary Thyroid Carcinoma Cells. *J. Biol. Chem.* **2007**, *282*, 2776–2784. [[CrossRef](#)]
426. Oner, M.; Lin, E.; Chen, M.-C.; Hsu, F.-N.; Prince, G.M.S.H.; Chiu, K.-Y.; Teng, C.-L.J.; Yang, T.-Y.; Wang, H.-Y.; Yue, C.-H.; et al. Future Aspects of CDK5 in Prostate Cancer: From Pathogenesis to Therapeutic Implications. *Int. J. Mol. Sci.* **2019**, *20*, 3881. [[CrossRef](#)]
427. Huang, P.-H.; Chen, M.-C.; Peng, Y.-T.; Kao, W.-H.; Chang, C.-H.; Wang, Y.-C.; Lai, C.-H.; Hsieh, J.-T.; Wang, J.-H.; Lee, Y.-T.; et al. Cdk5 Directly Targets Nuclear p21CIP1 and Promotes Cancer Cell Growth. *Cancer Res* **2016**, *76*, 6888–6900. [[CrossRef](#)]
428. Sun, Y.-Q.; Xie, J.-W.; Xie, H.-T.; Chen, P.-C.; Zhang, X.-L.; Zheng, C.-H.; Li, P.; Wang, J.-B.; Lin, J.-X.; Cao, L.-L.; et al. Expression of CRM1 and CDK5 shows high prognostic accuracy for gastric cancer. *World J. Gastroenterol.* **2017**, *23*, 2012–2022. [[CrossRef](#)]
429. Kour, S.; Rana, S.; Contreras, J.I.; King, H.M.; Robb, C.M.; Sonawane, Y.A.; Bendjennat, M.; Crawford, A.J.; Barger, C.J.; Kizhake, S.; et al. CDK5 Inhibitor Downregulates Mcl-1 and Sensitizes Pancreatic Cancer Cell Lines to Navitoclax. *Mol. Pharmacol.* **2019**, *96*, 419–429. [[CrossRef](#)]
430. Mukherjee, S.; Tucker-Burden, C.; Kaissi, E.; Newsam, A.; Duggireddy, H.; Chau, M.; Zhang, C.; Diwedi, B.; Rupji, M.; Seby, S.; et al. CDK5 Inhibition Resolves PKA/cAMP-Independent Activation of CREB1 Signaling in Glioma Stem Cells. *Cell Rep.* **2018**, *23*, 1651–1664. [[CrossRef](#)]
431. Do, P.A.; Lee, C.H. The Role of CDK5 in Tumours and Tumour Microenvironments. *Cancers* **2020**, *13*, 101. [[CrossRef](#)]
432. Gao, G.-B.; Sun, Y.; Fang, R.-D.; Wang, Y.; Wang, Y.; He, Q.-Y. Post-translational modifications of CDK5 and their biological roles in cancer. *Mol. Biomed.* **2021**, *2*, 1–15. [[CrossRef](#)]
433. Peyressatre, M.; Laure, A.; Pellerano, M.; Boukhaddaoui, H.; Soussi, I.; Morris, M.C. Fluorescent Biosensor of CDK5 Kinase Activity in Glioblastoma Cell Extracts and Living Cells. *Biotechnol. J.* **2020**, *15*, e1900474. [[CrossRef](#)]
434. Binlath, T.; Reudhabadh, R.; Prommeenat, P.; Hutamekalin, P. Investigation of mechanisms underlying the inhibitory effects of metformin against proliferation and growth of neuroblastoma SH-SY5Y cells. *Toxicol. Vitro.* **2022**, *83*, 105410. [[CrossRef](#)] [[PubMed](#)]
435. Yushan, R.; Wenjie, C.; Suning, H.; Yiwu, D.; Tengfei, Z.; Madushi, W.M.; Feifei, L.; Changwen, Z.; Xin, W.; Roodrajeetsing, G.; et al. Insights into the clinical value of cyclin-dependent kinase 5 in glioma: A retrospective study. *World J. Surg. Oncol.* **2015**, *13*, 223. [[CrossRef](#)] [[PubMed](#)]
436. Catania, A.; Urban, S.; Yan, E.; Hao, C.; Barron, G.; Allalunis-Turner, J. Expression and localization of cyclin-dependent kinase 5 in apoptotic human glioma cells. *Neuro-Oncology* **2001**, *3*, 89–98. [[CrossRef](#)] [[PubMed](#)]
437. de Nigris, F.; Mancini, F.P.; Schiano, C.; Infante, T.; Zullo, A.; Minucci, P.B.; Al-Omran, M.; Giordano, A.; Napoli, C. Osteosarcoma cells induce endothelial cell proliferation during neo-angiogenesis. *J. Cell. Physiol.* **2012**, *228*, 846–852. [[CrossRef](#)] [[PubMed](#)]
438. Bao, H.-X.; Bi, Q.; Han, Y.; Zhao, C.; Zou, H. Potential mechanisms underlying CDK5 related Osteosarcoma progression. *Expert Opin. Ther. Targets* **2017**, *21*, 455–460. [[CrossRef](#)]
439. Fu, H.; Zhao, H.; Yang, Y.; Duan, K.; Guo, T. CDK5 Inhibitor Seliciclib Promotes Osteoblastic Differentiation of MSCs and Suppresses the Migration of MG-63 Osteosarcoma Cells. *BioRxiv* **2020**. [[CrossRef](#)]
440. Saidak, Z.; Le Henaff, C.; Azzi, S.; Marty, C.; Da Nascimento, S.; Sonnet, P.; Marie, P.J. Wnt/ $\beta$ -Catenin Signaling Mediates Osteoblast Differentiation Triggered by Peptide-induced  $\alpha$ 5 $\beta$ 1 Integrin Priming in Mesenchymal Skeletal Cells. *J. Biol. Chem.* **2015**, *290*, 6903–6912. [[CrossRef](#)]
441. Schwalbe, E.C.; Lindsey, J.C.; Nakjang, S.; Crosier, S.; Smith, A.J.; Hicks, D.; Rafiee, G.; Hill, R.M.; Iliasova, A.; Stone, T.; et al. Novel molecular subgroups for clinical classification and outcome prediction in childhood medulloblastoma: A cohort study. *Lancet Oncol.* **2017**, *18*, 958–971. [[CrossRef](#)]
442. Dorand, R.D.; Nthale, J.; Myers, J.T.; Barkauskas, D.S.; Avril, S.; Chirieleison, S.M.; Pareek, T.K.; Abbott, D.W.; Stearns, D.S.; Letterio, J.J.; et al. Cdk5 disruption attenuates tumor PD-L1 expression and promotes antitumor immunity. *Science* **2016**, *353*, 399–403. [[CrossRef](#)]
443. Sava, G.P.; Fan, H.; Coombes, R.C.; Buluwela, L.; Ali, S. CDK7 inhibitors as anticancer drugs. *Cancer Metastasis Rev.* **2020**, *39*, 805–823. [[CrossRef](#)]

444. Schachter, M.M.; Fisher, R.P. The CDK-activating kinase Cdk7: Taking yes for an answer. *Cell Cycle* **2013**, *12*, 3239–3240. [[CrossRef](#)]
445. Chipumuro, E.; Marco, E.; Christensen, C.L.; Kwiatkowski, N.; Zhang, T.; Hatheway, C.M.; Abraham, B.J.; Sharma, B.; Yeung, C.; Altabef, A.; et al. CDK7 Inhibition Suppresses Super-Enhancer-Linked Oncogenic Transcription in MYCN-Driven Cancer. *Cell* **2014**, *159*, 1126–1139. [[CrossRef](#)]
446. Fisher, R.P. Cdk7: A kinase at the core of transcription and in the crosshairs of cancer drug discovery. *Transcription* **2018**, *10*, 47–56. [[CrossRef](#)]
447. Bacon, C.W.; D’Orso, I. CDK9: A signaling hub for transcriptional control. *Transcription* **2018**, *10*, 57–75. [[CrossRef](#)]
448. Zhang, H.; Pandey, S.; Travers, M.; Sun, H.; Morton, G.; Madzo, J.; Chung, W.; Khowsathit, J.; Perez-Leal, O.; Barrero, C.A.; et al. Targeting CDK9 Reactivates Epigenetically Silenced Genes in Cancer. *Cell* **2018**, *175*, 1244–1258.e26. [[CrossRef](#)]
449. Patel, H.; Abduljabbar, R.; Lai, C.-F.; Periyasamy, M.; Harrod, A.; Gemma, C.; Steel, J.H.; Patel, N.; Busonero, C.; Jerjees, D.; et al. Expression of CDK7, Cyclin H, and MAT1 Is Elevated in Breast Cancer and Is Prognostic in Estrogen Receptor-Positive Breast Cancer. *Clin. Cancer Res.* **2016**, *22*, 5929–5938. [[CrossRef](#)]
450. Naseh, G.; Mohammadifard, M. Upregulation of cyclin-dependent kinase 7 and matrix metalloproteinase-14 expression contribute to metastatic properties of gastric cancer. *IUBMB Life* **2016**, *68*, 799–805. [[CrossRef](#)]
451. Jagomast, T.; Idel, C.; Klapper, L.; Kuppler, P.; Offermann, A.; Dreyer, E.; Bruchhage, K.-L.; Ribbat-Idel, J.; Perner, S. CDK7 Predicts Worse Outcome in Head and Neck Squamous-Cell Cancer. *Cancers* **2022**, *14*, 492. [[CrossRef](#)]
452. Kim, J.; Cho, Y.-J.; Ryu, J.-Y.; Hwang, I.; Han, H.D.; Ahn, H.J.; Kim, W.Y.; Cho, H.; Chung, J.-Y.; Hewitt, S.M.; et al. CDK7 is a reliable prognostic factor and novel therapeutic target in epithelial ovarian cancer. *Gynecol. Oncol.* **2020**, *156*, 211–221. [[CrossRef](#)]
453. Tang, L.; Zhu, C.; Jin, J.; Wang, X.; Yu, L.; Guan, X. Expression of CDK7 correlates with molecular subtypes and predicts clinical outcomes in breast cancer. *Transl. Cancer Res.* **2021**, *10*, 669–680. [[CrossRef](#)]
454. Kretz, A.-L.; Schaum, M.; Richter, J.; Kitzig, E.F.; Engler, C.C.; Leithäuser, F.; Henne-Bruns, D.; Knippschild, U.; Lemke, J. CDK9 is a prognostic marker and therapeutic target in pancreatic cancer. *Tumor Biol.* **2017**, *39*. [[CrossRef](#)] [[PubMed](#)]
455. Yang, W.; Liu, S.; Luo, Q.; Tan, X. Expression of CDK9 in endometrial cancer tissues and its effect on the proliferation of HEC-1B. *Open Life Sci.* **2021**, *16*, 1341–1346. [[CrossRef](#)] [[PubMed](#)]
456. Rasool, R.U.; Natesan, R.; Deng, Q.; Aras, S.; Lal, P.; Effron, S.S.; Mitchell-Velasquez, E.; Posimo, J.M.; Carskadon, S.; Baca, S.C.; et al. CDK7 Inhibition Suppresses Castration-Resistant Prostate Cancer through MED1 Inactivation. *Cancer Discov.* **2019**, *9*, 1538–1555. [[CrossRef](#)] [[PubMed](#)]
457. Gao, X.; Liang, J.; Wang, L.; Zhang, Z.; Yuan, P.; Wang, J.; Gao, Y.; Ma, F.; Calagua, C.; Ye, H.; et al. Phosphorylation of the androgen receptor at Ser81 is co-sustained by CDK1 and CDK9 and leads to AR-mediated transactivation in prostate cancer. *Mol. Oncol.* **2021**, *15*, 1901–1920. [[CrossRef](#)] [[PubMed](#)]
458. Nagaraja, S.; Vitanza, N.A.; Woo, P.J.; Taylor, K.R.; Liu, F.; Zhang, L.; Li, M.; Meng, W.; Ponnuswami, A.; Sun, W.; et al. Transcriptional Dependencies in Diffuse Intrinsic Pontine Glioma. *Cancer Cell* **2017**, *31*, 635–652.e6. [[CrossRef](#)]
459. Ma, H.; Dean, D.C.; Wei, R.; Hornicek, F.J.; Duan, Z. Cyclin-dependent kinase 7 (CDK7) is an emerging prognostic biomarker and therapeutic target in osteosarcoma. *Ther. Adv. Musculoskelet. Dis.* **2021**, *13*. [[CrossRef](#)]
460. Zhang, J.; Liu, W.; Zou, C.; Zhao, Z.; Lai, Y.; Shi, Z.; Xie, X.; Huang, G.; Wang, Y.; Zhang, X.; et al. Targeting Super-Enhancer-Associated Oncogenes in Osteosarcoma with THZ2, a Covalent CDK7 Inhibitor. *Clin. Cancer Res.* **2020**, *26*, 2681–2692. [[CrossRef](#)]
461. Ma, H.; Seebacher, N.A.; Hornicek, F.J.; Duan, Z. Cyclin-dependent kinase 9 (CDK9) is a novel prognostic marker and therapeutic target in osteosarcoma. *Ebiomedicine* **2018**, *39*, 182–193. [[CrossRef](#)]
462. Qin, J.-J. Is CDK9 a promising target for both primary and metastatic osteosarcoma? *Ebiomedicine* **2019**, *40*, 27–28. [[CrossRef](#)]
463. Iniguez, A.B.; Stolte, B.; Wang, E.J.; Conway, A.S.; Alexe, G.; Dharia, N.V.; Kwiatkowski, N.; Zhang, T.; Abraham, B.J.; Mora, J.; et al. EWS/FLI Confers Tumor Cell Synthetic Lethality to CDK12 Inhibition in Ewing Sarcoma. *Cancer Cell* **2018**, *33*, 202–216.e6. [[CrossRef](#)]
464. Ning, J.; Ma, X.; Long, C.; Mao, Y.; Kuang, X.; Huang, Z.; Fan, Y.; Zhang, H.; Xia, Q.; Wang, R.; et al. Anti-tumor Drug THZ1 Suppresses TGFβ2-mediated EMT in Lens Epithelial Cells via Notch and TGFβ/Smad Signaling Pathway. *J. Cancer* **2019**, *10*, 3778–3788. [[CrossRef](#)]
465. Cassandri, M.; Fioravanti, R.; Pomella, S.; Valente, S.; Rotili, D.; Del Baldo, G.; De Angelis, B.; Rota, R.; Mai, A. CDK9 as a Valuable Target in Cancer: From Natural Compounds Inhibitors to Current Treatment in Pediatric Soft Tissue Sarcomas. *Front. Pharmacol.* **2020**, *11*, 1230. [[CrossRef](#)]
466. Simone, C.; Giordano, A. Abrogation of signal-dependent activation of the cdk9/cyclin T2a complex in human RD rhabdomyosarcoma cells. *Cell Death Differ.* **2006**, *14*, 192–195. [[CrossRef](#)]
467. Richter, G.H.; Hensel, T.; Schmidt, O.; Saratov, V.; von Heyking, K.; Becker-Dettling, F.; Prexler, C.; Yen, H.-Y.; Steiger, K.; Fulda, S.; et al. Combined Inhibition of Epigenetic Readers and Transcription Initiation Targets the EWS-ETS Transcriptional Program in Ewing Sarcoma. *Cancers* **2020**, *12*, 304. [[CrossRef](#)]
468. De Falco, G.; Bellan, C.; D’Amuri, A.; Angeloni, G.; Leucci, E.; Giordano, A.; Leoncini, L. Cdk9 regulates neural differentiation and its expression correlates with the differentiation grade of neuroblastoma and PNET tumors. *Cancer Biol. Ther.* **2005**, *4*, 277–281. [[CrossRef](#)]
469. Poon, E.; Liang, T.; Jamin, Y.; Walz, S.; Kwok, C.; Hakkert, A.; Barker, K.; Urban, Z.; Thway, K.; Zeid, R.; et al. Orally bioavailable CDK9/2 inhibitor shows mechanism-based therapeutic potential in MYCN-driven neuroblastoma. *J. Clin. Investig.* **2020**, *130*, 5875–5892. [[CrossRef](#)]

470. Hamanaka, R.; Maloid, S.; Smith, M.R.; O'Connell, C.D.; Longo, D.L.; Ferris, D.K. Cloning and characterization of human and murine homologues of the Drosophila polo serine-threonine kinase. *Cell Growth Differ.* **1994**, *5*, 249–257.
471. Johnson, E.F.; Stewart, K.; Woods, K.W.; Giranda, V.L.; Luo, Y. Pharmacological and Functional Comparison of the Polo-like Kinase Family: Insight into Inhibitor and Substrate Specificity. *Biochemistry* **2007**, *46*, 9551–9563. [[CrossRef](#)]
472. Cizmecioglu, O.; Warnke, S.; Arnold, M.; Duensing, S.; Hoffmann, I. Plk2 regulated centriole duplication is dependent on its localization to the centrosome and a functional polo-box domain. *Cell Cycle* **2008**, *7*, 3548–3555. [[CrossRef](#)]
473. Archambault, V.; Glover, D. Polo-like kinases: Conservation and divergence in their functions and regulation. *Nat. Rev. Mol. Cell Biol.* **2009**, *10*, 265–275. [[CrossRef](#)]
474. Glover, D.M.; Hagan, I.M.; Tavares, A. Polo-like kinases: A team that plays throughout mitosis. *Genes Dev.* **1998**, *12*, 3777–3787. [[CrossRef](#)]
475. Barr, F.; Silljé, H.H.W.; Nigg, E. Polo-like kinases and the orchestration of cell division. *Nat. Rev. Mol. Cell Biol.* **2004**, *5*, 429–441. [[CrossRef](#)] [[PubMed](#)]
476. A Winkles, J.; Alberts, G.F. Differential regulation of polo-like kinase 1, 2, 3, and 4 gene expression in mammalian cells and tissues. *Oncogene* **2005**, *24*, 260–266. [[CrossRef](#)] [[PubMed](#)]
477. Iliaki, S.; Beyaert, R.; Afonina, I.S. Polo-like kinase 1 (PLK1) signaling in cancer and beyond. *Biochem. Pharmacol.* **2021**, *193*, 114747. [[CrossRef](#)] [[PubMed](#)]
478. Raab, C.A.; Raab, M.; Becker, S.; Strebhardt, K. Non-mitotic functions of polo-like kinases in cancer cells. *Biochim. Biophys. Acta (BBA)-Rev. Cancer* **2020**, *1875*, 188467. [[CrossRef](#)]
479. Cholewa, B.D.; Liu, X.; Ahmad, N. The Role of Polo-like Kinase 1 in Carcinogenesis: Cause or Consequence? *Cancer Res* **2013**, *73*, 6848–6855. [[CrossRef](#)]
480. Strebhardt, K. Multifaceted polo-like kinases: Drug targets and antitargets for cancer therapy. *Nat. Rev. Drug Discov.* **2010**, *9*, 643–660. [[CrossRef](#)]
481. Pellegrino, R.; Calvisi, D.F.; Ladu, S.; Ehemann, V.; Staniscia, T.; Evert, M.; Dombrowski, F.; Schirmacher, P.; Longerich, T. Oncogenic and tumor suppressive roles of polo-like kinases in human hepatocellular carcinoma. *Hepatology* **2009**, *51*, 857–868. [[CrossRef](#)]
482. Degenhardt, Y.; Lampkin, T. Targeting Polo-like kinase in cancer therapy. *Clin. Cancer Res. Off. J. Am. Assoc. Cancer Res.* **2010**, *16*, 384–389. [[CrossRef](#)]
483. Ando, K.; Ozaki, T.; Yamamoto, H.; Furuya, K.; Hosoda, M.; Hayashi, S.; Fukuzawa, M.; Nakagawara, A. Polo-like Kinase 1 (Plk1) Inhibits p53 Function by Physical Interaction and Phosphorylation. *J. Biol. Chem.* **2004**, *279*, 25549–25561. [[CrossRef](#)]
484. Li, S.; Li, H.; Cao, Y.; Geng, H.; Ren, F.; Li, K.; Dai, C.; Li, N. Integrated bioinformatics analysis reveals CDK1 and PLK1 as potential therapeutic targets of lung adenocarcinoma. *Medicine* **2021**, *100*, e26474. [[CrossRef](#)]
485. Li, H.; Wang, H.; Sun, Z.; Guo, Q.; Shi, H.; Jia, Y. The clinical and prognostic value of polo-like kinase 1 in lung squamous cell carcinoma patients: Immunohistochemical analysis. *Biosci. Rep.* **2017**, *37*. [[CrossRef](#)]
486. Ramani, P.; Nash, R.; Sowa-Avugrah, E.; A Rogers, C. High levels of polo-like kinase 1 and phosphorylated translationally controlled tumor protein indicate poor prognosis in neuroblastomas. *J. Neuro-Oncology* **2015**, *125*, 103–111. [[CrossRef](#)]
487. Harris, P.S.; Venkataraman, S.; Alimova, I.; Birks, D.K.; Donson, A.M.; Knipstein, J.; Dubuc, A.; Taylor, M.D.; Handler, M.H.; Foreman, N.K.; et al. Polo-like kinase 1 (PLK1) inhibition suppresses cell growth and enhances radiation sensitivity in medulloblastoma cells. *BMC Cancer* **2012**, *12*, 80. [[CrossRef](#)]
488. Triscott, J.; Lee, C.; Foster, C.; Manoranjan, B.; Pambid, M.R.; Berns, R.; Fotovati, A.; Venugopal, C.; O'Halloran, K.; Narendran, A.; et al. Personalizing the Treatment of Pediatric Medulloblastoma: Polo-like Kinase 1 as a Molecular Target in High-Risk Children. *Cancer Res* **2013**, *73*, 6734–6744. [[CrossRef](#)]
489. Pezuk, J.A.; Brassescio, M.S.; de Oliveira, R.S.; Machado, H.R.; Neder, L.; Scrideli, C.A.; Tone, L.G. PLK1-associated microRNAs are correlated with pediatric medulloblastoma prognosis. *Child's Nerv. Syst.* **2017**, *33*, 609–615. [[CrossRef](#)]
490. Ma, H.; Nie, C.; Chen, Y.; Li, J.; Xie, Y.; Tang, Z.; Gao, Y.; Ai, S.; Mao, Y.; Sun, Q.; et al. Therapeutic Targeting PLK1 by ON-01910.Na Is Effective in Local Treatment of Retinoblastoma. *Oncol. Res. Featur. Preclin. Clin. Cancer Ther.* **2021**, *28*, 745–761. [[CrossRef](#)]
491. Singh, L.; Pushker, N.; Sen, S.; Singh, M.K.; A Chauhan, F.; Kashyap, S. Prognostic significance of polo-like kinases in retinoblastoma: Correlation with patient outcome, clinical and histopathological parameters. *Clin. Exp. Ophthalmol.* **2015**, *43*, 550–557. [[CrossRef](#)]
492. Ackermann, S.; Goeser, F.; Schulte, J.H.; Schramm, A.; Ehemann, V.; Hero, B.; Eggert, A.; Berthold, F.; Fischer, M. Polo-Like Kinase 1 is a Therapeutic Target in High-Risk Neuroblastoma. *Clin. Cancer Res.* **2011**, *17*, 731–741. [[CrossRef](#)]
493. Mo, H.; He, J.; Yuan, Z.; Wu, Z.; Liu, B.; Lin, X.; Guan, J. PLK1 contributes to autophagy by regulating MYC stabilization in osteosarcoma cells. *Oncotargets Ther.* **2019**, *12*, 7527–7536. [[CrossRef](#)]
494. Mountzios, G.; Terpos, E.; Dimopoulos, M. Aurora kinases as targets for cancer therapy. *Cancer Treat. Rev.* **2008**, *34*, 175–182. [[CrossRef](#)] [[PubMed](#)]
495. Goldenson, B.; Crispino, J.D. The aurora kinases in cell cycle and leukemia. *Oncogene* **2014**, *34*, 537–545. [[CrossRef](#)] [[PubMed](#)]
496. Toji, S.; Yabuta, N.; Hosomi, T.; Nishihara, S.; Kobayashi, T.; Suzuki, S.; Tamai, K.; Nojima, H. The centrosomal protein Lats2 is a phosphorylation target of Aurora-A kinase. *Genes Cells* **2004**, *9*, 383–397. [[CrossRef](#)]
497. Naso, F.D.; Boi, D.; Ascanelli, C.; Pamfil, G.; Lindon, C.; Paiardini, A.; Guarguaglini, G. Nuclear localisation of Aurora-A: Its regulation and significance for Aurora-A functions in cancer. *Oncogene* **2021**, *40*, 3917–3928. [[CrossRef](#)] [[PubMed](#)]



498. Marumoto, T.; Honda, S.; Hara, T.; Nitta, M.; Hirota, T.; Kohmura, E.; Saya, H. Aurora-A Kinase Maintains the Fidelity of Early and Late Mitotic Events in HeLa Cells. *J. Biol. Chem.* **2003**, *278*, 51786–51795. [[CrossRef](#)]
499. Crane, R.; Gadea, B.; Littlepage, L.; Wu, H.; Ruderman, J.V. Aurora A, Meiosis and Mitosis. *Biol. Cell* **2004**, *96*, 215–229. [[CrossRef](#)]
500. Musacchio, A.; Hardwick, K.G. The spindle checkpoint: Structural insights into dynamic signalling. *Nat. Rev. Mol. Cell Biol.* **2002**, *3*, 731–741. [[CrossRef](#)]
501. Werner, M.; Mattis, A.; Aubele, M.; Cummings, M.; Zitzelsberger, H.; Hutzler, P.; Höfler, H. 20q13.2 Amplification in intraductal hyperplasia adjacent to in situ and invasive ductal carcinoma of the breast. *Virchows Arch.* **1999**, *435*, 469–472. [[CrossRef](#)]
502. Bui, V.-M.; Mettling, C.; Jou, J.; Sun, H.S. Genomic amplification of chromosome 20q13.33 is the early biomarker for the development of sporadic colorectal carcinoma. *BMC Med. Genom.* **2020**, *13*, 149. [[CrossRef](#)]
503. Tanner, M.M.; Tirkkonen, M.; Kallioniemi, A.; Holli, K.; Collins, C.; Kowbel, D.; Gray, J.W.; Kallioniemi, O.; Isola, J. Amplification of chromosomal region 20q13 in invasive breast cancer: Prognostic implications. *Clin. Cancer Res.* **1995**, *1*, 1455–1461.
504. Mou, P.K.; Yang, E.J.; Shi, C.; Ren, G.; Tao, S.; Shim, J.S. Aurora kinase A, a synthetic lethal target for precision cancer medicine. *Exp. Mol. Med.* **2021**, *53*, 835–847. [[CrossRef](#)]
505. Murga-Zamalloa, C.; Inamdar, K.V.; Wilcox, R.A. The role of aurora A and polo-like kinases in high-risk lymphomas. *Blood Adv.* **2019**, *3*, 1778–1787. [[CrossRef](#)]
506. Bast, R.C., Jr.; Hennessy, B.; Mills, G.B. The biology of ovarian cancer: New opportunities for translation. *Nat. Rev. Cancer* **2009**, *9*, 415–428. [[CrossRef](#)]
507. Sun, H.; Wang, H.; Wang, X.; Aoki, Y.; Wang, X.; Yang, Y.; Cheng, X.; Wang, Z.; Wang, X. Aurora-A/SOX8/FOXK1 signaling axis promotes chemoresistance via suppression of cell senescence and induction of glucose metabolism in ovarian cancer organoids and cells. *Theranostics* **2020**, *10*, 6928–6945. [[CrossRef](#)]
508. Wan, X.-B.; Long, Z.-J.; Yan, M.; Xu, J.; Xia, L.-P.; Liu, L.; Zhao, Y.; Huang, X.-F.; Wang, X.-R.; Zhu, X.-F.; et al. Inhibition of Aurora-A suppresses epithelial–mesenchymal transition and invasion by downregulating MAPK in nasopharyngeal carcinoma cells. *Carcinog.* **2008**, *29*, 1930–1937. [[CrossRef](#)]
509. Nguyen, T.T.B.; Silva, F.N.M.; Golemis, E.A. Aurora Kinases as Therapeutic Targets in Head and Neck Cancer. *Cancer J.* **2022**, *28*, 387–400. [[CrossRef](#)]
510. Do, T.-V.; Xiao, F.; E Bickel, L.; Klein-Szanto, A.J.; Pathak, H.B.; Hua, X.; Howe, C.; O'Brien, S.W.; Maglady, M.; A Ecsedy, J.; et al. Aurora kinase A mediates epithelial ovarian cancer cell migration and adhesion. *Oncogene* **2013**, *33*, 539–549. [[CrossRef](#)]
511. Katayama, H.; Wang, J.; Treekitkarnmongkol, W.; Kawai, H.; Sasai, K.; Zhang, H.; Wang, H.; Adams, H.P.; Jiang, S.; Chakraborty, S.N.; et al. Aurora Kinase-A Inactivates DNA Damage-Induced Apoptosis and Spindle Assembly Checkpoint Response Functions of p73. *Cancer Cell* **2012**, *21*, 196–211. [[CrossRef](#)]
512. Anand, S.; Penrhyn-Lowe, S.; Venkitaraman, A.R. AURORA-A amplification overrides the mitotic spindle assembly checkpoint, inducing resistance to Taxol. *Cancer Cell* **2003**, *3*, 51–62. [[CrossRef](#)]
513. Cirak, Y.; Furuncuoglu, Y.; Yapicier, O.; Aksu, A.; Cubukcu, E. Aurora A overexpression in breast cancer patients induces taxane resistance and results in worse prognosis. *J. BUON.* **2016**, *20*, 1414–1419.
514. Reiter, R.; Gais, P.; Jütting, U.; Steuer-Vogt, M.K.; Pickhard, A.; Bink, K.; Rauser, S.; Lassmann, S.; Höfler, H.; Werner, M.; et al. Aurora Kinase A Messenger RNA Overexpression Is Correlated with Tumor Progression and Shortened Survival in Head and Neck Squamous Cell Carcinoma. *Clin. Cancer Res.* **2006**, *12*, 5136–5141. [[CrossRef](#)] [[PubMed](#)]
515. Noh, E.-M.; Lee, Y.-R.; Hong, O.-Y.; Jung, S.H.; Youn, H.J.; Kim, J.-S. Aurora kinases are essential for PKC-induced invasion and matrix metalloproteinase-9 expression in MCF-7 breast cancer cells. *Oncol. Rep.* **2015**, *34*, 803–810. [[CrossRef](#)] [[PubMed](#)]
516. Landen, C.N.; Lin, Y.G.; Immaneni, A.; Deavers, M.T.; Merritt, W.M.; Spanuth, W.A.; Bodurka, D.C.; Gershenson, D.M.; Brinkley, W.R.; Sood, A.K. Overexpression of the Centrosomal Protein Aurora-A Kinase is Associated with Poor Prognosis in Epithelial Ovarian Cancer Patients. *Clin. Cancer Res.* **2007**, *13*, 4098–4104. [[CrossRef](#)] [[PubMed](#)]
517. Tuncel, H.; Shimamoto, F.; Qi, H.K.; Aoki, E.; Jikihara, H.; Nakai, S.; Takata, T.; Tatsuka, M. Nuclear Aurora B and cytoplasmic Survivin expression is involved in lymph node metastasis of colorectal cancer. *Oncol. Lett.* **2012**, *3*, 1109–1114. [[CrossRef](#)]
518. Rannou, Y.; Troadec, M.-B.; Petretti, C.; Hans, F.; Dutertre, S.; Dimitrov, S.; Prigent, C. Localization of aurora A and aurora B kinases during interphase: Role of the N-terminal domain. *Cell Cycle* **2008**, *7*, 3012–3020. [[CrossRef](#)]
519. Xia, Z.; Wei, P.; Zhang, H.; Ding, Z.; Yang, L.; Huang, Z.; Zhang, N. AURKA Governs Self-Renewal Capacity in Glioma-Initiating Cells via Stabilization/Activation of  $\beta$ -catenin/Wnt Signaling. *Mol. Cancer Res.* **2013**, *11*, 1101–1111. [[CrossRef](#)]
520. Lu, T.; Li, L.; Zhu, J.; Liu, J.; Lin, A.; Fu, W.; Liu, G.; Xia, H.; Zhang, T.; He, J. AURKA rs8173 G>C Polymorphism Decreases Wilms Tumor Risk in Chinese Children. *J. Oncol.* **2019**, *2019*, 9074908–7. [[CrossRef](#)]
521. Otto, T.; Horn, S.; Brockmann, M.; Eilers, U.; Schüttrumpf, L.; Popov, N.; Kenney, A.M.; Schulte, J.H.; Beijersbergen, R.; Christiansen, H.; et al. Stabilization of N-Myc Is a Critical Function of Aurora A in Human Neuroblastoma. *Cancer Cell* **2009**, *15*, 67–78. [[CrossRef](#)]
522. Maris, J.M. Unholy Matrimony: Aurora A and N-Myc as Malignant Partners in Neuroblastoma. *Cancer Cell* **2009**, *15*, 5–6. [[CrossRef](#)]
523. Ommer, J.; Selfe, J.L.; Wachtel, M.; O'Brien, E.M.; Laubscher, D.; Roemmele, M.; Kasper, S.; Delattre, O.; Surdez, D.; Petts, G.; et al. Aurora A Kinase Inhibition Destabilizes PAX3-FOXO1 and MYCN and Synergizes with Navitoclax to Induce Rhabdomyosarcoma Cell Death. *Cancer Res* **2020**, *80*, 832–842. [[CrossRef](#)]



524. Goldstein, M.; Meller, I.; Issakov, J.; Orr-Urtreger, A. Novel Genes Implicated in Embryonal, Alveolar, and Pleomorphic Rhabdomyosarcoma: A Cytogenetic and Molecular Analysis of Primary Tumors. *Neoplasia* **2006**, *8*, 332–343. [[CrossRef](#)]
525. Zhao, R.; Li, Z.; Huang, Y.; Xiong, C.; Zhang, C.; Liang, H.; Xu, J.; Luo, X. A Novel Ferroptosis-Related Gene Signature for Prognosis Prediction in Ewing Sarcoma. *Anal. Cell Pathol.* **2022**, *2022*, 1–22. [[CrossRef](#)]
526. Liao, H.; Xie, X.; Xu, Y.; Huang, G. Identification of driver genes associated with chemotherapy resistance of Ewing's sarcoma. *OncoTargets Ther.* **2018**, *11*, 6947–6956. [[CrossRef](#)]
527. Huang, W.-T.; Liu, A.-G.; Cai, K.-T.; He, R.-Q.; Li, Z.; Wei, Q.-J.; Chen, M.-Y.; Huang, J.-Y.; Yan, W.-Y.; Zhou, H.; et al. Exploration and validation of downregulated microRNA-199a-3p, downstream messenger RNA targets and transcriptional regulation in osteosarcoma. *Am. J. Transl. Res.* **2019**, *11*, 7538–7554.
528. Zhu, X.; Mei, J.; Wang, Z. Aurora-A kinase: Potential tumor marker of osteosarcoma. *J. Cancer Res. Ther.* **2014**, *10*, 102–107. [[CrossRef](#)]
529. Yang, W.; Jiang, X.; Zhao, X.; Mao, P. Treatment of RB -deficient retinoblastoma with Aurora-A kinase inhibitor. *Kaohsiung J. Med. Sci.* **2021**, *38*, 244–252. [[CrossRef](#)]
530. Lehman, N.L.; O'Donnell, J.P.; Whiteley, L.J.; Stapp, R.T.; Lehman, T.D.; Roszka, K.M.; Schultz, L.R.; Williams, C.J.; Mikkelsen, T.; Brown, S.L.; et al. Aurora A is differentially expressed in gliomas, is associated with patient survival in glioblastoma and is a potential chemotherapeutic target in gliomas. *Cell Cycle* **2012**, *11*, 489–502. [[CrossRef](#)]
531. Liang, B.; Zhou, Y.; Jiao, J.; Xu, L.; Yan, Y.; Wu, Q.; Tong, X.; Yan, H. Integrated Analysis of Transcriptome Data Revealed AURKA and KIF20A as Critical Genes in Medulloblastoma Progression. *Front. Oncol.* **2022**, *12*. [[CrossRef](#)]
532. Vader, G.; Medema, R.; Lens, S.M. The chromosomal passenger complex: Guiding Aurora-B through mitosis. *J. Cell Biol.* **2006**, *173*, 833–837. [[CrossRef](#)]
533. Vagnarelli, P.; Earnshaw, W.C. Chromosomal passengers: The four-dimensional regulation of mitotic events. *Chromosoma* **2004**, *113*, 211–222. [[CrossRef](#)]
534. Adams, R.R.; Carmena, M.; Earnshaw, W.C. Chromosomal passengers and the (aurora) ABCs of mitosis. *Trends Cell Biol.* **2001**, *11*, 49–54. [[CrossRef](#)] [[PubMed](#)]
535. Minoshima, Y.; Kawashima, T.; Hirose, K.; Tonozuka, Y.; Kawajiri, A.; Bao, Y.C.; Deng, X.; Tatsuka, M.; Narumiya, S.; May, W.; et al. Phosphorylation by Aurora B Converts MgcRacGAP to a RhoGAP during Cytokinesis. *Dev. Cell* **2003**, *4*, 549–560. [[CrossRef](#)] [[PubMed](#)]
536. Hsu, J.-Y.; Sun, Z.-W.; Li, X.; Reuben, M.; Tatchell, K.; Bishop, D.K.; Grushcow, J.M.; Brame, C.J.; A Caldwell, J.; Hunt, D.F.; et al. Mitotic Phosphorylation of Histone H3 Is Governed by Ipl1/aurora Kinase and Glc7/PP1 Phosphatase in Budding Yeast and Nematodes. *Cell* **2000**, *102*, 279–291. [[CrossRef](#)] [[PubMed](#)]
537. Murnion, M.E.; Adams, R.R.; Callister, D.M.; Allis, C.D.; Earnshaw, W.C.; Swedlow, J.R. Chromatin-associated Protein Phosphatase 1 Regulates Aurora-B and Histone H3 Phosphorylation. *J. Biol. Chem.* **2001**, *276*, 26656–26665. [[CrossRef](#)] [[PubMed](#)]
538. Lan, W.; Zhang, X.; Kline-Smith, S.L.; E Rosasco, S.; A Barrett-Wilt, G.; Shabanowitz, J.; Hunt, D.F.; E Walczak, C.; Stukenberg, T. Aurora B Phosphorylates Centromeric MCAK and Regulates Its Localization and Microtubule Depolymerization Activity. *Curr. Biol.* **2004**, *14*, 273–286. [[CrossRef](#)]
539. Ma, H.T.; Poon, R.Y. Aurora kinases and DNA damage response. *Mutat. Res. Mol. Mech. Mutagen.* **2020**, *821*, 111716. [[CrossRef](#)]
540. González-Loyola, A.; Fernández-Miranda, G.; Trakala, M.; Partida, D.; Samejima, K.; Ogawa, H.; Cañamero, M.; de Martino, A.; Martínez-Ramírez, A.; de Cáncer, G.; et al. Aurora B Overexpression Causes Aneuploidy and p21<sup>Cip1</sup> Repression during Tumor Development. *Mol. Cell. Biol.* **2015**, *35*, 3566–3578. [[CrossRef](#)]
541. Takeshita, M.; Koga, T.; Takayama, K.; Ijichi, K.; Yano, T.; Maehara, Y.; Nakanishi, Y.; Sueishi, K. Aurora-B overexpression is correlated with aneuploidy and poor prognosis in non-small cell lung cancer. *Lung Cancer* **2013**, *80*, 85–90. [[CrossRef](#)]
542. Porcelli, L.; Guida, G.; E Quatrala, A.; Cocco, T.; Sidella, L.; Maida, I.; Iacobazzi, R.M.; Ferretta, A.; A Stolfa, D.; Strippoli, S.; et al. Aurora kinase B inhibition reduces the proliferation of metastatic melanoma cells and enhances the response to chemotherapy. *J. Transl. Med.* **2015**, *13*, 26. [[CrossRef](#)]
543. Wang, C.; Chen, J.; Cao, W.; Sun, L.; Sun, H.; Liu, Y. Aurora-B and HDAC synergistically regulate survival and proliferation of lymphoma cell via AKT, mTOR and Notch pathways. *Eur. J. Pharmacol.* **2016**, *779*, 1–7. [[CrossRef](#)]
544. Wan, B.; Huang, Y.; Liu, B.; Lu, L.; Lv, C. AURKB: A promising biomarker in clear cell renal cell carcinoma. *PeerJ* **2019**, *7*, e7718. [[CrossRef](#)]
545. Twu, N.-F.; Yuan, C.-C.; Yen, M.-S.; Lai, C.-R.; Chao, K.-C.; Wang, P.-H.; Wu, H.-H.; Chen, Y.-J. Expression of Aurora kinase A and B in normal and malignant cervical tissue: High Aurora A kinase expression in squamous cervical cancer. *Eur. J. Obstet. Gynecol. Reprod. Biol.* **2009**, *142*, 57–63. [[CrossRef](#)]
546. Pannone, G.; Hindi, S.; Santoro, A.; Sanguedolce, F.; Rubini, C.; Cincione, R.; De Maria, S.; Tortorella, S.; Rocchetti, R.; Cagiano, S.; et al. Aurora B Expression as a Prognostic Indicator and Possible Therapeutic Target in Oral Squamous Cell Carcinoma. *Int. J. Immunopathol. Pharmacol.* **2011**, *24*, 79–88. [[CrossRef](#)]
547. Alafate, W.; Zuo, J.; Deng, Z.; Guo, X.; Wu, W.; Zhang, W.; Xie, W.; Wang, M.; Wang, J. Combined elevation of AURKB and UBE2C predicts severe outcomes and therapy resistance in glioma. *Pathol.-Res. Pract.* **2019**, *215*, 152557. [[CrossRef](#)] [[PubMed](#)]
548. Liu, M.; Li, Y.; Zhang, C.; Zhang, Q. Role of aurora kinase B in regulating resistance to paclitaxel in breast cancer cells. *Hum. Cell* **2022**, *35*, 678–693. [[CrossRef](#)]

549. Wang, Z.; Yu, Z.; Wang, G.-H.; Zhou, Y.-M.; Deng, J.-P.; Feng, Y.; Chen, J.-Q.; Tian, L. AURKB Promotes the Metastasis of Gastric Cancer, Possibly by Inducing EMT. *Cancer Manag. Res.* **2020**, *12*, 6947–6958. [[CrossRef](#)]
550. Nie, M.; Wang, Y.; Yu, Z.; Li, X.; Deng, Y.; Wang, Y.; Yang, D.; Li, Q.; Zeng, X.; Ju, J.; et al. AURKB promotes gastric cancer progression via activation of CCND1 expression. *Aging* **2020**, *12*, 1304–1321. [[CrossRef](#)]
551. Yuan, K.; Wu, M.; Lyu, S.; Li, Y. Identification of prognostic genes for early basal-like breast cancer with weighted gene co-expression network analysis. *Medicine* **2022**, *101*, e30581. [[CrossRef](#)]
552. Gao, X.; Jiang, A.; Shen, Y.; Lu, H.M.; Chen, R. Expression and clinical significance of AURKB gene in lung adenocarcinoma. *Medicine* **2021**, *100*, e26439. [[CrossRef](#)]
553. Yang, Y.; Sheng, Y.; Sun, D.; Sun, J.; Li, L.; Sun, L. AURKB promotes tumorigenesis and carboplatin resistance by regulating the ERK pathway in neuroblastoma cells. *Int. J. Neurosci.* **2021**, 1–11. [[CrossRef](#)]
554. Bogen, D.; Wei, J.S.; Azorsa, D.O.; Ormanoglu, P.; Buehler, E.; Guha, R.; Keller, J.M.; Griner, L.A.M.; Ferrer, M.; Song, Y.K.; et al. Aurora B kinase is a potent and selective target in MYCN-driven neuroblastoma. *Oncotarget* **2015**, *6*, 35247–35262. [[CrossRef](#)] [[PubMed](#)]
555. Hartsink-Segers, S.; Zwaan, C.; Exalto, C.; Luijendijk, M.W.J.; Calvert, V.; Petricoin, E.F.; Evans, W.; Reinhardt, D.; De Haas, V.; Hedtj rn, M.; et al. Aurora kinases in childhood acute leukemia: The promise of aurora B as therapeutic target. *Leukemia* **2012**, *27*, 560–568. [[CrossRef](#)] [[PubMed](#)]
556. Saletta, F.; Wadham, C.; Ziegler, D.; Marshall, G.M.; Haber, M.; McCowage, G.; Norris, M.D.; Byrne, J.A. Molecular profiling of childhood cancer: Biomarkers and novel therapies. *BBA Clin.* **2014**, *1*, 59–77. [[CrossRef](#)]
557. Gibson, S.E.; Zeng, W.F.; Weil, R.J.; Prayson, R.A. Aurora B Kinase Expression in Ependymal Neoplasms. *Appl. Immunohistochem. Mol. Morphol.* **2008**, *16*, 274–278. [[CrossRef](#)]
558. Wang, S.; Hwang, E.E.; Guha, R.; O'Neill, A.F.; Melong, N.; Veinotte, C.J.; Saur, A.C.; Wuerthele, K.; Shen, M.; McKnight, C.; et al. High-throughput Chemical Screening Identifies Focal Adhesion Kinase and Aurora Kinase B Inhibition as a Synergistic Treatment Combination in Ewing Sarcoma. *Clin. Cancer Res.* **2019**, *25*, 4552–4566. [[CrossRef](#)] [[PubMed](#)]
559. Zhao, Z.; Jin, G.; Yao, K.; Liu, K.; Liu, F.; Chen, H.; Wang, K.; Gorja, D.R.; Reddy, K.; Bode, A.M.; et al. Aurora B kinase as a novel molecular target for inhibition the growth of osteosarcoma. *Mol. Carcinog.* **2019**, *58*, 1056–1067. [[CrossRef](#)]
560. Borah, N.A.; Sradhanjali, S.; Barik, M.R.; Jha, A.; Tripathy, D.; Kaliki, S.; Rath, S.; Raghav, S.K.; Patnaik, S.; Mittal, R.; et al. Aurora Kinase B Expression, Its Regulation and Therapeutic Targeting in Human Retinoblastoma. *Investig. Ophthalmology Vis. Sci.* **2021**, *62*, 16. [[CrossRef](#)]
561. Kimura, M.; Matsuda, Y.; Yoshioka, T.; Okano, Y. Cell Cycle-dependent Expression and Centrosome Localization of a Third Human Aurora/Ipl1-related Protein Kinase, AIK3. *J. Biol. Chem.* **1999**, *274*, 7334–7340. [[CrossRef](#)]
562. Fujii, S.; Srivastava, V.; Hegde, A.; Kondo, Y.; Shen, L.; Hoshino, K.; Gonzalez, Y.; Wang, J.; Sasai, K.; Ma, X.; et al. Regulation of AURKC expression by CpG island methylation in human cancer cells. *Tumor Biol.* **2015**, *36*, 8147–8158. [[CrossRef](#)]
563. Tseng, T.-C.; Chen, S.-H.; Hsu, Y.-P.P.; Tang, T.K. Protein Kinase Profile of Sperm and Eggs: Cloning and Characterization of Two Novel Testis-Specific Protein Kinases (AIE1, AIE2) Related to Yeast and Fly Chromosome Segregation Regulators. *DNA Cell Biol.* **1998**, *17*, 823–833. [[CrossRef](#)]
564. Santos, M.A.; van de Werken, C.; de Vries, M.; Jahr, H.; Vromans, M.J.; Laven, J.S.; Fauser, B.C.; Kops, G.J.; Lens, S.M.; Baart, E.B. A role for Aurora C in the chromosomal passenger complex during human preimplantation embryo development. *Hum. Reprod.* **2011**, *26*, 1868–1881. [[CrossRef](#)]
565. Khan, J.; Ezan, F.; Cr met, J.-Y.; Fautrel, A.; Gilot, D.; Lambert, M.; Benaud, C.; Troadec, M.-B.; Prigent, C. Overexpression of Active Aurora-C Kinase Results in Cell Transformation and Tumour Formation. *PLoS ONE* **2011**, *6*, e26512. [[CrossRef](#)]
566. Yan, X.; Cao, L.; Li, Q.; Wu, Y.; Zhang, H.; Saiyin, H.; Liu, X.; Zhang, X.; Shi, Q.; Yu, L. Aurora C is directly associated with Survivin and required for cytokinesis. *Genes Cells* **2005**, *10*, 617–626. [[CrossRef](#)]
567. Kobayashi, K.; Kiyomura, H. The theoretical analysis on the tooth movement (II). *Nihon Kyosei Shika Gakkai zasshi = J. Jpn. Orthod. Soc.* **1982**, *41*, 716–722.
568. Tsou, J.-H.; Chang, K.-C.; Chang-Liao, P.-Y.; Yang, S.-T.; Lee, C.-T.; Chen, Y.-P.; Lee, Y.-C.; Lin, B.-W.; Lee, J.-C.; Shen, M.-R.; et al. Aberrantly expressed AURKC enhances the transformation and tumourigenicity of epithelial cells. *J. Pathol.* **2011**, *225*, 243–254. [[CrossRef](#)]
569. Zekri, A.; Lesan, V.; Ghaffari, S.H.; Tabrizi, M.H.; Modarressi, M.H. Gene Amplification and Overexpression of Aurora-C in Breast and Prostate Cancer Cell Lines. *Oncol. Res. Featur. Preclin. Clin. Cancer Ther.* **2012**, *20*, 241–250. [[CrossRef](#)]
570. Seu nez, H.N.; Pereira, H.S.; Lima, S.C.S.; de Faria, P.S.; Cardoso, L.C.D.A. i RPS6KA4 i i MIR1237 i and i AURKC i promoter regions are differentially methylated in Wilms tumor. *Front. Biosci.* **2018**, *10*, 143–154. [[CrossRef](#)]
571. Hsieh, C.-H.; Cheung, C.H.Y.; Liu, Y.-L.; Hou, C.-L.; Hsu, C.-L.; Huang, C.-T.; Yang, T.-S.; Chen, S.-F.; Chen, C.-N.; Hsu, W.-M.; et al. Quantitative Proteomics of Th-MYCN Transgenic Mice Reveals Aurora Kinase Inhibitor Altered Metabolic Pathways and Enhanced ACADM To Suppress Neuroblastoma Progression. *J. Proteome Res.* **2019**, *18*, 3850–3866. [[CrossRef](#)]
572. Bejar, J.F.; DiSanza, Z.; Quartuccio, S.M. The oncogenic role of meiosis-specific Aurora kinase C in mitotic cells. *Exp. Cell Res.* **2021**, *407*, 112803. [[CrossRef](#)]
573. Nowell, P.C.; Hungerford, D.A. Chromosome studies on normal and leukemic human leukocytes. *J. Natl. Cancer Inst.* **1960**, *25*, 85–109.
574. Seabright, M. A rapid banding technique for human chromosomes. *Lancet* **1971**, *298*, 971–972. [[CrossRef](#)] [[PubMed](#)]

575. Rowley, J.D. A New Consistent Chromosomal Abnormality in Chronic Myelogenous Leukaemia identified by Quinacrine Fluorescence and Giemsa Staining. *Nature* **1973**, *243*, 290–293. [[CrossRef](#)] [[PubMed](#)]
576. Heisterkamp, N.; Stephenson, J.R.; Groffen, J.; Hansen, P.F.; de Klein, A.; Bartram, C.R.; Grosveld, G. Localization of the c-abl oncogene adjacent to a translocation break point in chronic myelocytic leukaemia. *Nature* **1983**, *306*, 239–242. [[CrossRef](#)] [[PubMed](#)]
577. de Klein, A.; van Kessel, A.G.; Grosveld, G.; Bartram, C.R.; Hagemeijer, A.; Bootsma, D.; Spurr, N.K.; Heisterkamp, N.; Groffen, J.; Stephenson, J.R. A cellular oncogene is translocated to the Philadelphia chromosome in chronic myelocytic leukaemia. *Nature* **1982**, *300*, 765–767. [[CrossRef](#)] [[PubMed](#)]
578. Groffen, J.; Stephenson, J.R.; Heisterkamp, N.; De Klein, A.; Bartram, C.R.; Grosveld, G. Philadelphia chromosomal breakpoints are clustered within a limited region, bcr, on chromosome 22. *Cell* **1984**, *36*, 93–99. [[CrossRef](#)]
579. Koschmieder, S.; Göttgens, B.; Zhang, P. Inducible chronic phase of myeloid leukemia with expansion of hematopoietic stem cells in a trans-genic model of BCR-ABL leukemogenesis. *Blood* **2005**, *105*, 324–334. [[CrossRef](#)]
580. Druker, B.J.; Talpaz, M.; Resta, D.J.; Peng, B.; Buchdunger, E.; Ford, J.M.; Lydon, N.B.; Kantarjian, H.; Capdeville, R.; Ohno-Jones, S.; et al. Efficacy and Safety of a Specific Inhibitor of the BCR-ABL Tyrosine Kinase in Chronic Myeloid Leukemia. *N. Engl. J. Med.* **2001**, *344*, 1031–1037. [[CrossRef](#)]
581. Festuccia, C.; Gravina, G.L.; Biordi, L.; D’Ascenzo, S.; Dolo, V.; Ficorella, C.; Ricevuto, E.; Tombolini, V. Effects of EGFR tyrosine kinase inhibitor erlotinib in prostate cancer cells in vitro. *Prostate* **2009**, *69*, 1529–1537. [[CrossRef](#)]
582. Abdelgalil, A.A.; Al-Kahtani, H.M.; Al-Jenoobi, F.I. Erlotinib. *Profiles DrugSubst. Excip. Relat. Methodol.* **2019**, *45*, 93–117. [[CrossRef](#)]
583. Ms, K.N.R.; Zweidler-McKay, P.A.; Van Roy, N.; Speleman, F.; Trevino, J.; Zage, P.E.; Hughes, D.P.M. Signaling of ERBB receptor tyrosine kinases promotes neuroblastoma growth in vitro and in vivo. *Cancer* **2010**, *116*, 3233–3243. [[CrossRef](#)]
584. Ji, X.L.; He, M. Sodium cantharidate targets STAT3 and abrogates EGFR inhibitor resistance in osteosarcoma. *Aging* **2019**, *11*, 5848–5863. [[CrossRef](#)]
585. Bandyopadhyay, A.; Favours, E.; Phelps, D.A.; Del Pozo, V.; Ghilu, S.; Kurmashev, D.; Michalek, J.; Trevino, A.; Guttridge, D.; London, C.; et al. Evaluation of patritumab with or without erlotinib in combination with standard cytotoxic agents against pediatric sarcoma xenograft models. *Pediatr. Blood Cancer* **2017**, *65*, e26870. [[CrossRef](#)]
586. Abraham, J.; Nelon, L.D.; Kubicek, C.B.; Kilcoyne, A.; Hampton, S.T.; Zarzabal, L.A.; Giles, F.J.; Michalek, J.E.; Rubin, B.P.; Keller, C. Preclinical Testing of Erlotinib in a Transgenic Alveolar Rhabdomyosarcoma Mouse Model. *Sarcoma* **2011**, *2011*, 1–5. [[CrossRef](#)]
587. Hernan, R.; Fasheh, R.; Calabrese, C.; Frank, A.J.; MacLean, K.H.; Allard, D.; Barraclough, R.; Gilbertson, R.J. ERBB2 up-regulates S100A4 and several other prometastatic genes in medulloblastoma. *Cancer Res* **2003**, *63*, 140–148.
588. Guan, S.; Shen, R.; Lafortune, T.; Tiao, N.; Houghton, P.; Yung, W.A.; Koul, D. Establishment and characterization of clinically relevant models of ependymoma: A true challenge for targeted therapy. *Neuro-Oncology* **2011**, *13*, 748–758. [[CrossRef](#)]
589. Shao, Y.; Yu, Y.; Zong, R.; Quyang, L.; He, H.; Zhou, Q.; Pei, C. Erlotinib has tumor inhibitory effect in human retinoblastoma cells. *Biomed. Pharmacother.* **2017**, *85*, 479–485. [[CrossRef](#)]
590. Jakacki, R.I.; Hamilton, M.; Gilbertson, R.J.; Blaney, S.M.; Tersak, J.; Krailo, M.D.; Ingle, A.M.; Voss, S.D.; Dancey, J.E.; Adamson, P.C. Pediatric Phase I and Pharmacokinetic Study of Erlotinib Followed by the Combination of Erlotinib and Temozolomide: A Children’s Oncology Group Phase I Consortium Study. *J. Clin. Oncol.* **2008**, *26*, 4921–4927. [[CrossRef](#)]
591. Georger, B.; Hargrave, D.; Thomas, F.; Ndiaye, A.; Frappaz, D.; Andreiuolo, F.; Varlet, P.; Aerts, I.; Riccardi, R.; Jaspan, T.; et al. Innovative Therapies for Children with Cancer pediatric phase I study of erlotinib in brainstem glioma and relapsing/refractory brain tumors. *Neuro-Oncology* **2010**, *13*, 109–118. [[CrossRef](#)]
592. Frampton, J.E. Vandetanib. *Drugs* **2012**, *72*, 1423–1436. [[CrossRef](#)]
593. Karras, S.; Anagnostis, P.; E Krassas, G. Vandetanib for the treatment of thyroid cancer: An update. *Expert Opin. Drug Metab. Toxicol.* **2014**, *10*, 469–481. [[CrossRef](#)]
594. Hatem, R.; Labiod, D.; Château-Joubert, S.; de Plater, L.; El Botty, R.; Vacher, S.; Bonin, F.; Servely, J.-L.; Dieras, V.; Bièche, I.; et al. Vandetanib as a potential new treatment for estrogen receptor-negative breast cancers. *Int. J. Cancer* **2016**, *138*, 2510–2521. [[CrossRef](#)] [[PubMed](#)]
595. Valerio, L.; Bottici, V.; Matrone, A.; Piaggi, P.; Viola, D.; Cappagli, V.; Agate, L.; Molinaro, E.; Ciampi, R.; Tacito, A.; et al. Medullary thyroid cancer treated with vandetanib: Predictors of a longer and durable response. *Endocrine-Related Cancer* **2020**, *27*, 97–110. [[CrossRef](#)] [[PubMed](#)]
596. Ding, X.; Xiang, L.; Wang, N.; Zhao, Z.; Jin, X.; Sun, Y.; Duan, W.; Wang, S.; Jin, X. Vandetanib-induced inhibition of neuroblastoma cell migration and invasion is associated with downregulation of the SDF-1/CXCR4 axis and matrix metalloproteinase 14. *Oncol. Rep.* **2013**, *31*, 1165–1174. [[CrossRef](#)] [[PubMed](#)]
597. Li, H.; Li, C.-W.; Li, X.; Ding, Q.; Guo, L.; Liu, S.; Liu, C.; Lai, C.-C.; Hsu, J.-M.; Dong, Q.; et al. MET Inhibitors Promote Liver Tumor Evasion of the Immune Response by Stabilizing PDL1. *Gastroenterology* **2019**, *156*, 1849–1861.e7. [[CrossRef](#)]
598. Beaudry, P.; Nilsson, M.; Rioth, M.; Prox, D.; Poon, D.; Xu, L.; Zweidler-Mckay, P.; Ryan, A.; Folkman, J.; Ryeom, S.; et al. Potent antitumor effects of ZD6474 on neuroblastoma via dual targeting of tumor cells and tumor endothelium. *Mol. Cancer Ther.* **2008**, *7*, 418–424. [[CrossRef](#)]
599. Cazes, A.; Lopez-Delisle, L.; Tsarovina, K.; Pierre-Eugène, C.; De Preter, K.; Peuchmaur, M.; Nicolas, A.; Provost, C.; Louis-Brennetot, C.; Daveau, R.; et al. Activated Alk triggers prolonged neurogenesis and Ret upregulation providing a therapeutic target in ALK-mutated neuroblastoma. *Oncotarget* **2014**, *5*, 2688–2702. [[CrossRef](#)]



600. Li, C.; Yang, C.; Wei, G. Vandetanib inhibits cisplatin-resistant neuroblastoma tumor growth and invasion. *Oncol. Rep.* **2018**, *39*, 1757–1764. [[CrossRef](#)]
601. Zage, P.E.; Zeng, L.; Palla, S.; Fang, W.; Nilsson, M.B.; Heymach, J.V.; Zweidler-McKay, P.A. A novel therapeutic combination for neuroblastoma. *Cancer* **2010**, *116*, 2465–2475. [[CrossRef](#)]
602. Craveiro, R.B.; Ehrhardt, M.; Velz, J.; Olschewski, M.; Goetz, B.; Pietsch, T.; Dilloo, D. The anti-neoplastic activity of Vandetanib against high-risk medulloblastoma variants is profoundly enhanced by additional PI3K inhibition. *Oncotarget* **2017**, *8*, 46915–46927. [[CrossRef](#)]
603. Liu, J.; Wu, J.; Zhou, L.; Pan, C.; Zhou, Y.; Du, W.; Chen, J.-M.; Zhu, X.; Shen, J.; Chen, S.; et al. ZD6474, a new treatment strategy for human osteosarcoma, and its potential synergistic effect with celecoxib. *Oncotarget* **2015**, *6*, 21341–21352. [[CrossRef](#)]
604. Andersson, M.K.; Åman, P. Proliferation of Ewing sarcoma cell lines is suppressed by the receptor tyrosine kinase inhibitors gefitinib and vandetanib. *Cancer Cell Int.* **2008**, *8*, 1–6. [[CrossRef](#)]
605. Maloney, C.; Kallis, M.P.; Edelman, M.; Tzanavaris, C.; Lesser, M.; Soffer, S.Z.; Symons, M.; Steinberg, B.M. Gefitinib Inhibits Invasion and Metastasis of Osteosarcoma via Inhibition of Macrophage Receptor Interacting Serine-Threonine Kinase 2. *Mol. Cancer Ther.* **2020**, *19*, 1340–1350. [[CrossRef](#)]
606. Wakeling, A.E.; Guy, S.P.; Woodburn, J.R.; Ashton, S.E.; Curry, B.J.; Barker, A.J.; Gibson, K.H. ZD1839 (Iressa): An orally active inhibitor of epidermal growth factor signaling with potential for cancer therapy. *Cancer Res.* **2002**, *62*, 5749–5754.
607. Ciardiello, F.; Caputo, R.; Bianco, R.; Damiano, V.; Pomatico, G. Antitumor effect and potentiation of cytotoxic drugs activity in human cancer cells by ZD-1839 (Iressa), an epidermal growth factor receptor-selective tyrosine kinase inhibitor. *Clin Cancer Res* **2000**, *6*, 2053–2063.
608. Foreman, N.K.; Gore, L.; Wells, D.; Bs, J.S.; Heideman, R.; Donson, A.M. Gefitinib is effective against juvenile pilocytic astrocytoma in vitro. *Pediatr. Blood Cancer* **2005**, *47*, 293–298. [[CrossRef](#)]
609. Schaiquevich, P.; Panetta, J.C.; Throm, S.; Daw, N.C.; Geyer, J.R.; Furman, W.L.; Stewart, C.F. Population pharmacokinetic (PK) analysis of gefitinib in pediatric cancer patients. *J. Clin. Oncol.* **2008**, *26*, 2523. [[CrossRef](#)]
610. Daudigeos-Dubus, E.; Le Dret, L.; Lanvers-Kaminsky, C.; Bawa, O.; Opolon, P.; Vievard, A.; Villa, I.; Pagès, M.; Bosq, J.; Vassal, G.; et al. Regorafenib: Antitumor Activity upon Mono and Combination Therapy in Preclinical Pediatric Malignancy Models. *PLoS ONE* **2015**, *10*, e0142612. [[CrossRef](#)]
611. Ettrich, T.J.; Seufferlein, T. Regorafenib. In *Recent Results in Cancer Research*; Springer LLC: New York, NY, USA, 2018; Volume 211, pp. 45–56. [[CrossRef](#)]
612. Subramonian, D.; Phanthilath, N.; Rinehardt, H.; Flynn, S.; Huo, Y.; Zhang, J.; Messer, K.; Mo, Q.; Huang, S.; Lesperance, J.; et al. Regorafenib is effective against neuroblastoma in vitro and in vivo and inhibits the RAS/MAPK, PI3K/Akt/mTOR and Fos/Jun pathways. *Br. J. Cancer* **2020**, *123*, 568–579. [[CrossRef](#)]
613. Carpenter, R.L.; Lo, H.-W. Dacomitinib, an emerging HER-targeted therapy for non-small cell lung cancer. *J. Thorac. Dis.* **2012**, *4*, 639–642. [[CrossRef](#)]
614. Shirley, M. Dacomitinib: First Global Approval. *Drugs* **2018**, *78*, 1947–1953. [[CrossRef](#)]
615. Popat, S.; Yap, T. Toward precision medicine with next-generation EGFR inhibitors in non-small-cell lung cancer. *Pharmacogenomics Pers. Med.* **2014**, *7*, 285–295. [[CrossRef](#)] [[PubMed](#)]
616. Endersby, R.; Whitehouse, J.; Hii, H.; Greenall, S.A.; Johns, T.G.; Gottardo, N.G. A Pre-Clinical Assessment of the Pan-ERBB Inhibitor Dacomitinib in Pediatric and Adult Brain Tumors. *Neoplasia* **2018**, *20*, 432–442. [[CrossRef](#)] [[PubMed](#)]
617. Moreira, C.; Kaklamani, V. Lapatinib and breast cancer: Current indications and outlook for the future. *Expert Rev. Anticancer Ther.* **2010**, *10*, 1171–1182. [[CrossRef](#)] [[PubMed](#)]
618. Bouchalova, K.; Cizkova, M.; Cwierka, K.; Trojanec, R.; Friedecký, D.; Hajdich, M. Lapatinib in breast cancer—the predictive significance of her1 (egfr), her2, pten and pik3ca genes and lapatinib plasma level assessment. *Biomed. Pap.* **2010**, *154*, 281–288. [[CrossRef](#)]
619. Gorlick, R.; Kolb, E.A.; Houghton, P.J.; Morton, C.L.; Phelps, D.; Schaiquevich, P.; Stewart, C.; Keir, S.T.; Lock, R.; Carol, H.; et al. Initial testing (stage 1) of lapatinib by the pediatric preclinical testing program. *Pediatr. Blood Cancer* **2009**, *53*, 594–598. [[CrossRef](#)]
620. Tebbutt, N.; Pedersen, M.W.; Johns, T.G. Targeting the ERBB family in cancer: Couples therapy. *Nat. Rev. Cancer* **2013**, *13*, 663–673. [[CrossRef](#)]
621. Herrmann, D.; Seitz, G.; Warmann, S.W.; Bonin, M.; Fuchs, J.; Armeanu-Ebinger, S. Cetuximab Promotes Immunotoxicity Against Rhabdomyosarcoma In Vitro. *J. Immunother.* **2010**, *33*, 279–286. [[CrossRef](#)]
622. Yamamoto, Y.; Fukuda, K.; Fuchimoto, Y.; Matsuzaki, Y.; Saikawa, Y.; Kitagawa, Y.; Morikawa, Y.; Kuroda, T. Cetuximab promotes anticancer drug toxicity in rhabdomyosarcomas with EGFR amplification in vitro. *Oncol. Rep.* **2013**, *30*, 1081–1086. [[CrossRef](#)]
623. O’Farrell, A.-M.; Abrams, T.J.; Yuen, H.A.; Ngai, T.J.; Louie, S.G.; Yee, K.W.H.; Wong, L.M.; Hong, W.; Lee, L.B.; Town, A.; et al. SU11248 is a novel FLT3 tyrosine kinase inhibitor with potent activity in vitro and in vivo. *Blood* **2003**, *101*, 3597–3605. [[CrossRef](#)]
624. Zhang, L.; Smith, K.M.; Chong, A.L.; Stempak, D.; Yeger, H.; Marrano, P.; Thorner, P.S.; Irwin, M.S.; Kaplan, D.R.; Baruchel, S. In Vivo Antitumor and Antimetastatic Activity of Sunitinib in Preclinical Neuroblastoma Mouse Model. *Neoplasia* **2009**, *11*, 426–435. [[CrossRef](#)]
625. Maris, J.M.; Bs, J.C.; Houghton, P.J.; Bs, C.L.M.; Kolb, E.A.; Lock, R.; Bs, M.T.; Reynolds, C.P.; Keir, S.T.; Wu, J.; et al. Initial testing (stage 1) of sunitinib by the pediatric preclinical testing program. *Pediatr. Blood Cancer* **2008**, *51*, 42–48. [[CrossRef](#)]
626. Hao, Z.; Wang, P. Lenvatinib in Management of Solid Tumors. *Oncologist* **2020**, *25*, e30. [[CrossRef](#)]



627. Matsui, J.; Yamamoto, Y.; Funahashi, Y.; Tsuruoka, A.; Watanabe, T.; Wakabayashi, T.; Uenaka, T.; Asada, M. E7080, a novel inhibitor that targets multiple kinases, has potent antitumor activities against stem cell factor producing human small cell lung cancer H146, based on angiogenesis inhibition. *Int. J. Cancer* **2007**, *122*, 664–671. [[CrossRef](#)]
628. Capozzi, M.; De Divitiis, C.; Ottaiano, A.; von Arx, C.; Scala, S.; Tatangelo, F.; Delrio, P.; Tafuto, S. Lenvatinib, a molecule with versatile application: From preclinical evidence to future development in anti-cancer treatment. *Cancer Manag. Res.* **2019**, *11*, 3847–3860. [[CrossRef](#)]
629. Bruheim, S.; Kristian, A.; Uenaka, T.; Suo, Z.; Tsuruoka, A.; Nesland, J.M.; Fodstad, A. Antitumour activity of oral E7080, a novel inhibitor of multiple tyrosine kinases, in human sarcoma xenografts. *Int. J. Cancer* **2011**, *129*, 742–750. [[CrossRef](#)]
630. Glen, H.; Mason, S.; Patel, H.; MacLeod, K.; Brunton, V.G. E7080, a multi-targeted tyrosine kinase inhibitor suppresses tumor cell migration and invasion. *BMC Cancer* **2011**, *11*, 309. [[CrossRef](#)]
631. Gaspar, N.; Campbell-Hewson, Q.; Melcon, S.G.; Locatelli, F.; Venkatramani, R.; Hecker-Nolting, S.; Gambart, M.; Bautista, F.; Thebaud, E.; Aerts, I.; et al. Phase I/II study of single-agent lenvatinib in children and adolescents with refractory or relapsed solid malignancies and young adults with osteosarcoma (ITCC-050). *ESMO Open* **2021**, *6*, 100250. [[CrossRef](#)]
632. Wilky, B.A.; Meyer, C.F.; Trent, J.C. Pazopanib in sarcomas. *Curr. Opin. Oncol.* **2013**, *25*, 373–378. [[CrossRef](#)]
633. Kumar, S.; Mokhtari, R.B.; Oliveira, I.D.; Islam, S.; Toledo, S.R.C.; Yeger, H.; Baruchel, S. Tumor Dynamics in Response to Antiangiogenic Therapy with Oral Metronomic Topotecan and Pazopanib in Neuroblastoma Xenografts. *Transl. Oncol.* **2013**, *6*, 493–503. [[CrossRef](#)]
634. Kumar, S.; Mokhtari, R.B.; Sheikh, R.; Wu, B.; Zhang, L.; Xu, P.; Man, S.; Oliveira, I.D.; Yeger, H.; Kerbel, R.S.; et al. Metronomic Oral Topotecan with Pazopanib Is an Active Antiangiogenic Regimen in Mouse Models of Aggressive Pediatric Solid Tumor. *Clin. Cancer Res.* **2011**, *17*, 5656–5667. [[CrossRef](#)]
635. Chiabotto, G.; Grignani, G.; Todorovic, M.; Martin, V.; Centomo, M.L.; Prola, E.; Giordano, G.; Merlini, A.; Miglio, U.; Berrino, E.; et al. Pazopanib and Trametinib as a Synergistic Strategy against Osteosarcoma: Preclinical Activity and Molecular Insights. *Cancers* **2020**, *12*, 1519. [[CrossRef](#)] [[PubMed](#)]
636. Keir, S.T.; Bs, C.L.M.; Wu, J.; Kurmasheva, R.T.; Houghton, P.J.; Smith, M.A. Initial testing of the multitargeted kinase inhibitor pazopanib by the pediatric preclinical testing program. *Pediatr. Blood Cancer* **2011**, *59*, 586–588. [[CrossRef](#)] [[PubMed](#)]
637. Aggerholm-Pedersen, N.; Rossen, P.; Rose, H.; Safwat, A. Pazopanib in the Treatment of Bone Sarcomas: Clinical Experience. *Transl. Oncol.* **2020**, *13*, 295–299. [[CrossRef](#)] [[PubMed](#)]
638. Mori, Y.; Kinoshita, S.; Kanamori, T.; Kataoka, H.; Joh, T.; Iida, S.; Takemoto, M.; Kondo, M.; Kuroda, J.; Komatsu, H. The Successful Treatment of Metastatic Extraosseous Ewing Sarcoma with Pazopanib. *Intern. Med.* **2018**, *57*, 2753–2757. [[CrossRef](#)] [[PubMed](#)]
639. Donson, A.M.; Amani, V.; Warner, E.A.; Griesinger, A.M.; Witt, D.A.; Levy, J.M.M.; Hoffman, L.M.; Hankinson, T.C.; Handler, M.H.; Vibhakar, R.; et al. Identification of FDA-Approved Oncology Drugs with Selective Potency in High-Risk Childhood Ependymoma. *Mol. Cancer Ther.* **2018**, *17*, 1984–1994. [[CrossRef](#)]
640. Schoen, L.F.; Craveiro, R.B.; Pietsch, T.; Moritz, T.; Troeger, A.; Jordans, S.; Dilloo, D. The PI3K inhibitor pictilisib and the multikinase inhibitors pazopanib and sorafenib have an impact on Rac1 level and migration of medulloblastoma in vitro. *J. Cell. Mol. Med.* **2022**, *26*, 5832–5845. [[CrossRef](#)]
641. Craveiro, R.B.; Ehrhardt, M.; Holst, M.I.; Pietsch, T.; Dilloo, D. In Comparative Analysis of Multi-Kinase Inhibitors for Targeted Medulloblastoma Therapy Pazopanib Exhibits Promising In Vitro and In Vivo Efficacy, 2014. Available online: [www.impactjournals.com/oncotarget](http://www.impactjournals.com/oncotarget) (accessed on 22 October 2022).
642. Scott, B.J.; Quant, E.C.; McNamara, M.B.; Ryg, P.A.; Batchelor, T.T.; Wen, P.Y. Bevacizumab salvage therapy following progression in high-grade glioma patients treated with VEGF receptor tyrosine kinase inhibitors. *Neuro-Oncology* **2010**, *12*, 603–607. [[CrossRef](#)]
643. Duke, E.S.; Barone, A.K.; Chatterjee, S.; Mishra-Kalyani, P.S.; Shen, Y.-L.; Isikwei, E.; Zhao, H.; Bi, Y.; Liu, J.; Rahman, N.A.; et al. FDA Approval Summary: Cabozantinib for Differentiated Thyroid Cancer. *Clin. Cancer Res.* **2022**, *28*, 4173–4177. [[CrossRef](#)]
644. Leavitt, J.; Copur, M.S. FDA Approved Uses of Cabozantinib. *Oncology* **2019**, *33*, 685004.
645. Yakes, F.M.; Chen, J.; Tan, J.; Yamaguchi, K.; Shi, Y.; Yu, P.; Qian, F.; Chu, F.; Bentzien, F.; Cancilla, B.; et al. Cabozantinib (XL184), a Novel MET and VEGFR2 Inhibitor, Simultaneously Suppresses Metastasis, Angiogenesis, and Tumor Growth. *Mol. Cancer Ther.* **2011**, *10*, 2298–2308. [[CrossRef](#)]
646. Bentzien, F.; Zuzow, M.; Heald, N.; Gibson, A.; Shi, Y.; Goon, L.; Yu, P.; Engst, S.; Zhang, W.; Huang, D.; et al. In Vitro and In Vivo Activity of Cabozantinib (XL184), an Inhibitor of RET, MET, and VEGFR2, in a Model of Medullary Thyroid Cancer. *Thyroid* **2013**, *23*, 1569–1577. [[CrossRef](#)]
647. Santoni, M.; Iacovelli, R.; Colonna, V.; Klinz, S.; Mauri, G.; Nuti, M. Antitumor effects of the multi-target tyrosine kinase inhibitor cabozantinib: A comprehensive review of the preclinical evidence. *Expert Rev. Anticancer. Ther.* **2021**, *21*, 1029–1054. [[CrossRef](#)]
648. Fioramonti, M.; Fausti, V.; Pantano, F.; Iuliani, M.; Ribelli, G.; Lotti, F.; Pignochino, Y.; Grignani, G.; Santini, D.; Tonini, G.; et al. Cabozantinib Affects Osteosarcoma Growth Through A Direct Effect On Tumor Cells and Modifications In Bone Microenvironment. *Sci. Rep.* **2018**, *8*, 4177. [[CrossRef](#)]
649. Pagnuzzi-Boncompagni, M.; Picco, V.; Vial, V.; Planas-Bielsa, V.; Vandenberghe, A.; Daubon, T.; Derieppe, M.-A.; Montemagno, C.; Durivault, J.; Grépin, R.; et al. Antiangiogenic Compound Axitinib Demonstrates Low Toxicity and Antitumoral Effects against Medulloblastoma. *Cancers* **2021**, *14*, 70. [[CrossRef](#)]

650. Daudigeos-Dubus, E.; Le Dret, L.; Bawa, O.; Opolon, P.; Vievard, A.; Villa, I.; Bosq, J.; Vassal, G.; Geoerger, B. Dual inhibition using cabozantinib overcomes HGF/MET signaling mediated resistance to pan-VEGFR inhibition in orthotopic and metastatic neuroblastoma tumors. *Int. J. Oncol.* **2016**, *50*, 203–211. [[CrossRef](#)]
651. Wind, S.; Schmid, U.; Freiwald, M.; Marzin, K.; Lotz, R.; Ebner, T.; Stopfer, P.; Dallinger, C. Clinical Pharmacokinetics and Pharmacodynamics of Nintedanib. *Clin. Pharmacokinet.* **2019**, *58*, 1131–1147. [[CrossRef](#)]
652. A Kharlamova, L. Study of the radiobiological reactions of Amoeba proteus. 2. The effect of the binuclear state on the radiosensitivity of amebae. *Radiobiologiya* **1972**, *12*, 934–937.
653. Lin, T.; Gong, L. Inhibition of lymphangiogenesis in vitro and in vivo by the multikinase inhibitor nintedanib. *Drug Des. Dev. Ther.* **2017**, *11*, 1147–1158. [[CrossRef](#)]
654. Patwardhan, P.P.; Musi, E.; Schwartz, G.K. Preclinical Evaluation of Nintedanib, a Triple Angiokinase Inhibitor, in Soft-tissue Sarcoma: Potential Therapeutic Implication for Synovial Sarcoma. *Mol. Cancer Ther.* **2018**, *17*, 2329–2340. [[CrossRef](#)]
655. Zhang, W.; Zhao, J.-M.; Lin, J.; Hu, C.-Z.; Zhang, W.-B.; Yang, W.-L.; Zhang, J.; Zhu, J. Adaptive Fibrogenic Reprogramming of Osteosarcoma Stem Cells Promotes Metastatic Growth. *Cell Rep.* **2018**, *24*, 1266–1277.e5. [[CrossRef](#)]
656. Milton, C.I.; Selfe, J.; Aladowicz, E.; Man, S.Y.K.; Bernauer, C.; Missiaglia, E.; Walters, Z.S.; Gatz, S.A.; Kelsey, A.; Generali, M.; et al. FGF7-FGFR2 autocrine signaling increases growth and chemoresistance of fusion-positive rhabdomyosarcomas. *Mol. Oncol.* **2021**, *16*, 1272–1289. [[CrossRef](#)]
657. Loetsch, D.; Gojo, J.; Kirchhofer, D.; Van Schoonhoven, S.; Pajtlar, K.; Kool, M.; Haberler, C.; Czech, T.; Slavc, I.; Berger, W. OS5.2 FGFR a novel target in malignant pediatric ependymoma. *Neuro-Oncology* **2018**, *20*, iii224. [[CrossRef](#)]
658. Lötsch, D.; Kirchhofer, D.; Englinger, B.; Jiang, L.; Okonechnikov, K.; Senfter, D.; Laemmerer, A.; Gabler, L.; Pirker, C.; Donson, A.M.; et al. Targeting fibroblast growth factor receptors to combat aggressive ependymoma. *Acta Neuropathol.* **2021**, *142*, 339–360. [[CrossRef](#)] [[PubMed](#)]
659. Kasamon, Y.L.; Ko, C.-W.; Subramaniam, S.; Ma, L.; Yang, Y.; Nie, L.; Shord, S.; Przepiorka, D.; Farrell, A.T.; McKee, A.E.; et al. FDA Approval Summary: Midostaurin for the Treatment of Advanced Systemic Mastocytosis. *Oncologist* **2018**, *23*, 1511–1519. [[CrossRef](#)] [[PubMed](#)]
660. Midostaurin Gets FDA Nod for AML. *Cancer Discov.* **2017**, *7*, OF5. [[CrossRef](#)]
661. Kawamoto, T.; Akisue, T.; Kishimoto, K.; Hara, H.; Imabori, M.; Fujimoto, T.; Kurosaka, M.; Hitora, T.; Kawaguchi, Y.; Yamamoto, T. Inhibition of PKC $\alpha$  activation in human bone and soft tissue sarcoma cells by the selective PKC inhibitor PKC412. *Anticancer Res.* **2008**, *28*, 825–832.
662. Boro, A.; Prêtre, K.; Rechfeld, F.; Thalhammer, V.; Oesch, S.; Wachtel, M.; Schäfer, B.W.; Niggli, F.K. Small-molecule screen identifies modulators of EWS/FLI1 target gene expression and cell survival in Ewing’s sarcoma. *Int. J. Cancer* **2012**, *131*, 2153–2164. [[CrossRef](#)]
663. Brounais, B.; Chipoy, C.; Mori, K.; Charrier, C.; Battaglia, S.; Pilet, P.; Richards, C.D.; Heymann, D.; Rédini, F.; Blanchard, F. Oncostatin M Induces Bone Loss and Sensitizes Rat Osteosarcoma to the Antitumor Effect of Midostaurin *In vivo*. *Clin. Cancer Res.* **2008**, *14*, 5400–5409. [[CrossRef](#)]
664. Kelly, R.J.; Rixe, O. Axitinib (AG-013736). *Small Mol. Oncol.* **2009**, *184*, 33–44. [[CrossRef](#)]
665. Lu, L.; Saha, D.; Martuza, R.L.; Rabkin, S.; Wakimoto, H. Single agent efficacy of the VEGFR kinase inhibitor axitinib in preclinical models of glioblastoma. *J. Neuro-Oncology* **2014**, *121*, 91–100. [[CrossRef](#)]
666. Ehrhardt, M.; Craveiro, R.B.; Velz, J.; Olschewski, M.; Casati, A.; Schönberger, S.; Pietsch, T.; Dilloo, D. The FDA approved PI 3K inhibitor GDC -0941 enhances in vitro the anti-neoplastic efficacy of Axitinib against c-myc-amplified high-risk medulloblastoma. *J. Cell. Mol. Med.* **2018**, *22*, 2153–2161. [[CrossRef](#)]
667. Suri, A.; Bailey, A.W.; Tavares, M.T.; Gunosewoyo, H.; Dyer, C.P.; Grupenmacher, A.T.; Piper, D.R.; Horton, R.A.; Tomita, T.; Kozikowski, A.P.; et al. Evaluation of Protein Kinase Inhibitors with PLK4 Cross-Over Potential in a Pre-Clinical Model of Cancer. *Int. J. Mol. Sci.* **2019**, *20*, 2112. [[CrossRef](#)]
668. Schwinn, S.; Mokhtari, Z.; Thusek, S.; Schneider, T.; Sirén, A.-L.; Tiemeyer, N.; Caruana, I.; Miele, E.; Schlegel, P.G.; Beilhack, A.; et al. Cytotoxic effects and tolerability of gemcitabine and axitinib in a xenograft model for c-myc amplified medulloblastoma. *Sci. Rep.* **2021**, *11*, 1–15. [[CrossRef](#)]
669. Saha, D.; Wakimoto, H.; Peters, C.W.; Antoszczyk, S.J.; Rabkin, S.D.; Martuza, R.L. Combinatorial Effects of VEGFR Kinase Inhibitor Axitinib and Oncolytic Virotherapy in Mouse and Human Glioblastoma Stem-Like Cell Models. *Clin. Cancer Res.* **2018**, *24*, 3409–3422. [[CrossRef](#)]
670. Rössler, J.; Monnet, Y.; Farace, F.; Opolon, P.; Daudigeos-Dubus, E.; Bourredjem, A.; Vassal, G.; Geoerger, B. The selective VEGFR1-3 inhibitor axitinib (AG-013736) shows antitumor activity in human neuroblastoma xenografts. *Int. J. Cancer* **2010**, *128*, 2748–2758. [[CrossRef](#)]
671. Lu, D.; Jimenez, X.; Zhang, H.; Bohlen, P.; Witte, L.; Zhu, Z. Selection of high affinity human neutralizing antibodies to VEGFR2 from a large antibody phage display library for antiangiogenesis therapy. *Int. J. Cancer* **2001**, *97*, 393–399. [[CrossRef](#)]
672. Lowery, C.D.; Blosser, W.; Dowless, M.; Renschler, M.; Perez, L.V.; Stephens, J.; Pytowski, B.; Wasserstrom, H.; Stancato, L.F.; Falcon, B. Anti-VEGFR2 therapy delays growth of preclinical pediatric tumor models and enhances anti-tumor activity of chemotherapy. *Oncotarget* **2019**, *10*, 5523–5533. [[CrossRef](#)]
673. Casak, S.J.; Fashoyin-Aje, I.; Lemery, S.J.; Zhang, L.; Jin, R.; Li, H.; Zhao, L.; Zhao, H.; Zhang, H.; Chen, H.; et al. FDA Approval Summary: Ramucirumab for Gastric Cancer. *Clin. Cancer Res.* **2015**, *21*, 3372–3376. [[CrossRef](#)]

674. Syed, Y.Y. Ramucirumab: A Review in Hepatocellular Carcinoma. *Drugs* **2020**, *80*, 315–322. [[CrossRef](#)]
675. Tiwari, P. Ramucirumab: Boon or bane. *J. Egypt. Natl. Cancer Inst.* **2016**, *28*, 133–140. [[CrossRef](#)]
676. Debeuckelaere, C.; Murgioni, S.; Lonardi, S.; Girardi, N.; Alberti, G.; Fano, C.; Gallimberti, S.; Magro, C.; Ahcene-Djaballah, S.; Daniel, F.; et al. Ramucirumab: The long and winding road toward being an option for mCRC treatment. *Expert Opin. Biol. Ther.* **2019**, *19*, 399–409. [[CrossRef](#)] [[PubMed](#)]
677. Larkins, E.; Blumenthal, G.M.; Chen, H.; He, K.; Agarwal, R.; Gieser, G.; Stephens, O.; Zahalka, E.; Ringgold, K.; Helms, W.; et al. FDA Approval: Alectinib for the Treatment of Metastatic, ALK-Positive Non-Small Cell Lung Cancer Following Crizotinib. *Clin. Cancer Res.* **2016**, *22*, 5171–5176. [[CrossRef](#)] [[PubMed](#)]
678. Kodama, T.; Hasegawa, M.; Takanashi, K.; Sakurai, Y.; Kondoh, O.; Sakamoto, H. Antitumor activity of the selective ALK inhibitor alectinib in models of intracranial metastases. *Cancer Chemother. Pharmacol.* **2014**, *74*, 1023–1028. [[CrossRef](#)] [[PubMed](#)]
679. Ryu, S.; Hayashi, M.; Aikawa, H.; Okamoto, I.; Fujiwara, Y.; Hamada, A. Heterogeneous distribution of alectinib in neuroblastoma xenografts revealed by matrix-assisted laser desorption ionization mass spectrometry imaging: A pilot study. *Br. J. Pharmacol.* **2017**, *175*, 29–37. [[CrossRef](#)]
680. Chen, L.; Humphreys, A.; Turnbull, L.; Bellini, A.; Schleiermacher, G.; Salwen, H.; Cohn, S.L.; Bown, N.; Tweddle, D.A. Identification of different ALK mutations in a pair of neuroblastoma cell lines established at diagnosis and relapse. *Oncotarget* **2016**, *7*, 87301–87311. [[CrossRef](#)]
681. Lu, J.; Guan, S.; Zhao, Y.; Yu, Y.; Woodfield, S.E.; Zhang, H.; Yang, K.L.; Bieerkehazhi, S.; Qi, L.; Li, X.; et al. The second-generation ALK inhibitor alectinib effectively induces apoptosis in human neuroblastoma cells and inhibits tumor growth in a TH-MYCN transgenic neuroblastoma mouse model. *Cancer Lett.* **2017**, *400*, 61–68. [[CrossRef](#)]
682. Yang, K.; Chen, Y.; Wang, F. Correction: Alectinib (CH5424802) antagonizes ABCB1- and ABCG2-mediated multidrug resistance in vitro, in vivo and ex vivo. *Exp. Mol. Med.* **2020**, *52*, 989–990. [[CrossRef](#)]
683. Brunac, A.; Laprie, A.; Castex, M.; Laurent, C.; Le Loarer, F.; Karanian, M.; Le Guellec, S.; Guillemot, D.; Pierron, G.; Gomez-Bouchet, A. The combination of radiotherapy and ALK inhibitors is effective in the treatment of intraosseous rhabdomyosarcoma with *FUS-TFCP2* fusion transcript. *Pediatr. Blood Cancer* **2020**, *67*, e28185. [[CrossRef](#)]
684. Hagiwara, K.; Tokunaga, T.; Iida, H.; Nagai, H. Combined Inhibition of ALK and HDAC Induces Synergistic Cytotoxicity in Neuroblastoma Cell Lines. *Anticancer. Res.* **2019**, *39*, 3579–3584. [[CrossRef](#)]
685. Berezowska, S.; Diermeier-Daucher, S.; Brockhoff, G.; Busch, R.; Duyster, J.; Grosu, A.-L.; Schlegel, J. Effect of additional inhibition of human epidermal growth factor receptor 2 with the bispecific tyrosine kinase inhibitor AEE788 on the resistance to specific EGFR inhibition in glioma cells. *Int. J. Mol. Med.* **2010**, *26*, 713–721. [[CrossRef](#)]
686. Servidei, T.; Meco, D.; Trivieri, N.; Patriarca, V.; Vellone, V.G.; Zannoni, G.F.; Lamorte, G.; Pallini, R.; Riccardi, R. Effects of epidermal growth factor receptor blockade on ependymoma stem cells in vitro and in orthotopic mouse models. *Int. J. Cancer* **2012**, *131*, E791–E803. [[CrossRef](#)]
687. Park, Y.W.; Younes, M.N.; Jasser, S.A.; Yigitbasi, O.G.; Zhou, G.; Bucana, C.D.; Bekele, B.N.; Myers, J.N. AEE788, a Dual Tyrosine Kinase Receptor Inhibitor, Induces Endothelial Cell Apoptosis in Human Cutaneous Squamous Cell Carcinoma Xenografts in Nude Mice. *Clin. Cancer Res.* **2005**, *11*, 1963–1973. [[CrossRef](#)]
688. Meco, D.; Servidei, T.; Zannoni, G.F.; Martinelli, E.; Prisco, M.G.; de Waure, C.; Riccardi, R. Dual Inhibitor AEE78 Reduces Tumor Growth in Preclinical Models of Medulloblastoma. *Transl. Oncol.* **2010**, *3*, 326–335. [[CrossRef](#)]
689. Heigener, D.F.; Reck, M. Crizotinib. In *Recent Results in Cancer Research*; Springer LLC: New York, NY, USA, 2018; Volume 211, pp. 57–65. [[CrossRef](#)]
690. Zomeran, W.W.; Plasschaert, S.L.A.; Diks, S.H.; Lourens, H.-J.; Boer, T.M.-D.; Hoving, E.W.; Dunnen, W.F.A.D.; De Bont, E.S.J.M. Exogenous HGF Bypasses the Effects of ErbB Inhibition on Tumor Cell Viability in Medulloblastoma Cell Lines. *PLoS ONE* **2015**, *10*, e0141381. [[CrossRef](#)]
691. Sie, M.; Dunnen, W.F.A.D.; Lourens, H.J.; Boer, T.G.J.M.-D.; Scherpen, F.J.G.; Zomeran, W.W.; Kampen, K.; Hoving, E.W.; De Bont, E.S.J.M. Growth-Factor-Driven Rescue to Receptor Tyrosine Kinase (RTK) Inhibitors through Akt and Erk Phosphorylation in Pediatric Low Grade Astrocytoma and Ependymoma. *PLoS ONE* **2015**, *10*, e0122555. [[CrossRef](#)]
692. Megiorni, F.; McDowell, H.P.; Camero, S.; Mannarino, O.; Ceccarelli, S.; Paiano, M.; Losty, P.D.; Pizer, B.; Shukla, R.; Pizzuti, A.; et al. Crizotinib-induced antitumour activity in human alveolar rhabdomyosarcoma cells is not solely dependent on ALK and MET inhibition. *J. Exp. Clin. Cancer Res.* **2015**, *34*, 1–16. [[CrossRef](#)]
693. Schöffski, P.; Wozniak, A.; Leahy, M.G.; Aamdal, S.; Rutkowski, P.; Bauer, S.; Richter, S.; Grünwald, V.; Debiec-Rychter, M.; Sciort, R.; et al. The tyrosine kinase inhibitor crizotinib does not have clinically meaningful activity in heavily pre-treated patients with advanced alveolar rhabdomyosarcoma with FOXO rearrangement: European Organisation for Research and Treatment of Cancer phase 2 trial 90101 'CREATE'. *Eur. J. Cancer* **2018**, *94*, 156–167. [[CrossRef](#)]
694. Heuckmann, J.M.; Hölzel, M.; Sos, M.L.; Heynck, S.; Balke-Want, H.; Koker, M.; Peifer, M.; Weiss, J.; Lovly, C.M.; Grütter, C.; et al. ALK Mutations Conferring Differential Resistance to Structurally Diverse ALK Inhibitors. *Clin. Cancer Res.* **2011**, *17*, 7394–7401. [[CrossRef](#)]
695. George, R.E.; Sanda, T.; Hanna, M.; Fröhling, S.; Luther, W., II; Zhang, J.; Ahn, Y.; Zhou, W.; London, W.B.; McGrady, P.; et al. Activating mutations in ALK provide a therapeutic target in neuroblastoma. *Nature* **2008**, *455*, 975–978. [[CrossRef](#)]



696. Bresler, S.C.; Weiser, D.A.; Huwe, P.J.; Park, J.H.; Krytska, K.; Ryles, H.; Laudenslager, M.; Rappaport, E.F.; Wood, A.C.; McGrady, P.W.; et al. ALK Mutations Confer Differential Oncogenic Activation and Sensitivity to ALK Inhibition Therapy in Neuroblastoma. *Cancer Cell* **2014**, *26*, 682–694. [[CrossRef](#)]
697. Dagogo-Jack, I.; Shaw, A.T. Crizotinib resistance: Implications for therapeutic strategies. *Ann. Oncol.* **2016**, *27* (Suppl. S3), iii42–iii50. [[CrossRef](#)] [[PubMed](#)]
698. Zuckermann, M.; He, C.; Andrews, J.; Sloan-Henry, R.; Bianski, B.; Xie, J.; Wang, Y.; Twarog, N.; Onar-Thomas, A.; Ernst, K.; et al. MODL-06. Targeting c-MET in combination with radiation is effective in MET-fusion driven high-grade glioma. *Neuro-Oncology* **2022**, *24*, i169. [[CrossRef](#)]
699. Esaki, T.; Hirai, F.; Makiyama, A.; Seto, T.; Bando, H.; Naito, Y.; Yoh, K.; Ishihara, K.; Kakizume, T.; Natsume, K.; et al. Phase I dose-escalation study of capmatinib (INC 280) in Japanese patients with advanced solid tumors. *Cancer Sci.* **2019**, *110*, 1340–1351. [[CrossRef](#)] [[PubMed](#)]
700. Markham, A. Tepotinib: First Approval. *Drugs* **2020**, *80*, 829–833. [[CrossRef](#)]
701. Zou, H.Y.; Li, Q.; Lee, J.H.; Arango, M.E.; Burgess, K.; Qiu, M.; Engstrom, L.D.; Yamazaki, S.; Parker, M.; Timofeevski, S.; et al. Sensitivity of Selected Human Tumor Models to PF-04217903, a Novel Selective c-Met Kinase Inhibitor. *Mol. Cancer Ther.* **2012**, *11*, 1036–1047. [[CrossRef](#)]
702. Niswander, L.M.; Guenther, L.M.; Mendoza, A.; Khanna, C.; Christensen, J.G.; Helman, L.J.; Kim, S. Effect of modulation of MET with the small molecule inhibitor PF-04217903 on osteosarcoma metastasis in vivo. *J. Clin. Oncol.* **2010**, *28*, 9567. [[CrossRef](#)]
703. Lock, R.; Ingraham, R.; Maertens, O.; Miller, A.L.; Weledji, N.; Legius, E.; Konicek, B.M.; Yan, S.-C.B.; Graff, J.R.; Cichowski, K. Cotargeting MNK and MEK kinases induces the regression of NF1-mutant cancers. *J. Clin. Investig.* **2016**, *126*, 2181–2190. [[CrossRef](#)]
704. Katayama, R.; Aoyama, A.; Yamori, T.; Qi, J.; Oh-Hara, T.; Song, Y.; Engelman, J.A.; Fujita, N. Cytotoxic Activity of Tivantinib (ARQ 197) Is Not Due Solely to c-MET Inhibition. *Cancer Res* **2013**, *73*, 3087–3096. [[CrossRef](#)]
705. Geller, J.I.; Perentesis, J.P.; Liu, X.; Minard, C.G.; Kudgus, R.A.; Reid, J.M.; Fox, E.; Blaney, S.M.; Weigel, B.J. A phase 1 study of the c-Met inhibitor, tivantinib (ARQ197) in children with relapsed or refractory solid tumors: A Children’s Oncology Group study phase 1 and pilot consortium trial (ADVL1111). *Pediatr. Blood Cancer* **2017**, *64*, e26565. [[CrossRef](#)]
706. Goldberg, J.M.; Gavcovich, T.; Saigal, G.; Goldman, J.W.; Rosen, L.S. Extended Progression-Free Survival in Two Patients With Alveolar Soft Part Sarcoma Exposed to Tivantinib. *J. Clin. Oncol.* **2014**, *32*, e114–e116. [[CrossRef](#)]
707. Johnson, T.W.; Richardson, P.F.; Bailey, S.; Brooun, A.; Burke, B.J.; Collins, M.R.; Cui, J.J.; Deal, J.G.; Deng, Y.-L.; Dinh, D.; et al. Discovery of (10R)-7-Amino-12-fluoro-2,10,16-trimethyl-15-oxo-10,15,16,17-tetrahydro-2H-8,4-(metheno)pyrazolo[4,3-h][[2,5,11]-benzoxadiazacyclotetradecine-3-carbonitrile (PF-06463922), a Macrocyclic Inhibitor of Anaplastic Lymphoma Kinase (ALK) and c-ros Oncogene 1 (ROS1) with Preclinical Brain Exposure and Broad-Spectrum Potency against ALK-Resistant Mutations. *J. Med. Chem.* **2014**, *57*, 4720–4744. [[CrossRef](#)]
708. Infarinato, N.R.; Park, J.H.; Krytska, K.; Ryles, H.T.; Sano, R.; Szigety, K.M.; Li, Y.; Zou, H.Y.; Lee, N.V.; Smeal, T.; et al. The ALK/ROS1 Inhibitor PF-06463922 Overcomes Primary Resistance to Crizotinib in ALK-Driven Neuroblastoma. *Cancer Discov.* **2016**, *6*, 96–107. [[CrossRef](#)]
709. Guan, J.; Tucker, E.R.; Wan, H.; Chand, D.; Danielson, L.S.; Ruuth, K.; El Wakil, A.; Witek, B.; Jamin, Y.; Umaphathy, G.; et al. The ALK inhibitor PF-06463922 is effective as a single agent in neuroblastoma driven by expression of ALK and MYCN. *Dis. Model. Mech.* **2016**, *9*, 941–952. [[CrossRef](#)]
710. Collier, T.L.; Maresca, K.P.; Normandin, M.D.; Richardson, P.; McCarthy, T.J.; Liang, S.H.; Waterhouse, R.N.; Vasdev, N. Brain Penetration of the ROS1/ALK Inhibitor Lorlatinib Confirmed by PET. *Mol. Imaging* **2017**, *16*. [[CrossRef](#)] [[PubMed](#)]
711. Bagchi, A.; Orr, B.A.; Campagne, O.; Dhanda, S.; Nair, S.; Tran, Q.; Christensen, A.M.; Gajjar, A.; Furtado, L.V.; Vasilyeva, A.; et al. Lorlatinib in a Child with ALK-Fusion-Positive High-Grade Glioma. *New Engl. J. Med.* **2021**, *385*, 761–763. [[CrossRef](#)] [[PubMed](#)]
712. Liu, T.; Merguerian, M.D.; Rowe, S.P.; Pratilas, C.A.; Chen, A.R.; Ladle, B.H. Exceptional response to the ALK and ROS1 inhibitor lorlatinib and subsequent mechanism of resistance in relapsed ALK F1174L-mutated neuroblastoma. *Mol. Case Stud.* **2021**, *7*, a006064. [[CrossRef](#)] [[PubMed](#)]
713. Khozin, S.; Blumenthal, G.M.; Zhang, L.; Tang, S.; Brower, M.; Fox, E.; Helms, W.; Leong, R.; Song, P.; Pan, Y.; et al. FDA Approval: Ceritinib for the Treatment of Metastatic Anaplastic Lymphoma Kinase-Positive Non-Small Cell Lung Cancer. *Clin. Cancer Res.* **2015**, *21*, 2436–2439. [[CrossRef](#)] [[PubMed](#)]
714. Guan, J.; Fransson, S.; Siaw, J.T. Clinical response of the novel activating ALK-I1171T mutation in neuroblastoma to the ALK inhibitor ceritinib Running title: Clinical response to ceritinib in ALK-positive neuroblastoma. *Cold Spring Harb. Mol. Case Stud.* **2018**, *4*, a002550. [[CrossRef](#)]
715. Mittal, A.; Gupta, A.; Rastogi, S.; Barwad, A.; Sharma, S. Near-complete response to low-dose ceritinib in recurrent infantile inflammatory myofibroblastic tumour. *Ecancermedicalscience* **2021**, *15*. [[CrossRef](#)]
716. Russo, A.; Paret, C.; Lehmann, N.; Bender, H.; Wingerter, A.; Neu, M.A.; Alt, F.; Backes, N.; Roth, L.; Seidmann, L.; et al. Epen-29. individualized therapy of an anaplastic ependymoma pediatric patient with a notch1 germline mutation. *Neuro-Oncology* **2018**, *20*, i79. [[CrossRef](#)]
717. Russo, A.; Paret, C.; Alt, F.; Burhenne, J.; Fresnais, M.; Wagner, W.; Glaser, M.; Bender, H.; Huprich, S.; Harter, P.N.; et al. Ceritinib-Induced Regression of an Insulin-Like Growth Factor-Driven Neuroepithelial Brain Tumor. *Int. J. Mol. Sci.* **2019**, *20*, 4267. [[CrossRef](#)] [[PubMed](#)]



718. Lin, C.Y.; Erkek, S.; Tong, Y.; Yin, L.; Federation, A.J.; Zapatka, M.; Haldipur, P.; Kawauchi, D.; Risch, T.; Warnatz, H.-J.; et al. Active medulloblastoma enhancers reveal subgroup-specific cellular origins. *Nature* **2016**, *530*, 57–62. [[CrossRef](#)] [[PubMed](#)]
719. Tsoi, M.; Wadham, C.; Pinese, M.; Failes, T.; Joshi, S.; Mould, E.; Yin, J.X.; Gayevskiy, V.; Kumar, A.; Kaplan, W.; et al. Integration of genomics, high throughput drug screening, and personalized xenograft models as a novel precision medicine paradigm for high risk pediatric cancer. *Cancer Biol. Ther.* **2018**, *19*, 1078–1087. [[CrossRef](#)] [[PubMed](#)]
720. Beck, O.; Paret, C.; Russo, A.; Burhenne, J.; Fresnais, M.; Steimel, K.; Seidmann, L.; Wagner, D.-C.; Vewinger, N.; Lehmann, N.; et al. Safety and Activity of the Combination of Ceritinib and Dasatinib in Osteosarcoma. *Cancers* **2020**, *12*, 793. [[CrossRef](#)] [[PubMed](#)]
721. van Erp, A.E.M.; Hillebrandt-Roeffen, M.H.S.; van Houdt, L.; Fleuren, E.D.G.; van der Graaf, W.T.A.; Versleijen-Jonkers, Y.M.H. Targeting Anaplastic Lymphoma Kinase (ALK) in Rhabdomyosarcoma (RMS) with the Second-Generation ALK Inhibitor Ceritinib. *Target. Oncol.* **2017**, *12*, 815–826. [[CrossRef](#)] [[PubMed](#)]
722. Dolgikh, N.; Fulda, S. Rhabdomyosarcoma cells are susceptible to cell death by LDK378 alone or in combination with sorafenib independently of anaplastic lymphoma kinase status. *Anti-Cancer Drugs* **2017**, *28*, 1118–1125. [[CrossRef](#)]
723. Huang, W.-S.; Liu, S.; Zou, D.; Thomas, M.; Wang, Y.; Zhou, T.; Romero, J.; Kohlmann, A.; Li, F.; Qi, J.; et al. Discovery of Brigatinib (AP26113), a Phosphine Oxide-Containing, Potent, Orally Active Inhibitor of Anaplastic Lymphoma Kinase. *J. Med. Chem.* **2016**, *59*, 4948–4964. [[CrossRef](#)]
724. Mo, F.; Pellerino, A.; Soffietti, R.; Rudà, R. Blood–Brain Barrier in Brain Tumors: Biology and Clinical Relevance. *Int. J. Mol. Sci.* **2021**, *22*, 12654. [[CrossRef](#)]
725. Spencer, S.A.; Riley, A.C.; Matthew, A.; Di Pasqua, A.J. Brigatinib: Novel ALK Inhibitor for Non–Small-Cell Lung Cancer. *Ann. Pharmacother.* **2019**, *53*, 621–626. [[CrossRef](#)]
726. Siaw, J.T.; Wan, H.; Pfeifer, K.; Rivera, V.M.; Guan, J.; Palmer, R.H.; Hallberg, B. Brigatinib, an anaplastic lymphoma kinase inhibitor, abrogates activity and growth in ALK-positive neuroblastoma cells, *Drosophila* and mice. *Oncotarget* **2016**, *7*, 29011–29022. [[CrossRef](#)]
727. Drilon, A.; De Braud, F.G.; Siena, S.; Ou, S.-H.I.; Patel, M.; Ahn, M.-J.; Lee, J.; Bauer, T.M.; Farago, A.F.; Liu, S.V.; et al. Abstract CT007: Entrectinib, an oral pan-Trk, ROS1, and ALK inhibitor in TKI-naïve patients with advanced solid tumors harboring gene rearrangements: Updated phase I results. *Cancer Res* **2016**, *76*, CT007. [[CrossRef](#)]
728. Jiang, Q.; Li, M.; Li, H.; Chen, L. Entrectinib, a new multi-target inhibitor for cancer therapy. *Biomed. Pharmacother.* **2022**, *150*, 112974. [[CrossRef](#)]
729. Iyer, R.; Wehrmann, L.; Golden, R.L.; Naraparaju, K.; Croucher, J.L.; MacFarland, S.P.; Guan, P.; Kolla, V.; Wei, G.; Cam, N.; et al. Entrectinib is a potent inhibitor of Trk-driven neuroblastomas in a xenograft mouse model. *Cancer Lett.* **2016**, *372*, 179–186. [[CrossRef](#)]
730. Desai, A.V.; Robinson, G.W.; Gauvain, K.; Basu, E.M.; E Macy, M.; Maese, L.; Whipple, N.S.; Sabnis, A.J.; Foster, J.H.; Shusterman, S.; et al. Entrectinib in children and young adults with solid or primary CNS tumors harboring *NTRK*, *ROS1*, or *ALK* aberrations (STARTRK-NG). *Neuro-Oncology* **2022**, *24*, 1776–1789. [[CrossRef](#)]
731. MacFarland, S.P.; Naraparaju, K.; Iyer, R.; Guan, P.; Kolla, V.; Hu, Y.; Tan, K.; Brodeur, G.M. Mechanisms of Entrectinib Resistance in a Neuroblastoma Xenograft Model. *Mol. Cancer Ther.* **2020**, *19*, 920–926. [[CrossRef](#)]
732. Spitaleri, G.; Passaro, A.; de Marinis, F. Ensartinib (X-396) a novel drug for anaplastic lymphoma kinase-positive non-small cell lung cancer patients: We need smart trials to avoid wasting good bullets. *Chin. Clin. Oncol.* **2019**, *8*, S1. [[CrossRef](#)]
733. Lovly, C.M.; Heuckmann, J.M.; de Stanchina, E.; Chen, H.; Thomas, R.K.; Liang, C.; Pao, W. Insights into ALK-Driven Cancers Revealed through Development of Novel ALK Tyrosine Kinase Inhibitors. *Cancer Res* **2011**, *71*, 4920–4931. [[CrossRef](#)]
734. Di Paolo, D.; Yang, D.; Pastorino, F.; Emionite, L.; Cilli, M.; Daga, A.; Destafanis, E.; Di Fiore, A.; Piaggio, F.; Brignole, C.; et al. New therapeutic strategies in neuroblastoma: Combined targeting of a novel tyrosine kinase inhibitor and liposomal siRNAs against *ALK*. *Oncotarget* **2015**, *6*, 28774–28789. [[CrossRef](#)]
735. Perera, T.P.; Jovcheva, E.; Mevellec, L.; Vialard, J.; De Lange, D.; Verhulst, T.; Paulussen, C.; Van De Ven, K.; King, P.; Freyne, E.; et al. Discovery and Pharmacological Characterization of JNJ-42756493 (Erdafitinib), a Functionally Selective Small-Molecule FGFR Family Inhibitor. *Mol. Cancer Ther.* **2017**, *16*, 1010–1020. [[CrossRef](#)]
736. Luzzi, S.; Brambilla, I.; Mantelli, S.S.; Mosconi, M.; Foadelli, T.; Savasta, S. Targeting the medulloblastoma: A molecular-based approach. *Acta Biomed.* **2020**, *91*, 79–100. [[CrossRef](#)]
737. Musumeci, F.; Ciancusi, A.; D’Agostino, I.; Grossi, G.; Carbone, A.; Schenone, S. Synthetic Heterocyclic Derivatives as Kinase Inhibitors Tested for the Treatment of Neuroblastoma. *Molecules* **2021**, *26*, 7069. [[CrossRef](#)] [[PubMed](#)]
738. Holzhauser, S.; Lukoseviciute, M.; Papachristofi, C.; Vasilopoulou, C.; Herold, N.; Wickström, M.; Kostopoulou, O.N.; Dalianis, T. Effects of PI3K and FGFR inhibitors alone and in combination, and with/without cytostatics in childhood neuroblastoma cell lines. *Int. J. Oncol.* **2021**, *58*, 211–225. [[CrossRef](#)] [[PubMed](#)]
739. Lukoseviciute, M.; Maier, H.; Poulou-Sidiropoulou, E.; Rosendahl, E.; Holzhauser, S.; Dalianis, T.; Kostopoulou, O.N. Targeting PI3K, FGFR, CDK4/6 Signaling Pathways Together With Cytostatics and Radiotherapy in Two Medulloblastoma Cell Lines. *Front. Oncol.* **2021**, *11*, 748657. [[CrossRef](#)] [[PubMed](#)]
740. Angevin, E.; Lopez-Martin, J.A.; Lin, C.-C.; Gschwend, J.E.; Harzstark, A.; Castellano, D.; Soria, J.-C.; Sen, P.; Chang, J.; Shi, M.; et al. Phase I Study of Dovitinib (TKI258), an Oral FGFR, VEGFR, and PDGFR Inhibitor, in Advanced or Metastatic Renal Cell Carcinoma. *Clin. Cancer Res.* **2013**, *19*, 1257–1268. [[CrossRef](#)]

741. Arnz, E.; Hertwig, F.; Krämer, A.; Ikram, F.; Roels, F.; Kocak, H.; Engesser, A.; Thomas, R.; Peifer, M.; Ackermann, S.; et al. Fibroblast growth factor receptors as therapeutic targets in neuroblastoma. *Klin. Pädiatrie* **2014**, *226*, A22. [[CrossRef](#)]
742. Li, S.Q.; Cheuk, A.T.; Shern, J.F.; Song, Y.K.; Hurd, L.; Liao, H.; Wei, J.S.; Khan, J. Targeting Wild-Type and Mutationally Activated FGFR4 in Rhabdomyosarcoma with the Inhibitor Ponatinib (AP24534). *PLoS ONE* **2013**, *8*, e76551. [[CrossRef](#)]
743. Schramm, K.; Iskar, M.; Statz, B.; Jäger, N.; Haag, D.; Slabicki, M.; Pfister, S.M.; Zapatka, M.; Gronych, J.; Jones, D.T.W.; et al. DECIPHER pooled shRNA library screen identifies PP2A and FGFR signaling as potential therapeutic targets for diffuse intrinsic pontine gliomas. *Neuro-Oncology* **2019**, *21*, 867–877. [[CrossRef](#)]
744. Yan, D.; Kowal, J.; Akkari, L.; Schuhmacher, A.J.; Huse, J.T.; West, B.L.; Joyce, J. Inhibition of colony stimulating factor-1 receptor abrogates microenvironment-mediated therapeutic resistance in gliomas. *Oncogene* **2017**, *36*, 6049–6058. [[CrossRef](#)]
745. Georjin-Lavialle, S.; Lhermitte, L.; Suarez, F.; Yang, Y.; Létard, S.; Hanssens, K.; Feger, F.; Renand, A.; Brouze, C.; Canioni, D.; et al. Mast cell leukemia: Identification of a new c-Kit mutation, dup(501-502), and response to masitinib, a c-Kit tyrosine kinase inhibitor. *Eur. J. Haematol.* **2012**, *89*, 47–52. [[CrossRef](#)]
746. Dubreuil, P.; Letard, S.; Ciufolini, M.; Gros, L.; Humbert, M.; Castéran, N.; Borge, L.; Hajem, B.; Lermet, A.; Sippl, W.; et al. Masitinib (AB1010), a Potent and Selective Tyrosine Kinase Inhibitor Targeting KIT. *PLoS ONE* **2009**, *4*, e7258. [[CrossRef](#)]
747. Marech, I.; Patrino, R.; Zizzo, N.; Gadaleta, C.; Introna, M.; Zito, A.F.; Gadaleta, C.D.; Ranieri, G. Masitinib (AB1010), from canine tumor model to human clinical development: Where we are? *Crit. Rev. Oncol.* **2014**, *91*, 98–111. [[CrossRef](#)]
748. Fleming, T.; Cunningham, C.; Keir, S. *The Effect of Masitinib on Pediatric Glioblastoma*; Duke University: Durham, NC, USA, 2014.
749. Buti, S.; Leonetti, A.; Dallatomasina, A.; Bersanelli, M. Everolimus in the management of metastatic renal cell carcinoma: An evidence-based review of its place in therapy. *Core Evid.* **2016**, *11*, 23–36. [[CrossRef](#)]
750. Nashan, B. Review of the proliferation inhibitor everolimus. *Expert Opin. Investig. Drugs* **2002**, *11*, 1845–1857. [[CrossRef](#)]
751. Pignochino, Y.; Dell’Aglia, C.; Basiricò, M.; Capozzi, F.; Soster, M.; Marchiò, S.; Bruno, S.; Gammaitoni, L.; Sangiolo, D.; Torchiario, E.; et al. The Combination of Sorafenib and Everolimus Abrogates mTORC1 and mTORC2 Upregulation in Osteosarcoma Preclinical Models. *Clin. Cancer Res.* **2013**, *19*, 2117–2131. [[CrossRef](#)]
752. Miklja, Z.; Yadav, V.N.; Cartaxo, R.T.; Siada, R.; Thomas, C.C.; Cummings, J.R.; Mullan, B.; Stallard, S.; Paul, A.; Bruzek, A.K.; et al. Everolimus improves the efficacy of dasatinib in PDGFR $\alpha$ -driven glioma. *J. Clin. Investig.* **2020**, *130*, 5313–5325. [[CrossRef](#)]
753. Poore, B.; Yuan, M.; Arnold, A.; Price, A.; Alt, J.; A Rubens, J.; Slusher, B.S.; Eberhart, C.G.; Raabe, E.H. Inhibition of mTORC1 in pediatric low-grade glioma depletes glutathione and therapeutically synergizes with carboplatin. *Neuro-Oncology* **2018**, *21*, 252–263. [[CrossRef](#)]
754. Salussolia, C.L.; Klonowska, K.; Kwiatkowski, D.J.; Sahin, M. Genetic Etiologies, Diagnosis, and Treatment of Tuberous Sclerosis Complex. *Annu. Rev. Genom. Hum. Genet.* **2019**, *20*, 217–240. [[CrossRef](#)]
755. Rosset, C.; Netto, C.B.O.; Ashton-Prolla, P. TSC1 and TSC2 gene mutations and their implications for treatment in Tuberous Sclerosis Complex: A review. *Genet. Mol. Biol.* **2017**, *40*, 69–79. [[CrossRef](#)]
756. Krueger, D.A.; Care, M.M.; Holland, K.; Agricola, K.; Tudor, C.; Mangeshkar, P.; Wilson, K.A.; Byars, A.; Sahnoud, T.; Franz, D.N. Everolimus for Subependymal Giant-Cell Astrocytomas in Tuberous Sclerosis. *N. Engl. J. Med.* **2010**, *363*, 1801–1811. [[CrossRef](#)]
757. Franz, D.N.; Lawson, J.A.; Yapici, Z.; Ikeda, H.; Polster, T.; Nabbout, R.; Curatolo, P.; de Vries, P.J.; Dlugos, D.J.; Voi, M.; et al. Everolimus for treatment-refractory seizures in TSC. *Neurol. Clin. Pract.* **2018**, *8*, 412–420. [[CrossRef](#)]
758. Xue, Q.; Hopkins, B.; Perruzzi, C.; Udayakumar, D.; Sherris, D.; Benjamin, L.E. Palomid 529, a Novel Small-Molecule Drug, Is a TORC1/TORC2 Inhibitor That Reduces Tumor Growth, Tumor Angiogenesis, and Vascular Permeability. *Cancer Res* **2008**, *68*, 9551–9557. [[CrossRef](#)] [[PubMed](#)]
759. Lin, F.; Buil, L.; Sherris, D.; Beijnen, J.H.; van Tellingen, O. Dual mTORC1 and mTORC2 inhibitor Palomid 529 penetrates the Blood-Brain Barrier without restriction by ABCB1 and ABCG2. *Int. J. Cancer* **2013**, *133*, 1222–1233. [[CrossRef](#)] [[PubMed](#)]
760. Gravina, G.L.; Mancini, A.; Colapietro, A.; Monache, S.D.; Sferra, R.; Pompili, S.; Vitale, F.; Martellucci, S.; Marampon, F.; Mattei, V.; et al. The Brain Penetrating and Dual TORC1/TORC2 Inhibitor, RES529, Elicits Anti-Glioma Activity and Enhances the Therapeutic Effects of Anti-Angiogenic Compounds in Preclinical Murine Models. *Cancers* **2019**, *11*, 1604. [[CrossRef](#)] [[PubMed](#)]
761. Cerna, D.; Carter, D.; Flaherty, S.; Cao, L.; Sherris, D.; Yoo, S.S. Abstract 2506: Palomid 529, a PI3K/Akt/mTOR dual TORC1/2 inhibitor, is a radiosensitizer with effect in both subcutaneous and orthotopic U251 glioblastoma tumor xenograft models. *Cancer Res.* **2010**, *70*, 2506. [[CrossRef](#)]
762. Hu, X.; Wang, Z.; Chen, M.; Chen, X.; Liang, W. The anti-osteosarcoma cell activity by a mTORC1/2 dual inhibitor RES-529. *Biochem. Biophys. Res. Commun.* **2018**, *497*, 499–505. [[CrossRef](#)]
763. Bhagwat, S.V.; Gokhale, P.C.; Crew, A.P.; Cooke, A.; Yao, Y.; Mantis, C.; Kahler, J.; Workman, J.; Bittner, M.; Dudkin, L.; et al. Preclinical Characterization of OSI-027, a Potent and Selective Inhibitor of mTORC1 and mTORC2: Distinct from Rapamycin. *Mol. Cancer Ther.* **2011**, *10*, 1394–1406. [[CrossRef](#)]
764. Eckerdt, F.; Clymer, J.; Bell, J.B.; Beauchamp, E.M.; Blyth, G.T.; Goldman, S.; Plataniias, L.C. Pharmacological mTOR targeting enhances the antineoplastic effects of selective PI3K $\alpha$  inhibition in medulloblastoma. *Sci. Rep.* **2019**, *9*, 12822. [[CrossRef](#)]
765. Srivastava, R.K.; Guroji, P.; Jin, L.; Mukhtar, M.S.; Athar, M. Combined inhibition of BET bromodomain and mTORC1/2 provides therapeutic advantage for rhabdomyosarcoma by switching cell death mechanism. *Mol. Carcinog.* **2022**, *61*, 737–751. [[CrossRef](#)]
766. Eckerdt, F.; Beauchamp, E.; Bell, J.; Iqbal, A.; Su, B.; Fukunaga, R.; Lulla, R.R.; Goldman, S.; Plataniias, L.C. Regulatory effects of a Mnk2-eIF4E feedback loop during mTORC1 targeting of human medulloblastoma cells. *Oncotarget* **2014**, *5*, 8442–8451. [[CrossRef](#)]

767. Clymer, J.; Eckerd, F.; Bell, J.; Lulla, R.; Goldman, S.; Platanius, L. MEDU-44. TARGETING SHH SIGNALING VIA PI3K/MTOR INHIBITION IN MEDULLOBLASTOMA AND EWING SARCOMA. *Neuro-Oncology* **2017**, *19*, iv47. [[CrossRef](#)]
768. Calimeri, T.; Ferreri, A.J.M. m-TOR inhibitors and their potential role in haematological malignancies. *Br. J. Haematol.* **2017**, *177*, 684–702. [[CrossRef](#)]
769. Kolev, V.N.; Wright, Q.G.; Vidal, C.M.; Ring, J.E.; Shapiro, I.M.; Ricono, J.; Weaver, D.T.; Padval, M.V.; Pachter, J.A.; Xu, Q. PI3K/mTOR Dual Inhibitor VS-5584 Preferentially Targets Cancer Stem Cells. *Cancer Res* **2015**, *75*, 446–455. [[CrossRef](#)]
770. Sun, J.-Y.; Hou, Y.-J.; Cui, H.-J.; Zhang, C.; Yang, M.-F.; Wang, F.-Z.; Sun, Z.; Fan, C.-D.; Sun, B.-L.; Oh, J.R. VS-5584 Inhibits Human Osteosarcoma Cells Growth by Induction of G1- phase Arrest through Regulating PI3K/mTOR and MAPK Pathways. *Curr. Cancer Drug Targets* **2020**, *20*, 616–623. [[CrossRef](#)]
771. Sun, J.-Y.; Hou, Y.-J.; Yin, Y.-B.; Wang, F.-Z.; Yang, M.-F.; Zhang, Y.-Y.; Fan, C.-D.; Sun, B.-L. CCT128930 induces G1-phase arrest and apoptosis and synergistically enhances the anticancer efficiency of VS5584 in human osteosarcoma cells. *Biomed. Pharmacother.* **2020**, *130*, 110544. [[CrossRef](#)] [[PubMed](#)]
772. Maira, S.-M.; Stauffer, F.; Brueggen, J.; Furet, P.; Schnell, C.; Fritsch, C.; Brachmann, S.; Chène, P.; De Pover, A.; Schoemaker, K.; et al. Identification and characterization of NVP-BEZ235, a new orally available dual phosphatidylinositol 3-kinase/mammalian target of rapamycin inhibitor with potent in vivo antitumor activity. *Mol. Cancer Ther.* **2008**, *7*, 1851–1863. [[CrossRef](#)]
773. Manara, M.C.; Nicoletti, G.; Zambelli, D.; Ventura, S.; Guerzoni, C.; Landuzzi, L.; Lollini, P.-L.; Maira, S.-M.; García-Echeverría, C.; Mercuri, M.; et al. NVP-BEZ235 as a New Therapeutic Option for Sarcomas. *Clin. Cancer Res.* **2010**, *16*, 530–540. [[CrossRef](#)]
774. Giorgi, C.; Boro, A.; Rechfeld, F.; Lopez-Garcia, L.A.; Gierisch, M.E.; Schäfer, B.W.; Niggli, F.K. PI3K/AKT signaling modulates transcriptional expression of EWS/FLI1 through specificity protein 1. *Oncotarget* **2015**, *6*, 28895–28910. [[CrossRef](#)]
775. Zhu, Y.-R.; Min, H.; Fang, J.-F.; Zhou, F.; Deng, X.-W.; Zhang, Y.-Q. Activity of the novel dual phosphatidylinositol 3-kinase/mammalian target of rapamycin inhibitor NVP-BEZ235 against osteosarcoma. *Cancer Biol. Ther.* **2015**, *16*, 602–609. [[CrossRef](#)]
776. Meng, W.; Wang, B.; Mao, W.; Wang, J.; Zhao, Y.; Li, Q.; Zhang, C.; Ma, J. Enhanced efficacy of histone deacetylase inhibitor panobinostat combined with dual PI3K/mTOR inhibitor BEZ235 against glioblastoma. *Nagoya J. Med. Sci.* **2019**, *81*, 93–102. [[CrossRef](#)]
777. Vazquez, N.; Lopez, A.; Cuello, V.; Persans, M.; Schuenzel, E.; Innis-Whitehouse, W.; Keniry, M. NVP-BEZ235 or JAKi Treatment leads to decreased survival of examined GBM and BBC cells. *Cancer Treat. Res. Commun.* **2021**, *27*, 100340. [[CrossRef](#)]
778. Xie, C.; Freeman, M.J.; Lu, H.; Wang, X.; Forster, C.L.; Sarver, A.L.; Hallstrom, T.C. Retinoblastoma cells activate the AKT pathway and are vulnerable to the PI3K/mTOR inhibitor NVP-BEZ235. *Oncotarget* **2017**, *8*, 38084–38098. [[CrossRef](#)] [[PubMed](#)]
779. Holzhauser, S.; Lukoseviciute, M.; Andonova, T.; Ursu, R.G.; Dalianis, T.; Wickström, M.; Kostopoulou, O.N. Targeting Fibroblast Growth Factor Receptor (FGFR) and Phosphoinositide 3-kinase (PI3K) Signaling Pathways in Medulloblastoma Cell Lines. *Anticancer. Res.* **2019**, *40*, 53–66. [[CrossRef](#)] [[PubMed](#)]
780. Gobin, B.; Battaglia, S.; Lanel, R.; Chesneau, J.; Amiaud, J.; Rédini, F.; Ory, B.; Heymann, D. NVP-BEZ235, a dual PI3K/mTOR inhibitor, inhibits osteosarcoma cell proliferation and tumor development in vivo with an improved survival rate. *Cancer Lett.* **2014**, *344*, 291–298. [[CrossRef](#)] [[PubMed](#)]
781. Chaturvedi, N.K.; Kling, M.J.; Griggs, C.N.; Kesharwani, V.; Shukla, M.; McIntyre, E.M.; Ray, S.; Liu, Y.; McGuire, T.R.; Sharp, J.G.; et al. A Novel Combination Approach Targeting an Enhanced Protein Synthesis Pathway in MYC-driven (Group 3) Medulloblastoma. *Mol. Cancer Ther.* **2020**, *19*, 1351–1362. [[CrossRef](#)] [[PubMed](#)]
782. Buonamici, S.; Williams, J.; Morrissey, M.; Wang, A.; Guo, R.; Vattay, A.; Hsiao, K.; Yuan, J.; Green, J.; Ospina, B.; et al. Interfering with Resistance to Smoothed Antagonists by Inhibition of the PI3K Pathway in Medulloblastoma. *Sci. Transl. Med.* **2010**, *2*, 51ra70. [[CrossRef](#)]
783. Garlich, J.R.; De, P.; Dey, N.; Su, J.D.; Peng, X.; Miller, A.; Murali, R.; Lu, Y.; Mills, G.B.; Kundra, V.; et al. A Vascular Targeted Pan Phosphoinositide 3-Kinase Inhibitor Prodrug, SF1126, with Antitumor and Antiangiogenic Activity. *Cancer Res* **2008**, *68*, 206–215. [[CrossRef](#)]
784. Mahadevan, D.; Chiorean, E.; Harris, W.; Von Hoff, D.; Stejskal-Barnett, A.; Qi, W.; Anthony, S.; Younger, A.; Rensvold, D.; Cordova, F.; et al. Phase I pharmacokinetic and pharmacodynamic study of the pan-PI3K/mTORC vascular targeted pro-drug SF1126 in patients with advanced solid tumours and B-cell malignancies. *Eur. J. Cancer* **2012**, *48*, 3319–3327. [[CrossRef](#)]
785. Singh, A.R.; Joshi, S.; Zulcic, M.; Alcaraz, M.; Garlich, J.R.; Morales, G.A.; Cho, Y.J.; Bao, L.; Levy, M.L.; Newbury, R.; et al. PI-3K Inhibitors Preferentially Target CD15+ Cancer Stem Cell Population in SHH Driven Medulloblastoma. *PLoS ONE* **2016**, *11*, e0150836. [[CrossRef](#)]
786. Goldin, A.N.; Singh, A.; Joshi, S.; Jamieson, C.; Durden, D.L.M. Augmented Antitumor Activity for Novel Dual PI3K/BDR4 Inhibitors, SF2523 and SF1126 in Ewing Sarcoma. *J. Pediatr. Hematol.* **2021**, *43*, e304–e311. [[CrossRef](#)]
787. Peirce, S.; Durden, D.; Garlich, J.; Findley, H. SF1126, a novel pan-PI3K inhibitor, inhibits activation of Mdm2 and in-crases sensitivity to doxorubicin in wild type p53 neuroblastoma cell lines. *Cancer Res.* **2007**, *67*, LB-294.
788. Erdreich-Epstein, A.; Singh, A.R.; Joshi, S.; Vega, F.M.; Guo, P.; Xu, J.; Groshen, S.; Ye, W.; Millard, M.; Campan, M.; et al. Association of high microvessel  $\alpha\beta 3$  and low PTEN with poor outcome in stage 3 neuroblastoma: Rationale for using first in class dual PI3K/BRD4 inhibitor, SF1126. *Oncotarget* **2016**, *8*, 52193–52210. [[CrossRef](#)]
789. Beljanski, V. Triciribine. In *xPharm: The Comprehensive Pharmacology Reference*; Elsevier: Amsterdam, The Netherlands, 2009; pp. 1–5. [[CrossRef](#)]



790. Bahmad, H.F.; Mouhieddine, T.H.; Chalhoub, R.M.; Assi, S.; Araji, T.; Chamaa, F.; Itani, M.M.; Nokkari, A.; Kobeissy, F.; Daoud, G.; et al. The Akt/mTOR pathway in cancer stem/progenitor cells is a potential therapeutic target for glioblastoma and neuroblastoma. *Oncotarget* **2018**, *9*, 33549–33561. [[CrossRef](#)]
791. Cubitt, C.L.; Menth, J.; Dawson, J.; Martinez, G.V.; Foroutan, P.; Morse, D.L.; Bui, M.M.; Letson, G.D.; Sullivan, D.M.; Reed, D.R. Rapid Screening of Novel Agents for Combination Therapy in Sarcomas. *Sarcoma* **2013**, *2013*, 1–12. [[CrossRef](#)]
792. Smeester, B.A.; Draper, G.M.; Slipek, N.J.; Larsson, A.T.; Stratton, N.; Pomeroy, E.J.; Becklin, K.L.; Yamamoto, K.; Williams, K.B.; Laoharawee, K.; et al. Implication of ZNF217 in Accelerating Tumor Development and Therapeutically Targeting ZNF217-Induced PI3K–AKT Signaling for the Treatment of Metastatic Osteosarcoma. *Mol. Cancer Ther.* **2020**, *19*, 2528–2541. [[CrossRef](#)]
793. Voss, M.H.; Gordon, M.S.; Mita, M.; Rini, B.; Makker, V.; Macarulla, T.; Smith, D.C.; Cervantes, A.; Puzanov, I.; Pili, R.; et al. Phase 1 study of mTORC1/2 inhibitor sapanisertib (TAK-228) in advanced solid tumours, with an expansion phase in renal, endometrial or bladder cancer. *Br. J. Cancer* **2020**, *123*, 1590–1598. [[CrossRef](#)]
794. Slotkin, E.K.; Patwardhan, P.P.; Vasudeva, S.D.; de Stanchina, E.; Tap, W.D.; Schwartz, G.K. MLN0128, an ATP-Competitive mTOR Kinase Inhibitor with Potent In Vitro and In Vivo Antitumor Activity, as Potential Therapy for Bone and Soft-Tissue Sarcoma. *Mol. Cancer Ther.* **2015**, *14*, 395–406. [[CrossRef](#)]
795. Jiang, H.; Zeng, Z. Dual mTORC1/2 inhibition by INK-128 results in antitumor activity in preclinical models of osteosarcoma. *Biochem. Biophys. Res. Commun.* **2015**, *468*, 255–261. [[CrossRef](#)]
796. Maynard, R.E.; Poore, B.; Hanaford, A.R.; Pham, K.; James, M.; Alt, J.; Park, Y.; Slusher, B.S.; Tamayo, P.; Mesirov, J.; et al. TORC1/2 kinase inhibition depletes glutathione and synergizes with carboplatin to suppress the growth of MYC-driven medulloblastoma. *Cancer Lett.* **2021**, *504*, 137–145. [[CrossRef](#)]
797. Zhang, H.; Dou, J.; Yu, Y.; Zhao, Y.; Fan, Y.; Cheng, J.; Xu, X.; Liu, W.; Guan, S.; Chen, Z.; et al. mTOR ATP-competitive inhibitor INK128 inhibits neuroblastoma growth via blocking mTORC signaling. *Apoptosis* **2014**, *20*, 50–62. [[CrossRef](#)]
798. Miyahara, H.; Yadavilli, S.; Natsumeda, M.; Rubens, J.A.; Rodgers, L.; Kambhampati, M.; Taylor, I.C.; Kaur, H.; Asnaghi, L.; Eberhart, C.G.; et al. The dual mTOR kinase inhibitor TAK228 inhibits tumorigenicity and enhances radiosensitization in diffuse intrinsic pontine glioma. *Cancer Lett.* **2017**, *400*, 110–116. [[CrossRef](#)]
799. Arnold, A.; Yuan, M.; Price, A.; Harris, L.; Eberhart, C.G.; Raabe, E.H. Synergistic activity of mTORC1/2 kinase and MEK inhibitors suppresses pediatric low-grade glioma tumorigenicity and vascularity. *Neuro-Oncology* **2019**, *22*, 563–574. [[CrossRef](#)]
800. Tang, L.; Fu, Y.; Song, J.; Hu, T.; Li, K.; Li, Z. mTOR inhibition by TAK-228 is effective against growth, survival and angiogenesis in preclinical retinoblastoma models. *Pharmacol. Res. Perspect.* **2022**, *10*, e00930. [[CrossRef](#)] [[PubMed](#)]
801. Gray, J.E.; Infante, J.R.; Brail, L.H.; Simon, G.R.; Cooksey, J.F.; Jones, S.F.; Farrington, D.L.; Yeo, A.; Jackson, K.A.; Chow, K.H.; et al. A first-in-human phase I dose-escalation, pharmacokinetic, and pharmacodynamic evaluation of intravenous LY2090314, a glycogen synthase kinase 3 inhibitor, administered in combination with pemetrexed and carboplatin. *Investig. New Drugs* **2015**, *33*, 1187–1196. [[CrossRef](#)] [[PubMed](#)]
802. Kunnimalaiyaan, S.; Schwartz, V.K.; Jackson, I.A.; Gamblin, T.C.; Kunnimalaiyaan, M. Antiproliferative and apoptotic effect of LY2090314, a GSK-3 inhibitor, in neuroblastoma in vitro. *BMC Cancer* **2018**, *18*, 560. [[CrossRef](#)]
803. Wei, D.; Zhu, X.; Li, S.; Liu, G.; Wang, Y.; Wang, W.; Zhang, Q.; Jiang, S. Tideglusib suppresses stem-cell-like features and progression of osteosarcoma by inhibiting GSK-3 $\beta$ /NOTCH1 signaling. *Biochem. Biophys. Res. Commun.* **2021**, *554*, 206–213. [[CrossRef](#)]
804. Bahmad, H.F.; Chalhoub, R.M.; Harati, H.; Bou-Gharios, J.; Assi, S.; Ballout, F.; Monzer, A.; Msheik, H.; Araji, T.; Elajami, M.K.; et al. Tideglusib attenuates growth of neuroblastoma cancer stem/progenitor cells in vitro and in vivo by specifically targeting GSK-3 $\beta$ . *Pharmacol. Rep.* **2020**, *73*, 211–226. [[CrossRef](#)]
805. Mathuram, T.L.; Ravikumar, V.; Reece, L.M.; Karthik, S.; Sasikumar, C.S.; Cherian, K.M. Tideglusib induces apoptosis in human neuroblastoma IMR32 cells, provoking sub-G 0 /G 1 accumulation and ROS generation. *Environ. Toxicol. Pharmacol.* **2016**, *46*, 194–205. [[CrossRef](#)]
806. Bharathy, N.; Svalina, M.N.; Settelmeier, T.P.; Cleary, M.M.; Berlow, N.E.; Airhart, S.D.; Xiang, S.; Keck, J.; Hayden, J.B.; Shern, J.F.; et al. Preclinical testing of the glycogen synthase kinase-3 $\beta$  inhibitor tideglusib for rhabdomyosarcoma. *Oncotarget* **2017**, *8*, 62976–62983. [[CrossRef](#)]
807. Hirai, H.; Sootome, H.; Nakatsuru, Y.; Miyama, K.; Taguchi, S.; Tsujioka, K.; Ueno, Y.; Hatch, H.; Majumder, P.K.; Pan, B.-S.; et al. MK-2206, an Allosteric Akt Inhibitor, Enhances Antitumor Efficacy by Standard Chemotherapeutic Agents or Molecular Targeted Drugs In vitro and In vivo. *Mol. Cancer Ther.* **2010**, *9*, 1956–1967. [[CrossRef](#)]
808. Gorlick, R.; Maris, J.M.; Houghton, P.J.; Lock, R.; Carol, H.; Kurmasheva, R.T.; Kolb, E.A.; Keir, S.T.; Reynolds, C.P.; Kang, M.H.; et al. Testing of the Akt/PKB inhibitor MK-2206 by the pediatric preclinical testing program. *Pediatr. Blood Cancer* **2011**, *59*, 518–524. [[CrossRef](#)]
809. Duan, L.; E Perez, R.; Hansen, M.; Gitelis, S.; Maki, C.G. Increasing cisplatin sensitivity by schedule-dependent inhibition of AKT and Chk1. *Cancer Biol. Ther.* **2014**, *15*, 1600–1612. [[CrossRef](#)]
810. Santo, E.E.; Stroeken, P.; Sluis, P.V.; Koster, J.; Versteeg, R.; Westerhout, E.M. FOXO3a Is a Major Target of Inactivation by PI3K/AKT Signaling in Aggressive Neuroblastoma. *Cancer Res* **2013**, *73*, 2189–2198. [[CrossRef](#)]
811. Qi, L.; Toyoda, H.; Xu, D.-Q.; Zhou, Y.; Sakurai, N.; Amano, K.; Kihira, K.; Hori, H.; Azuma, E.; Komada, Y. PDK1-mTOR signaling pathway inhibitors reduce cell proliferation in MK2206 resistant neuroblastoma cells. *Cancer Cell Int.* **2015**, *15*, 1–14. [[CrossRef](#)]



812. Li, Z.; Yan, S.; Attayan, N.; Ramalingam, S.; Thiele, C.J. Combination of an Allosteric Akt Inhibitor MK-2206 with Etoposide or Rapamycin Enhances the Antitumor Growth Effect in Neuroblastoma. *Clin. Cancer Res.* **2012**, *18*, 3603–3615. [[CrossRef](#)]
813. Kang, B.W.; Chau, I. Molecular target: Pan-AKT in gastric cancer. *ESMO Open* **2020**, *5*, e000728. [[CrossRef](#)]
814. Choo, F.; Odintsov, I.; Nusser, K.; Nicholson, K.S.; Davis, L.; Corless, C.L.; Stork, L.; Somwar, R.; Ladanyi, M.; Davis, J.L.; et al. Functional impact and targetability of *PI3KCA*, *GNAS*, and *PTEN* mutations in a spindle cell rhabdomyosarcoma with MYOD1 L122R mutation. *Mol. Case Stud.* **2022**, *8*, a006140. [[CrossRef](#)]
815. Abdelgalil, A.A.; Alkahtani, H.M.; Al-Jenoobi, F.I. Sorafenib. *Profiles Drug Subst. Excip. Relat. Methodol.* **2019**, *44*, 239–266. [[CrossRef](#)]
816. Yang, F.; Van Meter, T.E.; Buettner, R.; Hedvat, M.; Liang, W.; Kowolik, C.M.; Mepani, N.; Mirosevich, J.; Nam, S.; Chen, M.Y.; et al. Sorafenib inhibits signal transducer and activator of transcription 3 signaling associated with growth arrest and apoptosis of medulloblastomas. *Mol. Cancer Ther.* **2008**, *7*, 3519–3526. [[CrossRef](#)]
817. Chai, H.; Luo, A.Z.; Weerasinghe, P.; E Brown, R. Sorafenib downregulates ERK/Akt and STAT3 survival pathways and induces apoptosis in a human neuroblastoma cell line. *Int. J. Clin. Exp. Pathol.* **2010**, *3*, 408–415.
818. Kakodkar, N.C.; Peddinti, R.R.; Tian, Y.; Guerrero, L.J.; Chlenski, A.; Yang, Q.; Salwen, H.R.; Maitland, M.L.; Cohn, S.L. Sorafenib inhibits neuroblastoma cell proliferation and signaling, blocks angiogenesis, and impairs tumor growth. *Pediatr. Blood Cancer* **2011**, *59*, 642–647. [[CrossRef](#)]
819. Karajannis, M.A.; Legault, G.; Fisher, M.J.; Milla, S.S.; Cohen, K.J.; Wisoff, J.H.; Harter, D.H.; Goldberg, J.D.; Hochman, T.; Merkelson, A.; et al. Phase II study of sorafenib in children with recurrent or progressive low-grade astrocytomas. *Neuro-Oncology* **2014**, *16*, 1408–1416. [[CrossRef](#)] [[PubMed](#)]
820. Albarrán, V.; Villamayor, M.L.; Chamorro, J.; Rosero, D.I.; Pozas, J.; Román, M.S.; Calvo, J.C.; de Aguado, P.P.; Moreno, J.; Guerrero, P.; et al. Receptor Tyrosine Kinase Inhibitors for the Treatment of Recurrent and Unresectable Bone Sarcomas. *Int. J. Mol. Sci.* **2022**, *23*, 13784. [[CrossRef](#)] [[PubMed](#)]
821. Tian, Z.; Niu, X.; Yao, W. Receptor Tyrosine Kinases in Osteosarcoma Treatment: Which Is the Key Target? *Front. Oncol.* **2020**, *10*, 1642. [[CrossRef](#)] [[PubMed](#)]
822. Wu, C.-H.; Lin, K.-H.; Fu, B.-S.; Hsu, F.-T.; Tsai, J.-J.; Weng, M.-C.; Pan, P.-J. Sorafenib Induces Apoptosis and Inhibits NF- $\kappa$ B-mediated Anti-apoptotic and Metastatic Potential in Osteosarcoma Cells. *Anticancer. Res.* **2021**, *41*, 1251–1259. [[CrossRef](#)]
823. Dumont, S.; Yang, D.; Dumont, A.; Reynoso, D.; Blay, J.-Y.; Trent, J. Targeted polytherapy in small cell sarcoma and its association with doxorubicin. *Mol. Oncol.* **2014**, *8*, 1458–1468. [[CrossRef](#)]
824. Higuchi, T.; Igarashi, K.; Yamamoto, N.; Hayashi, K.; Kimura, H.; Miwa, S.; Bouvet, M.; Tsuchiya, H.; Hoffman, R.M. Osteosarcoma Patient-derived Orthotopic Xenograft (PDOX) Models Used to Identify Novel and Effective Therapeutics: A Review. *Anticancer. Res.* **2021**, *41*, 5865–5871. [[CrossRef](#)]
825. Chen, Z.; Zhao, Y.; Yu, Y.; Pang, J.C.; Woodfield, S.E.; Tao, L.; Guan, S.; Zhang, H.; Bieerkehazhi, S.; Shi, Y.; et al. Small molecule inhibitor regorafenib inhibits RET signaling in neuroblastoma cells and effectively suppresses tumor growth in vivo. *Oncotarget* **2017**, *8*, 104090–104103. [[CrossRef](#)]
826. Harrison, D.J.; Gill, J.D.; Roth, M.E.; Zhang, W.; Teicher, B.; Erickson, S.; Gatto, G.; Kurmasheva, R.T.; Houghton, P.J.; Smith, M.A.; et al. Initial in vivo testing of a multitarget kinase inhibitor, regorafenib, by the Pediatric Preclinical Testing Consortium. *Pediatr. Blood Cancer* **2020**, *67*, e28222. [[CrossRef](#)]
827. Jindal, A.; Thadi, A.; Shailubhai, K. Hepatocellular Carcinoma: Etiology and Current and Future Drugs. *J. Clin. Exp. Hepatol.* **2019**, *9*, 221–232. [[CrossRef](#)]
828. Aspeslagh, S.; Shailubhai, K.; Bahleda, R.; Gazzah, A.; Varga, A.; Hollebecque, A.; Massard, C.; Spreafico, A.; Reni, M.; Soria, J.-C. Phase I dose-escalation study of miliclib in combination with gemcitabine in patients with refractory solid tumors. *Cancer Chemother. Pharmacol.* **2017**, *79*, 1257–1265. [[CrossRef](#)]
829. Bolin, S.; Borgenvik, A.; Persson, C.U.; Sundström, A.; Qi, J.; Bradner, J.E.; Weiss, W.A.; Cho, Y.-J.; Weishaupt, H.; Swartling, F.J. Combined BET bromodomain and CDK2 inhibition in MYC-driven medulloblastoma. *Oncogene* **2018**, *37*, 2850–2862. [[CrossRef](#)]
830. Albanese, C.; Alzani, R.; Amboldi, N.; Degrassi, A.; Festuccia, C.; Fiorentini, F.; Gravina, G.L.; Mercurio, C.; Pastori, W.; Brasca, M.; et al. Anti-tumour efficacy on glioma models of PHA-848125, a multi-kinase inhibitor able to cross the blood-brain barrier. *Br. J. Pharmacol.* **2013**, *169*, 156–166. [[CrossRef](#)]
831. Martínez-Chávez, A.; Broeders, J.; Lebre, M.C.; Tibben, M.T.; Rosing, H.; Beijnen, J.H.; Schinkel, A.H. The role of drug efflux and uptake transporters ABCB1 (P-gp), ABCG2 (BCRP) and OATP1A/1B and of CYP3A4 in the pharmacokinetics of the CDK inhibitor miliclib. *Eur. J. Pharm. Sci.* **2021**, *159*, 105740. [[CrossRef](#)]
832. Smolewski, P. Terameprocol, a novel site-specific transcription inhibitor with anticancer activity. *IDrugs.* **2008**, *11*, 204–214.
833. Castro-Gamero, A.M.; Borges, K.S.; Moreno, D.A.; Suazo, V.K.; Fujinami, M.M.; Queiroz, R.D.P.G.; de Oliveira, H.F.; Carlotti, C.G.; Scrideli, C.; Tone, L.G. Tetra-O-methyl nordihydroguaiaretic acid, an inhibitor of Sp1-mediated survivin transcription, induces apoptosis and acts synergistically with chemo-radiotherapy in glioblastoma cells. *Investig. New Drugs* **2013**, *31*, 858–870. [[CrossRef](#)]
834. Akinaga, S.; Sugiyama, K.; Akiyama, T. UCN-01 (7-hydroxystaurosporine) and other indolocarbazole compounds: A new generation of anti-cancer agents for the new century? *Anti-Cancer Drug Des.* **2000**, *15*, 43–52.
835. Shao, R.-G.; Shimizu, T.; Pommier, Y. 7-Hydroxystaurosporine (UCN-01) Induces Apoptosis in Human Colon Carcinoma and Leukemia Cells Independently of p53. *Exp. Cell Res.* **1997**, *234*, 388–397. [[CrossRef](#)]

836. Lien, W.C.; Chen, T.Y.; Sheu, S.Y.; Lin, T.C.; Kang, F.C.; Yu, C.H.; Kuan, T.S.; Huang, B.M.; Wang, C.Y. 7-hydroxy-staurosporine, UCN-01, induces DNA damage response, and autophagy in human osteosarcoma U2-OS cells. *J. Cell. Biochem.* **2018**, *119*, 4729–4741. [[CrossRef](#)]
837. Shankar, S.L.; Krupski, M.; Parashar, B.; Okwuaka, C.; O'Guin, K.; Mani, S.; Shafit-Zagardo, B. UCN-01 alters phosphorylation of Akt and GSK3beta and induces apoptosis in six independent human neuroblastoma cell lines. *J. Neurochem.* **2004**, *90*, 702–711. [[CrossRef](#)]
838. Zhang, J.; Liu, S.; Ye, Q.; Pan, J. Transcriptional inhibition by CDK7/9 inhibitor SNS-032 abrogates oncogene addiction and reduces liver metastasis in uveal melanoma. *Mol. Cancer* **2019**, *18*, 1–17. [[CrossRef](#)]
839. Wu, Y.; Chen, C.; Sun, X.; Shi, X.; Jin, B.; Ding, K.; Yeung, S.-C.J.; Pan, J. Cyclin-Dependent Kinase 7/9 Inhibitor SNS-032 Abrogates FIP1-like-1 Platelet-Derived Growth Factor Receptor  $\alpha$  and Bcr-Abl Oncogene Addiction in Malignant Hematologic Cells. *Clin. Cancer Res.* **2012**, *18*, 1966–1978. [[CrossRef](#)] [[PubMed](#)]
840. Xie, G.; Tang, H.; Wu, S.; Chen, J.; Liu, J.; Liao, C. The cyclin-dependent kinase inhibitor SNS-032 induces apoptosis in breast cancer cells via depletion of Mcl-1 and X-linked inhibitor of apoptosis protein and displays antitumor activity in vivo. *Int. J. Oncol.* **2014**, *45*, 804–812. [[CrossRef](#)] [[PubMed](#)]
841. Scrae, S.F.; Kierstan, P.; Borgognoni, J.; Denny, S.; Wayne, J.; Bentley, C.; Cansfield, A.D.; Jackson, P.S.; Lockie, A.M.; Curtin, N.J.; et al. Transient treatment with CDK inhibitors eliminates proliferative potential even when their abilities to evoke apoptosis and DNA damage are blocked. *Cell Cycle* **2008**, *7*, 3898–3907. [[CrossRef](#)] [[PubMed](#)]
842. Löschmann, N.; Michaelis, M.; Rothweiler, F.; Voges, Y.; Balónová, B.; Blight, B.A.; Jr, J.C. ABCB1 as predominant resistance mechanism in cells with acquired SNS-032 resistance. *Oncotarget* **2016**, *7*, 58051–58064. [[CrossRef](#)] [[PubMed](#)]
843. Löschmann, N.; Michaelis, M.; Rothweiler, F.; Zehner, R.; Cinatl, J.; Voges, Y.; Sharifi, M.; Riecken, K.; Meyer, J.; von Deimling, A.; et al. Testing of SNS-032 in a Panel of Human Neuroblastoma Cell Lines with Acquired Resistance to a Broad Range of Drugs. *Transl. Oncol.* **2013**, *6*, 685–IN18. [[CrossRef](#)]
844. Saha, D.; Ali, M.A.; Reis, A.; Ding, L.-H.; Story, M.D.; Habib, A.A.; Chattopadhyay, A. SNS-032 prevents hypoxia-mediated glioblastoma cell invasion by inhibiting hypoxia inducible factor-1 $\alpha$  expression. *Int. J. Oncol.* **2009**, *34*, 1051–1060. [[CrossRef](#)]
845. Ali, M.A.; Choy, H.; Habib, A.A.; Saha, D. SNS-032 Prevents Tumor Cell-Induced Angiogenesis By Inhibiting Vascular Endothelial Growth Factor. *Neoplasia* **2007**, *9*, 370–381. [[CrossRef](#)]
846. Gaevyí, M.D.; Seif, M.N. Cerebral blood circulation during angiotensin-converting enzyme inhibition. *Fiziol Zh Im I M Sechenova* **1993**, *79*, 74–78.
847. Zhang, M.; Zhang, L.; Hei, R.; Li, X.; Cai, H.; Wu, X.; Zheng, Q.; Cai, C. CDK inhibitors in cancer therapy, an overview of recent development. *Am. J. Cancer Res.* **2021**, *11*, 1913–1935.
848. Aldoss, I.T.; Tashi, T.; Ganti, A.K. Seliciclib in malignancies. *Expert Opin. Investig. Drugs* **2009**, *18*, 1957–1965. [[CrossRef](#)]
849. Wu, T.; Qin, Z.; Tian, Y.; Wang, J.; Xu, C.; Li, Z.; Bian, J. Recent Developments in the Biology and Medicinal Chemistry of CDK9 Inhibitors: An Update. *J. Med. Chem.* **2020**, *63*, 13228–13257. [[CrossRef](#)]
850. Khalil, H.S.; Mitev, V.; Vlaykova, T.; Cavicchi, L.; Zhelev, N. Discovery and development of Seliciclib. How systems biology approaches can lead to better drug performance. *J. Biotechnol.* **2015**, *202*, 40–49. [[CrossRef](#)]
851. Tirado, O.M.; Mateo-Lozano, S.; Notario, V. Roscovitine Is an Effective Inducer of Apoptosis of Ewing's Sarcoma Family Tumor Cells In vitro and In vivo. *Cancer Res* **2005**, *65*, 9320–9327. [[CrossRef](#)]
852. Iurisci, I.; Filipski, E.; Reinhardt, J.; Bach, S.; Gianella-Borradori, A.; Iacobelli, S.; Meijer, L.; Lévi, F. Improved Tumor Control through Circadian Clock Induction by Seliciclib, a Cyclin-Dependent Kinase Inhibitor. *Cancer Res* **2006**, *66*, 10720–10728. [[CrossRef](#)]
853. Pizarro, J.G.; Folch, J.; Junyent, F.; Verdager, E.; Auladell, C.; Beas-Zarate, C.; Pallàs, M.; Camins, A. Antiapoptotic effects of roscovitine on camptothecin-induced DNA damage in neuroblastoma cells. *Apoptosis* **2011**, *16*, 536–550. [[CrossRef](#)]
854. Garrofé-Ochoa, X.; Cosiáls, A.M.; Ribas, J.; Gil, J.; Boix, J. Transcriptional modulation of apoptosis regulators by roscovitine and related compounds. *Apoptosis* **2011**, *16*, 660–670. [[CrossRef](#)]
855. Chen, Y.; Tsai, Y.-H.; Tseng, S.-H. Inhibition of cyclin-dependent kinase 1-induced cell death in neuroblastoma cells through the microRNA-34a–MYCN–survivin pathway. *Surgery* **2012**, *153*, 4–16. [[CrossRef](#)]
856. Ribas, J.; Boix, J.; Meijer, L. (R)-Roscovitine (CYC202, Seliciclib) sensitizes SH-SY5Y neuroblastoma cells to nutlin-3-induced apoptosis. *Exp. Cell Res.* **2006**, *312*, 2394–2400. [[CrossRef](#)]
857. Krüger, K.; Geist, K.; Stuhldreier, F.; Schumacher, L.; Blümel, L.; Remke, M.; Wesselborg, S.; Stork, B.; Klöcker, N.; Bormann, S.; et al. Multiple DNA damage-dependent and DNA damage-independent stress responses define the outcome of ATR/Chk1 targeting in medulloblastoma cells. *Cancer Lett.* **2018**, *430*, 34–46. [[CrossRef](#)]
858. Bhatia, B.; Hsieh, M.; Kenney, A.M.; Nahlé, Z. Mitogenic Sonic hedgehog signaling drives E2F1-dependent lipogenesis in progenitor cells and medulloblastoma. *Oncogene* **2010**, *30*, 410–422. [[CrossRef](#)]
859. Braal, C.L.; Jongbloed, E.M.; Wilting, S.M.; Mathijssen, R.H.J.; Koolen, S.L.W.; Jager, A. Inhibiting CDK4/6 in Breast Cancer with Palbociclib, Ribociclib, and Abemaciclib: Similarities and Differences. *Drugs* **2020**, *81*, 317–331. [[CrossRef](#)] [[PubMed](#)]
860. Guenther, L.M.; Dharia, N.V.; Ross, L.; Conway, A.; Robichaud, A.L.; Catlett, J.L., II; Wechsler, C.S.; Frank, E.S.; Goodale, A.; Church, A.J.; et al. A Combination CDK4/6 and IGF1R Inhibitor Strategy for Ewing Sarcoma. *Clin. Cancer Res.* **2019**, *25*, 1343–1357. [[CrossRef](#)]

861. Martinez-Monleon, A.; Öberg, H.K.; Gaarder, J.; Berbegall, A.P.; Javanmardi, N.; Djos, A.; Ussowicz, M.; Taschner-Mandl, S.; Ambros, I.M.; Øra, I.; et al. Amplification of CDK4 and MDM2: A detailed study of a high-risk neuroblastoma subgroup. *Sci. Rep.* **2022**, *12*, 1–17. [[CrossRef](#)] [[PubMed](#)]
862. Guntner, A.S.; Peyrl, A.; Mayr, L.; Englinger, B.; Berger, W.; Slavic, I.; Buchberger, W.; Gojo, J. Cerebrospinal fluid penetration of targeted therapeutics in pediatric brain tumor patients. *Acta Neuropathol. Commun.* **2020**, *8*, 78. [[CrossRef](#)] [[PubMed](#)]
863. Liang, M.-L.; Hsieh, T.-H.; Liu, Y.-R.; Chen, Y.-W.; Lee, Y.-Y.; Chang, F.-C.; Lin, S.-C.; Huang, M.-C.; Ho, D.M.-T.; Wong, T.-T.; et al. Significance of cyclin D1 overexpression in progression and radio-resistance of pediatric ependymomas. *Oncotarget* **2017**, *9*, 2527–2542. [[CrossRef](#)]
864. D’Oto, A.; Fang, J.; Jin, H.; Xu, B.; Singh, S.; Mullasseril, A.; Jones, V.; Abu-Zaid, A.; von Buttlar, X.; Cooke, B.; et al. KDM6B promotes activation of the oncogenic CDK4/6-pRB-E2F pathway by maintaining enhancer activity in MYCN-amplified neuroblastoma. *Nat. Commun.* **2021**, *12*, 7204. [[CrossRef](#)]
865. Swadi, R.R.; Sampat, K.; Herrmann, A.; Losty, P.D.; See, V.; Moss, D.J. CDK inhibitors reduce cell proliferation and reverse hypoxia-induced metastasis of neuroblastoma tumours in a chick embryo model. *Sci. Rep.* **2019**, *9*, 9136. [[CrossRef](#)]
866. Rihani, A.; Vandesompele, J.; Speleman, F.; Van Maerken, T. Inhibition of CDK4/6 as a novel therapeutic option for neuroblastoma. *Cancer Cell Int.* **2015**, *15*, 76. [[CrossRef](#)]
867. Perez, M.; Muñoz-Galván, S.; Jiménez-García, M.P.; Marín, J.J.; Carnero, A. Efficacy of CDK4 inhibition against sarcomas depends on their levels of CDK4 and p16ink4 mRNA. *Oncotarget* **2015**, *6*, 40557–40574. [[CrossRef](#)]
868. Böhm, M.J.; Marienfeld, R.; Jäger, D.; Mellert, K.; von Witzleben, A.; Brüderlein, S.; Wittau, M.; von Baer, A.; Schultheiss, M.; Mayer-Steinacker, R.; et al. Analysis of the CDK4/6 Cell Cycle Pathway in Leiomyosarcomas as a Potential Target for Inhibition by Palbociclib. *Sarcoma* **2019**, *2019*, 1–10. [[CrossRef](#)]
869. Murakami, T.; Singh, A.S.; Kiyuna, T.; Dry, S.M.; Li, Y.; James, A.W.; Igarashi, K.; Kawaguchi, K.; DeLong, J.C.; Zhang, Y.; et al. Effective molecular targeting of CDK4/6 and IGF-1R in a rare *FUS-ERG* fusion *CDKN2A*-deletion doxorubicin-resistant Ewing’s sarcoma patient-derived orthotopic xenograft (PDOX) nude-mouse model. *Oncotarget* **2016**, *7*, 47556–47564. [[CrossRef](#)]
870. Tramontana, T.F.; Marshall, M.S.; Helvie, A.E.; Schmitt, M.R.; Ivanovich, J.; Carter, J.L.; Renbarger, J.L.; Ferguson, M.J. Sustained Complete Response to Palbociclib in a Refractory Pediatric Sarcoma With *BCOR-CCNB3* Fusion and Germline *CDKN2B* Variant. *JCO Precis. Oncol.* **2020**, 466–471. [[CrossRef](#)]
871. Riess, C.; Koczan, D.; Schneider, B.; Linke, C.; del Moral, K.; Classen, C.F.; Maletzki, C. Cyclin-dependent kinase inhibitors exert distinct effects on patient-derived 2D and 3D glioblastoma cell culture models. *Cell Death Discov.* **2021**, *7*, 1–15. [[CrossRef](#)]
872. Sun, Y.; Sun, Y.; Yan, K.; Li, Z.; Xu, C.; Geng, Y.; Pan, C.; Chen, X.; Zhang, L.; Xi, Q. Potent anti-tumor efficacy of palbociclib in treatment-naïve H3.3K27M-mutant diffuse intrinsic pontine glioma. *Ebiomedicine* **2019**, *43*, 171–179. [[CrossRef](#)]
873. Morris-Hanon, O.; Marazita, M.C.; Romorini, L.; Isaja, L.; Fernandez-Espinosa, D.D.; Sevlever, G.E.; Scassa, M.E.; Videla-Richardson, G.A. Palbociclib Effectively Halts Proliferation but Fails to Induce Senescence in Patient-Derived Glioma Stem Cells. *Mol. Neurobiol.* **2019**, *56*, 7810–7821. [[CrossRef](#)]
874. Huillard, E.; Hashizume, R.; Phillips, J.J.; Griveau, A.; Ihrle, R.A.; Aoki, Y.; Nicolaidis, T.; Perry, A.; Waldman, T.; McMahon, M.; et al. Cooperative interactions of BRAF<sup>V600E</sup> kinase and *CDKN2A* locus deficiency in pediatric malignant astrocytoma as a basis for rational therapy. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 8710–8715. [[CrossRef](#)]
875. Barton, K.L.; Misuraca, K.; Cordero, F.; Dobrikova, E.; Min, H.D.; Gromeier, M.; Kirsch, D.G.; Becher, O.J. PD-0332991, a CDK4/6 Inhibitor, Significantly Prolongs Survival in a Genetically Engineered Mouse Model of Brainstem Glioma. *PLoS ONE* **2013**, *8*, e77639. [[CrossRef](#)]
876. Lallena, M.J.; Boehnke, K.; Torres, R.; Hermoso, A.; Amat, J.; Calsina, B.; De Dios, A.; Buchanan, S.; Du, J.; Beckmann, R.P.; et al. Abstract 3101: In-vitro characterization of Abemaciclib pharmacology in ER+ breast cancer cell lines. *Cancer Res* **2015**, *75*, 3101. [[CrossRef](#)]
877. Torres-Guzmán, R.; Calsina, B.; Hermoso, A.; Baquero, C.; Alvarez, B.; Amat, J.; McNulty, A.M.; Gong, X.; Boehnke, K.; Du, J.; et al. Preclinical characterization of abemaciclib in hormone receptor positive breast cancer. *Oncotarget* **2017**, *8*, 69493–69507. [[CrossRef](#)]
878. Chong, Q.-Y.; Kok, Z.-H.; Bui, N.-L.; Xiang, X.; Wong, A.L.-A.; Yong, W.-P.; Sethi, G.; Lobie, P.E.; Wang, L.; Goh, B.-C. A unique CDK4/6 inhibitor: Current and future therapeutic strategies of abemaciclib. *Pharmacol. Res.* **2020**, *156*, 104686. [[CrossRef](#)]
879. Goetz, M.P.; Toi, M.; Campone, M.; Sohn, J.; Paluch-Shimon, S.; Huober, J.; Park, I.H.; Trédan, O.; Chen, S.-C.; Manso, L.; et al. MONARCH 3: Abemaciclib As Initial Therapy for Advanced Breast Cancer. *J. Clin. Oncol.* **2017**, *35*, 3638–3646. [[CrossRef](#)] [[PubMed](#)]
880. Dowless, M.S.; Lowery, C.D.; Shackelford, T.J.; Renschler, M.; Stephens, J.R.; Flack, R.; Blosser, W.; Gupta, S.; Stewart, J.; Webster, Y.; et al. Abemaciclib Is Active in Preclinical Models of Ewing Sarcoma via Multipronged Regulation of Cell Cycle, DNA Methylation, and Interferon Pathway Signaling. *Clin. Cancer Res.* **2018**, *24*, 6028–6039. [[CrossRef](#)] [[PubMed](#)]
881. Wang, D.; Bao, H. Abemaciclib is synergistic with doxorubicin in osteosarcoma pre-clinical models via inhibition of CDK4/6–Cyclin D–Rb pathway. *Cancer Chemother. Pharmacol.* **2021**, *89*, 31–40. [[CrossRef](#)] [[PubMed](#)]
882. Schubert, N.A.; Schild, L.; van Oirschot, S.; Keller, K.M.; Alles, L.K.; Vernooij, L.; Nulle, M.E.; Dolman, M.E.M.; van den Boogaard, M.L.; Molenaar, J.J. Combined targeting of the p53 and pRb pathway in neuroblastoma does not lead to synergistic responses. *Eur. J. Cancer* **2020**, *142*, 1–9. [[CrossRef](#)] [[PubMed](#)]
883. Cao, Y.; Li, X.; Kong, S.; Shang, S.; Qi, Y. CDK4/6 inhibition suppresses tumour growth and enhances the effect of temozolomide in glioma cells. *J. Cell. Mol. Med.* **2020**, *24*, 5135–5145. [[CrossRef](#)]



884. Mayr, L.; Guntner, A.S.; Madlener, S.; Schmook, M.T.; Peyrl, A.; Azizi, A.A.; Dieckmann, K.; Reisinger, D.; Stepien, N.M.; Schramm, K.; et al. Cerebrospinal Fluid Penetration and Combination Therapy of Entrectinib for Disseminated *ROS1/NTRK*-Fusion Positive Pediatric High-Grade Glioma. *J. Pers. Med.* **2020**, *10*, 290. [[CrossRef](#)]
885. Stewart, E.; McEvoy, J.; Wang, H.; Chen, X.; Honnell, V.; Ocarz, M.; Gordon, B.; Dapper, J.; Blankenship, K.; Yang, Y.; et al. Identification of Therapeutic Targets in Rhabdomyosarcoma through Integrated Genomic, Epigenomic, and Proteomic Analyses. *Cancer Cell* **2018**, *34*, 411–426.e19. [[CrossRef](#)]
886. Rana, S.; Kour, S.; Sonawane, Y.A.; Robb, C.M.; Contreras, J.I.; Kizhake, S.; Zahid, M.; Karpf, A.R.; Natarajan, A. Symbiotic prodrugs (SymProDs) dual targeting of NFkappaB and CDK. *Chem. Biol. Drug Des.* **2020**, *96*, 773–784. [[CrossRef](#)]
887. Squires, M.S.; Feltell, R.E.; Wallis, N.G.; Lewis, E.J.; Smith, D.-M.; Cross, D.M.; Lyons, J.F.; Thompson, N.T. Biological characterization of AT7519, a small-molecule inhibitor of cyclin-dependent kinases, in human tumor cell lines. *Mol. Cancer Ther.* **2009**, *8*, 324–332. [[CrossRef](#)]
888. Oghabi, M.; Safaroghli-Azar, A.; Pourbagheri-Sigaroodi, A.; Sayyadi, M.; Hamidpour, M.; Mohammadi, M.H.; Bashash, D. Anti-proliferative effects of a small molecule inhibitor of CDK AT7519 on chronic myeloid leukemia (CML) cells through halting the transition of cells from G2/M phase of the cell cycle. *Biocell* **2020**, *44*, 183–192. [[CrossRef](#)]
889. Dolman, M.E.M.; Poon, E.; Ebus, M.E.; Hartog, I.J.D.; van Noesel, C.J.; Jamin, Y.; Hallsworth, A.; Robinson, S.P.; Petrie, K.; Sparidans, R.W.; et al. Cyclin-Dependent Kinase Inhibitor AT7519 as a Potential Drug for MYCN-Dependent Neuroblastoma. *Clin. Cancer Res.* **2015**, *21*, 5100–5109. [[CrossRef](#)]
890. Steegmaier, M.; Hoffmann, M.; Baum, A.; Lénárt, P.; Petronczki, M.; Krššák, M.; Gürtler, U.; Garin-Chesa, P.; Lieb, S.; Quant, J.; et al. BI 2536, a Potent and Selective Inhibitor of Polo-like Kinase 1, Inhibits Tumor Growth In Vivo. *Curr. Biol.* **2007**, *17*, 316–322. [[CrossRef](#)]
891. Pezuk, J.A.; Brassesco, M.S.; Morales, A.G.; de Oliveira, J.C.; de Oliveira, H.F.; Scrideli, C.A.; Tone, L.G. Inhibition of Polo-Like Kinase 1 Induces Cell Cycle Arrest and Sensitizes Glioblastoma Cells to Ionizing Radiation. *Cancer Biotherapy Radiopharm.* **2013**, *28*, 516–522. [[CrossRef](#)]
892. A Pezuk, J.; Brassesco, M.S.; Morales, A.G.; de Oliveira, J.C.; Queiroz, R.G.D.P.; Machado, H.R.; Carlotti, C.G.; Neder, L.; A Scrideli, C.; Tone, L.G. Polo-like kinase 1 inhibition causes decreased proliferation by cell cycle arrest, leading to cell death in glioblastoma. *Cancer Gene Ther.* **2013**, *20*, 499–506. [[CrossRef](#)]
893. Pezuk, J.A.; Brassesco, M.S.; Ramos, P.M.M.; Scrideli, C.A.; Tone, L.G.; Pezuk, M.S.B.J.A. Polo-Like Kinase 1 Pharmacological Inhibition as Monotherapy or in Combination: Comparative Effects of Polo-Like Kinase 1 Inhibition in Medulloblastoma Cells. *Anti-Cancer Agents Med. Chem.* **2017**, *17*, 1278–1291. [[CrossRef](#)]
894. Yunoki, T.; Tabuchi, Y.; Hayashi, A.; Kondo, T. Inhibition of Polo-Like Kinase 1 Promotes Hyperthermia Sensitivity via Inactivation of Heat Shock Transcription Factor 1 in Human Retinoblastoma Cells. *Investig. Ophthalmology Vis. Sci.* **2013**, *54*, 8353–8363. [[CrossRef](#)]
895. Li, Z.; Yang, C.; Li, X.; Du, X.; Tao, Y.; Ren, J.; Fang, F.; Xie, Y.; Li, M.; Qian, G.; et al. The dual role of BI 2536, a small-molecule inhibitor that targets PLK1, in induction of apoptosis and attenuation of autophagy in neuroblastoma cells. *J. Cancer* **2020**, *11*, 3274–3287. [[CrossRef](#)]
896. Grinshtein, N.; Datti, A.; Fujitani, M.; Uehling, D.; Prakesch, M.; Isaac, M.; Irwin, M.S.; Wrana, J.L.; Al-Awar, R.; Kaplan, D.R. Small Molecule Kinase Inhibitor Screen Identifies Polo-Like Kinase 1 as a Target for Neuroblastoma Tumor-Initiating Cells. *Cancer Res* **2011**, *71*, 1385–1395. [[CrossRef](#)]
897. Hsieh, C.-H.; Yeh, H.-N.; Huang, C.-T.; Wang, W.-H.; Hsu, W.-M.; Huang, H.-C.; Juan, H.-F. BI-2536 Promotes Neuroblastoma Cell Death via Minichromosome Maintenance Complex Components 2 and 10. *Pharmaceutics* **2021**, *15*, 37. [[CrossRef](#)]
898. Morales, A.G.; Brassesco, M.S.; Pezuk, J.A.; Oliveira, J.C.; Montaldi, A.P.; Sakamoto-Hojo, E.T.; Scrideli, C.A.; Tone, L.G. BI 2536-mediated PLK1 inhibition suppresses HOS and MG-63 osteosarcoma cell line growth and clonogenicity. *Anti-Cancer Drugs* **2011**, *22*, 995–1001. [[CrossRef](#)]
899. Liu, X.; Choy, E.; Harmon, D.; Yang, S.; Yang, C.; Mankin, H.; Hornicek, F.J.; Duan, Z. Inhibition of polo-like kinase 1 leads to the suppression of osteosarcoma cell growth in vitro and in vivo. *Anti-Cancer Drugs* **2011**, *22*, 444–453. [[CrossRef](#)] [[PubMed](#)]
900. Thalhammer, V.; Lopez-Garcia, L.A.; Herrero-Martin, D.; Hecker, R.; Laubscher, D.; Gierisch, M.E.; Wachtel, M.; Bode, P.; Nanni, P.; Blank, B.; et al. PLK1 Phosphorylates PAX3-FOXO1, the Inhibition of Which Triggers Regression of Alveolar Rhabdomyosarcoma. *Cancer Res* **2015**, *75*, 98–110. [[CrossRef](#)] [[PubMed](#)]
901. Stehle, A.; Hügler, M.; Fulda, S. Eribulin synergizes with Polo-like kinase 1 inhibitors to induce apoptosis in rhabdomyosarcoma. *Cancer Lett.* **2015**, *365*, 37–46. [[CrossRef](#)] [[PubMed](#)]
902. Valsasina, B.; Beria, I.; Alli, C.; Alzani, R.; Avanzi, N.; Ballinari, D.; Cappella, P.; Caruso, M.; Casolaro, A.; Ciavolella, A.; et al. NMS-P937, an Orally Available, Specific Small-Molecule Polo-like Kinase 1 Inhibitor with Antitumor Activity in Solid and Hematologic Malignancies. *Mol. Cancer Ther.* **2012**, *11*, 1006–1016. [[CrossRef](#)]
903. Sero, V.; Tavanti, E.; Vella, S.; Hattinger, C.M.; Fanelli, M.; Michelacci, F.; Versteeg, R.; Valsasina, B.; Gudeman, B.; Picci, P.; et al. Targeting polo-like kinase 1 by NMS-P937 in osteosarcoma cell lines inhibits tumor cell growth and partially overcomes drug resistance. *Investig. New Drugs* **2014**, *32*, 1167–1180. [[CrossRef](#)]
904. Wang, D.; Veo, B.; Pierce, A.; Fosmire, S.; Madhavan, K.; Balakrishnan, I.; Donson, A.; Alimova, I.; Sullivan, K.D.; Joshi, M.; et al. A novel PLK1 inhibitor onvansertib effectively sensitizes MYC-driven medulloblastoma to radiotherapy. *Neuro-Oncology* **2021**, *24*, 414–426. [[CrossRef](#)]



905. Wu, C.-P.; Hsiao, S.-H.; Luo, S.-Y.; Tuo, W.-C.; Su, C.-Y.; Li, Y.-Q.; Huang, Y.-H.; Hsieh, C.-H. Overexpression of Human ABCB1 in Cancer Cells Leads to Reduced Activity of GSK461364, a Specific Inhibitor of Polo-like Kinase 1. *Mol. Pharm.* **2014**, *11*, 3727–3736. [[CrossRef](#)]
906. Pajtlér, K.W.; Sadowski, N.; Ackermann, S.; Althoff, K.; Schönbeck, K.; Batzke, K.; Schäfers, S.; Odersky, A.; Heukamp, L.; Astrahantseff, K.; et al. The GSK461364 PLK1 inhibitor exhibits strong antitumoral activity in preclinical neuroblastoma models. *Oncotarget* **2016**, *8*, 6730–6741. [[CrossRef](#)]
907. Chou, Y.-S.; Yen, C.-C.; Chen, W.-M.; Lin, Y.-C.; Wen, Y.-S.; Ke, W.-T.; Wang, J.-Y.; Liu, C.-Y.; Yang, M.-H.; Chen, T.-H.; et al. Cytotoxic mechanism of PLK1 inhibitor GSK461364 against osteosarcoma: Mitotic arrest, apoptosis, cellular senescence, and synergistic effect with paclitaxel. *Int. J. Oncol.* **2016**, *48*, 1187–1194. [[CrossRef](#)]
908. Bogado, R.F.; Pezuk, J.A.; de Oliveira, H.F.; Tone, L.G.; Brassesco, M.S. BI 6727 and GSK461364 suppress growth and radiosensitize osteosarcoma cells, but show limited cytotoxic effects when combined with conventional treatments. *Anti-Cancer Drugs* **2015**, *26*, 56–63. [[CrossRef](#)]
909. Gorlick, R.; Kolb, E.A.; Keir, S.T.; Maris, J.M.; Reynolds, C.P.; Kang, M.H.; Carol, H.; Lock, R.; Billups, C.A.; Kurmasheva, R.T.; et al. Initial testing (stage 1) of the polo-like kinase inhibitor volasertib (BI 6727), by the Pediatric Preclinical Testing Program. *Pediatr. Blood Cancer* **2013**, *61*, 158–164. [[CrossRef](#)]
910. Carol, H.; Boehm, I.; Reynolds, C.P.; Kang, M.H.; Maris, J.M.; Morton, C.L.; Gorlick, R.; Kolb, E.A.; Keir, S.T.; Wu, J.; et al. Efficacy and pharmacokinetic/pharmacodynamic evaluation of the Aurora kinase A inhibitor MLN8237 against preclinical models of pediatric cancer. *Cancer Chemother. Pharmacol.* **2011**, *68*, 1291–1304. [[CrossRef](#)]
911. Maris, J.M.; Bs, C.L.M.; Gorlick, R.; Kolb, E.A.; Lock, R.; Carol, H.; Keir, S.T.; Reynolds, C.P.; Kang, M.H.; Wu, J.; et al. Initial testing of the aurora kinase a inhibitor MLN8237 by the Pediatric Preclinical Testing Program (PPTP). *Pediatr. Blood Cancer* **2010**, *55*, 26–34. [[CrossRef](#)]
912. Boi, D.; Souvalidou, F.; Capelli, D.; Polverino, F.; Marini, G.; Montanari, R.; Pochetti, G.; Tramonti, A.; Contestabile, R.; Trisciuglio, D.; et al. PHA-680626 Is an Effective Inhibitor of the Interaction between Aurora-A and N-Myc. *Int. J. Mol. Sci.* **2021**, *22*, 13122. [[CrossRef](#)]
913. Brockmann, M.; Poon, E.; Berry, T.; Carstensen, A.; Deubzer, H.E.; Rycak, L.; Jamin, Y.; Thway, K.; Robinson, S.P.; Roels, F.; et al. Small Molecule Inhibitors of Aurora-A Induce Proteasomal Degradation of N-Myc in Childhood Neuroblastoma. *Cancer Cell* **2013**, *24*, 75–89. [[CrossRef](#)]
914. Geron, L.; Borges, K.S.; Andrade, A.F.; Suazo, V.K.; Scrideli, C.A.; Tone, L.G. Antitumour activity of AMG 900 alone or in combination with histone deacetylase inhibitor SaHa on medulloblastoma cell lines. *Neurol. Res.* **2015**, *37*, 703–711. [[CrossRef](#)]
915. Hanahan, D. Hallmarks of Cancer: New Dimensions. *Cancer Discov.* **2022**, *12*, 31–46. [[CrossRef](#)]
916. Grignani, G.; Palmerini, E.; Ferraresi, V.; D’Ambrosio, L.; Bertulli, R.; Asaftei, S.D.; Tamburini, A.; Pignochino, Y.; Sangiolo, D.; Marchesi, E.; et al. Sorafenib and everolimus for patients with unresectable high-grade osteosarcoma progressing after standard treatment: A non-randomised phase 2 clinical trial. *Lancet Oncol.* **2015**, *16*, 98–107. [[CrossRef](#)]
917. Liu, Y.; Li, Y.; Wang, Y.; Lin, C.; Zhang, D.; Chen, J.; Ouyang, L.; Wu, F.; Zhang, J.; Chen, L. Recent progress on vascular endothelial growth factor receptor inhibitors with dual targeting capabilities for tumor therapy. *J. Hematol. Oncol.* **2022**, *15*, 1–28. [[CrossRef](#)]
918. Chambers, A.; Kundranda, M.; Rao, S.; Niu, J. Anti-angiogenesis Revisited: Combination with Immunotherapy in Solid Tumors. *Lung Cancer* **1912**, *23*, 100. [[CrossRef](#)]
919. Inaba, H.; Rubnitz, J.E.; Coustan-Smith, E.; Li, L.; Furmanski, B.D.; Mascara, G.P.; Heym, K.M.; Christensen, R.; Onciu, M.; Shurtleff, S.A.; et al. Phase I Pharmacokinetic and Pharmacodynamic Study of the Multikinase Inhibitor Sorafenib in Combination With Clofarabine and Cytarabine in Pediatric Relapsed/Refractory Leukemia. *J. Clin. Oncol.* **2011**, *29*, 3293–3300. [[CrossRef](#)] [[PubMed](#)]
920. Grignani, G.; Palmerini, E.; Dileo, P.; Asaftei, S.D.; D’Ambrosio, L.; Pignochino, Y.; Mercuri, M.; Picci, P.; Fagioli, F.; Casali, P.G.; et al. A phase II trial of sorafenib in relapsed and unresectable high-grade osteosarcoma after failure of standard multimodal therapy: An Italian Sarcoma Group study. *Ann. Oncol.* **2011**, *23*, 508–516. [[CrossRef](#)] [[PubMed](#)]
921. Keino, D.; Yokosuka, T.; Hirose, A.; Sakurai, Y.; Nakamura, W.; Fujita, S.; Hayashi, A.; Miyagawa, N.; Iwasaki, F.; Hamanoue, S.; et al. Pilot study of the combination of sorafenib and fractionated irinotecan in pediatric relapse/refractory hepatic cancer (FINEX pilot study). *Pediatr. Blood Cancer* **2020**, *67*, e28655. [[CrossRef](#)] [[PubMed](#)]
922. Geller, J.I.; Fox, E.; Turpin, B.K.; Goldstein, S.L.; Liu, X.; Minard, C.G.; Kudgus, R.A.; Reid, J.M.; Berg, S.L.; Weigel, B.J. A study of axitinib, a VEGF receptor tyrosine kinase inhibitor, in children and adolescents with recurrent or refractory solid tumors: A Children’s Oncology Group phase 1 and pilot consortium trial (ADVL1315). *Cancer* **2018**, *124*, 4548–4555. [[CrossRef](#)] [[PubMed](#)]
923. Daw, N.C.; Furman, W.L.; Stewart, C.F.; Iacono, L.C.; Krailo, M.; Bernstein, M.L.; Dancey, J.E.; Speights, R.A.; Blaney, S.M.; Croop, J.M.; et al. Phase I and Pharmacokinetic Study of Gefitinib in Children With Refractory Solid Tumors: A Children’s Oncology Group Study. *J. Clin. Oncol.* **2005**, *23*, 6172–6180. [[CrossRef](#)]
924. Pollack, I.F.; Stewart, C.F.; Kocak, M.; Poussaint, T.Y.; Broniscer, A.; Banerjee, A.; Douglas, J.G.; Kun, L.E.; Boyett, J.M.; Geyer, J.R. A phase II study of gefitinib and irradiation in children with newly diagnosed brainstem gliomas: A report from the Pediatric Brain Tumor Consortium. *Neuro-Oncology* **2011**, *13*, 290–297. [[CrossRef](#)]
925. Subbiah, V.; Wolf, J.; Konda, B.; Kang, H.; Spira, A.; Weiss, J.; Takeda, M.; Ohe, Y.; Khan, S.; Ohashi, K.; et al. Tumour-agnostic efficacy and safety of selpercatinib in patients with RET fusion-positive solid tumours other than lung or thyroid tumours (LIBRETTO-001): A phase 1/2, open-label, basket trial. *Lancet Oncol.* **2022**, *23*, 1261–1273. [[CrossRef](#)]

926. Del Rivero, J.; Edgerly, M.; Ward, J.; Madan, R.A.; Balasubramaniam, S.; Fojo, T.; Gramza, A.W. Phase I/II Trial of Vandetanib and Bortezomib in Adults with Locally Advanced or Metastatic Medullary Thyroid Cancer. *Oncol.* **2018**, *24*, 16–e14. [[CrossRef](#)]
927. Subbiah, V.; Wolf, J.; Konda, B.; Kang, H.; Spira, A.I.; Weiss, J.; Takeda, M.; Ohe, Y.; Khan, S.A.; Ohashi, K.; et al. Tumor agnostic efficacy of selpercatinib in patients with *RET* fusion+ solid tumors: A global, multicenter, registrational trial update (LIBRETTO-001). *J. Clin. Oncol.* **2022**, *40*, 3094. [[CrossRef](#)]
928. Subbiah, V.; Cassier, P.A.; Siena, S.; Garralda, E.; Paz-Ares, L.; Garrido, P.; Nadal, E.; Vuky, J.; Lopes, G.; Kalemkerian, G.P.; et al. Pan-cancer efficacy of pralsetinib in patients with *RET* fusion-positive solid tumors from the phase 1/2 ARROW trial. *Nat. Med.* **2022**, *28*, 1640–1645. [[CrossRef](#)]
929. Mossé, Y.P.; Lim, M.S.; Voss, S.D.; Wilner, K.; Ruffner, K.; Laliberte, J.; Rolland, D.; Balis, F.M.; Maris, J.M.; Weigel, B.J.; et al. Safety and activity of crizotinib for paediatric patients with refractory solid tumours or anaplastic large-cell lymphoma: A Children's Oncology Group phase 1 consortium study. *Lancet Oncol.* **2013**, *14*, 472–480. [[CrossRef](#)]
930. Fukano, R.; Mori, T.; Sekimizu, M.; Choi, I.; Kada, A.; Saito, A.M.; Asada, R.; Takeuchi, K.; Terauchi, T.; Tateishi, U.; et al. Alectinib for relapsed or refractory anaplastic lymphoma kinase-positive anaplastic large cell lymphoma: An open-label phase II trial. *Cancer Sci.* **2020**, *111*, 4540–4547. [[CrossRef](#)]
931. Franz, D.N.; Belousova, E.; Sparagana, S.; Bebin, E.M.; Frost, M.; Kuperman, R.; Witt, O.; Kohrman, M.H.; Flamini, J.R.; Wu, J.Y.; et al. Efficacy and safety of everolimus for subependymal giant cell astrocytomas associated with tuberous sclerosis complex (EXIST-1): A multicentre, randomised, placebo-controlled phase 3 trial. *Lancet* **2013**, *381*, 125–132. [[CrossRef](#)]
932. Adashek, J.J.; Menta, A.K.; Reddy, N.K.; Desai, A.P.; Roszik, J.; Subbiah, V. Tissue-Agnostic Activity of BRAF plus MEK Inhibitor in BRAF V600-Mutant Tumors. *Mol. Cancer Ther.* **2022**, *21*, 871–878. [[CrossRef](#)]
933. Bouffet, E.; Hansford, J.; Garré, M.L.; Hara, J.; Plant-Fox, A.; Aerts, I.; Locatelli, F.; Van der Lugt, J.; Papusha, L.; Sahm, F.; et al. Primary analysis of a phase II trial of dabrafenib plus trametinib (dab + tram) in *BRAF* V600-mutant pediatric low-grade glioma (pLGG). *J. Clin. Oncol.* **2022**, *40*, LBA2002. [[CrossRef](#)]
934. Hargrave, D.R.; Bouffet, E.; Tabori, U.; Broniscer, A.; Cohen, K.J.; Hansford, J.R.; Geoerger, B.; Hingorani, P.; Dunkel, I.J.; Russo, M.W.; et al. Efficacy and Safety of Dabrafenib in Pediatric Patients with *BRAF* V600 Mutation-Positive Relapsed or Refractory Low-Grade Glioma: Results from a Phase I/IIa Study. *Clin. Cancer Res.* **2019**, *25*, 7303–7311. [[CrossRef](#)]
935. Lau, N.; Feldkamp, M.M.; Roncari, L.; Loehr, A.H.; Shannon, P.; Gutmann, D.H.; Guha, A. Loss of Neurofibromin Is Associated with Activation of RAS/MAPK and PI3-K/AKT Signaling in a Neurofibromatosis 1 Astrocytoma. *J. Neuropathol. Exp. Neurol.* **2000**, *59*, 759–767. [[CrossRef](#)]
936. Gross, A.M.; Wolters, P.L.; Dombi, E.; Baldwin, A.; Whitcomb, P.; Fisher, M.J.; Weiss, B.; Kim, A.; Bornhorst, M.; Shah, A.C.; et al. Selumetinib in Children with Inoperable Plexiform Neurofibromas. *N. Engl. J. Med.* **2020**, *382*, 1430–1442. [[CrossRef](#)]
937. Bautista, F.; Paoletti, X.; Rubino, J.; Brard, C.; Rezai, K.; Nebchi, S.; Andre, N.; Aerts, I.; De Carli, E.; van Eijkelenburg, N.; et al. Phase I or II Study of Ribociclib in Combination With Topotecan-Temozolomide or Everolimus in Children With Advanced Malignancies: Arms A and B of the AcSé-ESMART Trial. *J. Clin. Oncol.* **2021**, *39*, 3546–3560. [[CrossRef](#)]
938. Yang, Y.; Li, S.; Wang, Y.; Zhao, Y.; Li, Q. Protein tyrosine kinase inhibitor resistance in malignant tumors: Molecular mechanisms and future perspective. *Signal Transduct. Target. Ther.* **2022**, *7*, 1–36. [[CrossRef](#)]
939. Van Mater, D.; Gururangan, S.; Becher, O.; Campagne, O.; Leary, S.; Phillips, J.J.; Huang, J.; Lin, T.; Poussaint, T.Y.; Goldman, S.; et al. A phase I trial of the CDK 4/6 inhibitor palbociclib in pediatric patients with progressive brain tumors: A Pediatric Brain Tumor Consortium study (PBTC-042). *Pediatr. Blood Cancer* **2021**, *68*, e28879. [[CrossRef](#)] [[PubMed](#)]
940. Mossé, Y.P.; Lipsitz, E.; Fox, E.; Teachey, D.T.; Maris, J.M.; Weigel, B.; Adamson, P.C.; Ingle, M.A.; Ahern, C.H.; Blaney, S.M. Pediatric Phase I Trial and Pharmacokinetic Study of MLN8237, an Investigational Oral Selective Small-Molecule Inhibitor of Aurora Kinase A: A Children's Oncology Group Phase I Consortium Study. *Clin. Cancer Res.* **2012**, *18*, 6058–6064. [[CrossRef](#)] [[PubMed](#)]
941. Moreno, L.; Marshall, L.V.; Pearson, A.D.; Morland, B.; Elliott, M.; Campbell-Hewson, Q.; Makin, G.; Halford, S.E.; Acton, G.; Ross, P.; et al. A Phase I Trial of AT9283 (a Selective Inhibitor of Aurora Kinases) in Children and Adolescents with Solid Tumors: A Cancer Research UK Study. *Clin. Cancer Res.* **2015**, *21*, 267–273. [[CrossRef](#)] [[PubMed](#)]
942. Mossé, Y.P.; Fox, E.; Teachey, D.T.; Reid, J.M.; Safgren, S.L.; Carol, H.; Lock, R.B.; Houghton, P.J.; Smith, M.A.; Hall, D.; et al. A Phase II Study of Alisertib in Children with Recurrent/Refractory Solid Tumors or Leukemia: Children's Oncology Group Phase I and Pilot Consortium (ADV0921). *Clin. Cancer Res.* **2019**, *25*, 3229–3238. [[CrossRef](#)] [[PubMed](#)]
943. Feliciano, S.V.M.; Santos, M.D.O.; Pombo-De-Oliveira, M.S. Incidência e Mortalidade por Câncer entre Crianças e Adolescentes: Uma Revisão Narrativa. *Rev. Bras. Cancerol.* **2019**, *64*, 389–396. [[CrossRef](#)]
944. Verhaak, R.G.W.; Hoadley, K.A.; Purdom, E.; Wang, V.; Wilkerson, M.D.; Miller, C.R.; Ding, L.; Golub, T.; Jill, P.; Alexe, G.; et al. Integrated Genomic Analysis Identifies Clinically Relevant Subtypes of Glioblastoma Characterized by Abnormalities in PDGFRA, IDH1, EGFR, and NF1. *Cancer Cell* **2010**, *17*, 98–110. [[CrossRef](#)]
945. Maximiano, S.; Magalhães, P.; Guerreiro, M.P.; Morgado, M. Trastuzumab in the Treatment of Breast Cancer. *Biodrugs* **2016**, *30*, 75–86. [[CrossRef](#)]
946. Lee, J.; Park, Y.H. Trastuzumab deruxtecan for HER2+ advanced breast cancer. *Futur. Oncol.* **2022**, *18*, 7–19. [[CrossRef](#)]
947. Hochhaus, A.; Baccarani, M.; Silver, R.T.; Schiffer, C.; Apperley, J.F.; Cervantes, F.; Clark, R.E.; Cortes, J.E.; Deininger, M.W.; Guilhot, F.; et al. European LeukemiaNet 2020 recommendations for treating chronic myeloid leukemia. *Leukemia* **2020**, *34*, 966–984. [[CrossRef](#)]

948. Louis, D.N.; Perry, A.; Wesseling, P.; Brat, D.J.; Cree, I.A.; Figarella-Branger, D.; Hawkins, C.; Ng, H.K.; Pfister, S.M.; Reifenberger, G.; et al. The 2021 WHO Classification of Tumors of the Central Nervous System: A summary. *Neuro-Oncology* **2021**, *23*, 1231–1251. [[CrossRef](#)]
949. Nørøxe, D.S.; Poulsen, H.S.; Lassen, U. Hallmarks of glioblastoma: A systematic review. *ESMO Open* **2016**, *1*, e000144. [[CrossRef](#)]
950. Roskoski, R., Jr. Properties of FDA-approved small molecule protein kinase inhibitors: A 2020 update. *Pharmacol. Res.* **2020**, *152*, 104609. [[CrossRef](#)]
951. Roskoski, R., Jr. Classification of small molecule protein kinase inhibitors based upon the structures of their drug-enzyme complexes. *Pharmacol. Res.* **2016**, *103*, 26–48. [[CrossRef](#)]
952. Wu, P.; Nielsen, T.E.; Clausen, M.H. FDA-approved small-molecule kinase inhibitors. *Trends Pharmacol. Sci.* **2015**, *36*, 422–439. [[CrossRef](#)]
953. Lopes, L.F.; Bacchi, C.E. Imatinib treatment for gastrointestinal stromal tumour (GIST). *J. Cell. Mol. Med.* **2009**, *14*, 42–50. [[CrossRef](#)]
954. Fischer, P.M. The Design of Drug Candidate Molecules as Selective Inhibitors of Therapeutically Relevant Protein Kinases. *Curr. Med. Chem.* **2004**, *11*, 1563–1583. [[CrossRef](#)]
955. A Bogoyevitch, M.; Fairlie, D. A new paradigm for protein kinase inhibition: Blocking phosphorylation without directly targeting ATP binding. *Drug Discov. Today* **2007**, *12*, 622–633. [[CrossRef](#)]
956. Gaumann, A.K.; Kiefer, F.; Alfer, J.; Lang, S.A.; Geissler, E.K.; Breier, G. Receptor tyrosine kinase inhibitors: Are they real tumor killers? *Int. J. Cancer* **2015**, *138*, 540–554. [[CrossRef](#)]
957. Roberti, M.; Bottegoni, G. Non-ATP Competitive Protein Kinase Inhibitors. *Curr. Med. Chem.* **2010**, *17*, 2804–2821. [[CrossRef](#)]
958. Suttorp, M.; Bornhäuser, M.; Metzler, M.; Millot, F.; Schleyer, E. Pharmacology and pharmacokinetics of imatinib in pediatric patients. *Expert Rev. Clin. Pharmacol.* **2017**, *11*, 219–231. [[CrossRef](#)]
959. Angel, M.C.-G.; Julia, A.P.; María, S.B.; Luiz, G.T. G2/M inhibitors as pharmacotherapeutic opportunities for glioblastoma: The old, the new, and the future. *Cancer Biol. Med.* **2018**, *15*, 354–374. [[CrossRef](#)] [[PubMed](#)]
960. Kwak, E.L.; Bang, Y.-J.; Camidge, D.R.; Shaw, A.T.; Solomon, B.; Maki, R.G.; Ou, S.-H.I.; Dezube, B.J.; Jänne, P.A.; Costa, D.B.; et al. Anaplastic Lymphoma Kinase Inhibition in Non-Small-Cell Lung Cancer. *N. Engl. J. Med.* **2010**, *363*, 1693–1703. [[CrossRef](#)] [[PubMed](#)]
961. De Brouwer, S.; De Preter, K.; Kumps, C.; Zabrocki, P.; Porcu, M.; Westerhout, E.M.; Lakeman, A.; Vandesompele, J.; Hoebeek, J.; Van Maerken, T.; et al. Meta-analysis of Neuroblastomas Reveals a Skewed *ALK* Mutation Spectrum in Tumors with *MYCN* Amplification. *Clin. Cancer Res.* **2010**, *16*, 4353–4362. [[CrossRef](#)] [[PubMed](#)]
962. Bresler, S.C.; Wood, A.C.; Haglund, E.A.; Courtright, J.; Belcastro, L.T.; Plegaria, J.S.; Cole, K.; Toporovskaya, Y.; Zhao, H.; Carpenter, E.L.; et al. Differential Inhibitor Sensitivity of Anaplastic Lymphoma Kinase Variants Found in Neuroblastoma. *Sci. Transl. Med.* **2011**, *3*, 108ra114. [[CrossRef](#)]
963. de Sousa, G.R.; Vieira, G.M.; das Chagas, P.F.; Pezuk, J.A.; Brassesco, M.S. Should we keep rocking? Portraits from targeting Rho kinases in cancer. *Pharmacol. Res.* **2020**, *160*, 105093. [[CrossRef](#)]
964. Martinsson, T.; Eriksson, T.; Abrahamsson, J.; Caren, H.; Hansson, M.; Kogner, P.; Kamaraj, S.; Schönherr, C.; Weinmar, J.; Ruuth, K.; et al. Appearance of the Novel Activating F1174S *ALK* Mutation in Neuroblastoma Correlates with Aggressive Tumor Progression and Unresponsiveness to Therapy. *Cancer Res* **2011**, *71*, 98–105. [[CrossRef](#)]
965. Braun, T.P.; Eide, C.A.; Druker, B.J. Response and Resistance to BCR-ABL1-Targeted Therapies. *Cancer Cell* **2020**, *37*, 530–542. [[CrossRef](#)]
966. Katayama, R. Drug resistance in anaplastic lymphoma kinase-rearranged lung cancer. *Cancer Sci.* **2018**, *109*, 572–580. [[CrossRef](#)]
967. Rolfo, C.; Passiglia, F.; Castiglia, M.; E Raez, L.; Germonpre, P.; Gil-Bazo, I.; Zwaenepoel, K.; De Wilde, A.; Bronte, G.; Russo, A.; et al. *ALK* and crizotinib: After the honeymoon . . . what else? Resistance mechanisms and new therapies to overcome it. *Transl. Lung Cancer Res.* **2014**, *3*, 250–261. [[CrossRef](#)]
968. Sharma, S.V.; Haber, D.A.; Settleman, J. Cell line-based platforms to evaluate the therapeutic efficacy of candidate anticancer agents. *Nat. Rev. Cancer* **2010**, *10*, 241–253. [[CrossRef](#)]
969. Massicotte, M.P.; Sofronas, M.; Deveber, G. Difficulties in performing clinical trials of antithrombotic therapy in neonates and children. *Thromb. Res.* **2006**, *118*, 153–163. [[CrossRef](#)]
970. Bond, M.C.; Pritchard, S. Understanding clinical trials in childhood cancer. *Paediatr. Child Health* **2006**, *11*, 148–150. [[CrossRef](#)]
971. Renfro, L.A.; Ji, L.; Piao, J.; Onar-Thomas, A.; Kairalla, J.A.; Alonzo, T.A. Trial Design Challenges and Approaches for Precision Oncology in Rare Tumors: Experiences of the Children’s Oncology Group. *JCO Precis. Oncol.* **2019**, *1*, 1–13. [[CrossRef](#)]
972. Bavcar, S.; Argyle, D.J. Receptor tyrosine kinase inhibitors: Molecularly targeted drugs for veterinary cancer therapy. *Veter- Comp. Oncol.* **2012**, *10*, 163–173. [[CrossRef](#)]

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