



Zinc nanocoated seeds: an alternative to boost soybean seed germination and seedling development

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Abstract

In this study, we characterized Zn seed coats adhered on soybean seed surface [*Glycine max* (L.) Merrill] and then investigated the effects of such coatings on germination and finally monitored the transference of Zn from the coatings to the seedlings. For that, seeds were coated with ZnSO₄, 40 nm ZnO, and 5 μm ZnO at 4 mg of Zn per kg of seed. The shape and homogeneity of Zn deposits on the seed surface depended on the type of Zn source. X-ray spectroscopy unraveled that before the germination, Zn neither crossed the seed coat nor was biotransformed. Regardless of the particle size, ZnO-based treatments increased germination ratio, seedlings root, and shoot development, whereas ZnSO₄ suppressed them. After sowing, most of Zn on the seed coat was transferred to soil, while less than 13 wt% was assimilated by the seedling 14 days after sowing. Either this minor fraction of Zn was taken up during the imbibition process or absorbed by during seedling growth. Zinc oxide-based treatments represent a viable alternative to Zn supply in order to boost soybean germination and seedling development.

Keywords Zinc · Nanoparticles · Seed coating · XRF · Rhizosphere

1 Introduction

Food security is a central issue of the twentieth century [1]. The agricultural output needs to increase by 70% until 2050 [2–4] to sustain the new 2.3 billion people worldwide in a few decades [5]. A sustainable response to this demand requires higher yields from crops [6] within the same area currently cultivated [3]. Hence, adequate strategies for mineral nutrition of plants are a significant pillar to accomplish these goals [7].

Zinc (Zn) is a micronutrient required for plants [8]. It is present in more than 300 enzymes (e.g., alcohol dehydrogenase and alkaline phosphatase) playing critical roles

in the structure and mechanism of DNA-related proteins [9], carbohydrate metabolism, and maintenance of cellular membranes integrity [10]. Despite variations among compartments and across species, Zn concentration usually ranges from 20 up to 200 mg Zn kg⁻¹ dry weight shoot and leaf tissues [11], which are inward exported to the agricultural products. However, nearly 50% of arable soils worldwide are Zn deficient [12], and thus, it must be supplied through fertilization.

Zinc fertilizers are mostly applied as zinc sulphates, carbonates, and oxides [13], usually employed through soil broadcasting and foliar spraying [14]. Both approaches require driving machines across the field, which increases

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costs and causes soil compaction. Additionally, soil broadcasting spreads fertilizers everywhere, which implies losses. On the other hand, localized foliar spraying, which delivers the nutrient directly to the plant, can only be accomplished when leaves are already developed. In this latter case, plant performance might be already compromised due to the lack of zinc.

An alternative for micronutrient supply consists of seed treatments (e.g., seed soaking, coating, or priming). Seed treatment is carried out in a warehouse, and therefore, it does not require proper machines trafficking in the field. Besides reducing energetic costs, it prevents soil compaction. Furthermore, it supposedly provides the nutrients to plants embryo even before germination [15]. Adequate Zn content in seeds can ensure higher germination, boost plant development, and enhance protection against pathogens [16].

In this scenario, nanomaterials are a promising source of nutrients for plants. In principle, its size-dependent solubility can yield controlled-release fertilizers, or small particles can be entirely taken up by plants and slowly dissolve at the target tissues [17].

Literature suggests that zinc oxide nanoparticles (ZnO NPs) can be used as a Zn source for crops [18–20]. Besides, some reports with pinto bean (*Phaseolus vulgaris*) [21], maize (*Zea mays*), soybean (*Glycine max* L.), pigeon pea (*Cajanus cajan*) and ladies finger (*Abelmoschus esculentus*) seeds treated with ZnO NPs at $< 100 \text{ mg Zn kg}^{-1}$ claim that it can also grant seeds' antimicrobial protection [20], which could be a desirable double function of ZnO NPs that deserves to be further investigated. On the other hand, phytotoxicity was observed in seeds exposed to higher ZnO doses [22], and the effects of nanoparticles with both seeds and the soil are still lacking.

Some of the gaps found in the current literature on the effects of nanomaterials on plants regard the employment of realistic concentrations and the presence of adequate experimental controls, i.e., to compare nanoparticles to microparticles and soluble counterparts.

Hence, the present study aimed at: (1) characterizing the zinc-based seed coats deposited on seed surface and check whether impregnated Zn would pass through the seed coat and reach the interior of cotyledons; (2) establishing the ratio of Zn in the seed coat that is taken up by seedlings and transferred to soil; (3) assessing whether the seed coating affects seed germination and seedling growth, namely shoot height and root system architectural development; (4) probing the microchemical root environment in terms of Zn spatial distribution and availability. For that, we employed soybean (*Glycine max* (L.) Merrill) as a model species. The seed coating procedures adopted in this study mimicked those carried out in farms; the seeds were coated with ZnSO_4 and 40 nm or 5 μm ZnO at 4 g of

Zn per kg of seeds, which is a dose usually employed for this purpose [15].

Soybean (*Glycine max* (L.) Merrill) was chosen due to its economic importance, since it is a major protein source for livestock feeding and used in the production of edible oil and biodiesel [23]. In 2018, 341.7 million metric tons of soybean grains were produced worldwide [24].

2 Materials and methods

2.1 Materials

Three sources of Zn were used in the present study: $\text{ZnSO}_4 \cdot 7\text{H}_2\text{O}$ (Dinâmica®, Brazil), 5 μm ZnO (Agrichem Company, Brazil), and 40 nm ZnO (MK Impex Corp, Canada).

2.2 Characterization of materials

The particle size of 5 μm ZnO source was determined using a Laser Particle Sizer (ANALYSETTE 22 MicroTec Plus, Fritsch, Germany). An ultrasonic water bath suspended a wet dispersion of the material at 50 W and 40 kHz, exposed to two 7-mW laser beams with 532 nm and 940 nm wavelengths and the variations scattered light recorded by a focal plane detector to measure the particles size [25]. Figure S1 presents the volume distribution of the particle size of the measured samples. The 40 nm ZnO was previously fully characterized [26].

2.3 Seed treatment

To promote uniform covering of the seeds surface, emulating the procedures carried out by farmers and seed treatment industry [27–30], the seed treatment was performed as described below.

Eighty milligrams (weight Zn basis) of each Zn source were added in a plastic vial, and the volume made up to 1 mL with deionized water. Subsequently, the micrometric and nanometric oxide sources were sonicated using an ultrasonic processor (705 Sonic Dismembrator, Fisher Scientific, USA) operated at 70 W for three cycles of 15 min each.

Shortly after, each Zn treatment solution/suspensions were transferred to 200-mL polypropylene bags. Then, 20 g of soybean seeds (*Glycine max* (L.) Merrill) cultivar M7739IPRO were added and immediately mixed manually by ca. 1 min. Then, they were dried in a fume hood for 24 h at room temperature. It represented an exposure of 4 mg of Zn per g of seed. Deionized water was used as a control.

2.4 Scanning electron microscopy (SEM) analysis of the seed surface

Seeds were longitudinally sectioned to expose the hilum region. Then, the samples were immediately glued on carbon tape covering aluminum stubs, gold-coated (Balttec model SCD 050), and examined with a scanning electron microscope (JEOL-JSM IT 300) at 15 kV. Two seeds from each treatment were scanned.

2.5 Zinc distribution on coated seeds by X-ray fluorescence spectroscopy microanalysis (μ -XRF)

The soybean seeds were cross-sectioned, preserving the hilum region. The samples were placed on an X-ray sample cup (Chemplex no. 1530, USA) sealed with 5- μ m-thick polypropylene film (Spex SamplePrep, no. 3520, USA). The analysis was performed on a benchtop μ -XRF system (Orbis PC EDAX, USA) furnished with a Rh X-ray tube operated at 40 kV and 900 μ A and using a 25- μ m-thick Ti filter. Sixty-four points along a ca. 10 mm internal seed part were investigated by a 30- μ m X-ray beam focused by a polycapillary optical element. The X-ray spectra were acquired during 200 s using a 30 mm² silicon drift detector (SDD), with a dead time smaller than 3%. Two independent seeds per treatment were herein probed.

2.6 Zinc chemical speciation on coated seeds by X-ray absorption near-edge structure (XANES)

The Zn chemical environment of treated soybean seed coats was probed by X-ray absorption near edge structure (XANES) prior to sowing. The measurements were performed at the XRF beamline of the Brazilian Synchrotron Light Laboratory (LNLS), which was equipped with Si (111) double-crystal monochromator, KB mirror system providing a 20 μ m wide X-ray focused beam spot. The spectra were recorded in XRF geometry using a silicon drift detector (SDD; KETEK GmbH, Germany). The calibration was done employing a Zn foil. The reference compounds were obtained by pelletizing the pristine zinc sources in cellulose (0.2 Zn wt%).

The analysis was carried out in triplicate and acquired XANES spectra were merged to improve the signal-to-noise ratio. The spectra were merged, energy-calibrated using a Zn foil, and then, normalized using the Athena software within the IFEFFIT package.

2.7 Cultivation assay

After the treatments, the soybean seeds were individually sowed in 15-mL vials containing ca. 18 g sandy oxisol (sand-798; silt-50; clay-152 g kg⁻¹, respectively) in a growth room at 80.6 K (27 °C) and under 12-h photoperiod at 250 μ mol of photons m⁻² s⁻¹, provided by LED lamps (Golden, Brazil) and daily irrigation with deionized water. The soil was chosen due to its low-Zn content (0.8 g dm⁻³) in order to assess as the seed treatment a practical approach for Zn nutrition to soybean plants. The experiment was carried out using twenty-one replicates per treatment ($n=21$), divided into three groups of seven replicates each. The experiments were repeated twice in two moments of 2018 ($n=21$ in each treatment). These results are presented independently in the main manuscript and Supplementary Material.

After the germination, the seed coats were carefully removed and dried in a laboratory oven (515/4A, FANEM, Brazil) at 140 K (60 °C) over 48 h. Then, past 14 days, the plants were removed from the soil, the germination was recorded, and the root tissues were separated from the shoots using a steel blade, photographed for root and shoot length calculation, and then, dried at the same conditions of the seed coats. For the root and shoot heights and area calculation, the ImageJ software version 1.52a was employed.

2.8 Quantification of Zn by energy-dispersive X-ray fluorescence analysis (EDXRF)

The plant parts (root, shoot, seed coat) of twenty-one seedlings were grouped for producing the necessary sample mass for the analysis. The plant parts were then ground using a cryogenic grinder for 5 min pre-cooling and subsequently 5 grinding cycles of 2 min each with 1 min of cooling between cycles (Spex Sample Prep, Freezer/Mill 6870, USA). One hundred milligrams of the homogenized ground sample was added in the X-ray sample cup (Spex SamplePrep no. 3577, USA) sealed with 5- μ m-thick polypropylene film (Spex SamplePrep, no. 3520, USA). The Zn content was determined by EDXRF (Shimadzu EDX-720, Japan) using a cellulose-based external calibration curve (Fig. S2).

2.9 Probing the presence and availability of Zn on soil and soybean roots

The soybean seeds were individually sown in acrylic sample holders (rhizoboxes) using the same soil of the experiment described above. An external and removable lid was covered with a 5- μ m-thick polypropylene film (Spex SamplePrep, no. 3520, USA), and Fig. S3 showed

the experimental setup. The plants were grown in a growth room at 80.6 K (27 °C) and under 12-h photoperiod at 250 μmol of photons $\text{m}^{-2} \text{s}^{-1}$, provided by LED lamps (Golden, Brazil) and daily irrigated with deionized water.

After 14 days, the outer lids of the sample holders were removed, and the roots and their immediate neighboring soil were investigated by X-ray fluorescence microanalysis ($\mu\text{-XRF}$, Orbis PC EDAX, USA). The measurements were performed using a Rh X-ray tube operated at 40 kV and 900 μA and a 25- μm -thick Ti filter. Each spot was investigated during 120 s by a 30- μm X-ray beam focused by a polycapillary optical element. The X-ray spectra were acquired using a 30 mm^2 silicon drift detector (SDD), with a dead time smaller than 5%. The XRF spectra were fitted, and the Zn K α net counts were determined. Then, aiming at correcting differences in sample density, the Zn K α net counts were normalized by Compton Rh K α net counts. Only the Zn K α signals above the instrumental limit of detection (ILOD) were herein considered. The ILOD was calculated according to Eq. S1.

The experiment was carried out using three seedlings for the controls and the ZnSO_4 treatment, and two seedlings micro- and nanometric ZnO. Six soil and six root points were probed per seedling.

2.10 Quantification of zinc availability on soil through DTPA extraction

The soils of three independent soybean cultivation assays were air-dried and manually homogenized. Only the soils where soybean seeds germinate was herein considered. Then, 10 g of air-dried soil and 20 mL of the DTPA extracting solution (0.005 mol L^{-1} diethylenetriaminepentaacetic acid + 0.1 mol L^{-1} triethanolamine + 0.01 mol L^{-1} calcium chloride at pH 7.3) were stirred for 2 h. After filtration, the Zn content at the supernatant was determined by inductively coupled plasma optical emission spectrometry (ICP-OES iCAP 6200, Thermo Scientific) [31].

2.11 Statistical analysis

All quantitative data (i.e., seed germination, seedling tissue lengths, zinc probing on roots and soil, and the extractable zinc contents) were subjected to a one-way analysis of variance (ANOVA) plus Tukey tests at 95% confidence interval. Only the parameters analyzed which presented normal distribution were considered for ANOVA, as probed by the Anderson–Darling normality test, reported on figure labels. The procedures were carried out using Minitab® software version 18.1.

3 Results

3.1 ZnO nanoparticles can change seed cuticle surface

Soybean seeds display a rugulose cuticle pattern at the tegument epidermal cells surrounding the hilum (Fig. 1a, b). The seeds coated with ZnSO_4 exhibited a thick layer of residue that partially covered both tegument and hilum (Fig. 1c, d). The seeds coated with ZnO dispersions (Fig. 1e–h) presented two granulometric deposited patterns on the hilum and tegument (Fig. 1e–g). High-magnification images showed that 40 nm ZnO dispersion induced cuticle deformation (arrows in Fig. 1h), which was not observed for the other treatments.

3.2 Tracing Zn distribution in soybean seeds

Zinc XRF intensity at each point of the linescan (Fig. 2) is directly proportional to the Zn content. Thus, it allows assessing the Zn distribution along with sectioned soybean seeds. Regardless of the treatment, Zn presented the maximum count rate at the seed coats, and then, it sharply decreased at the seed coat–cotyledon interface (Fig. 2b–d). The content of Zn within the cotyledon was similar to the control, independently of the Zn source used in the coating process. These results show that Zn remained mainly trapped at the seed coat instead of being incorporated by the embryo. Figure S4 presents images recorded for biological replicates, which confirmed the trends described above.

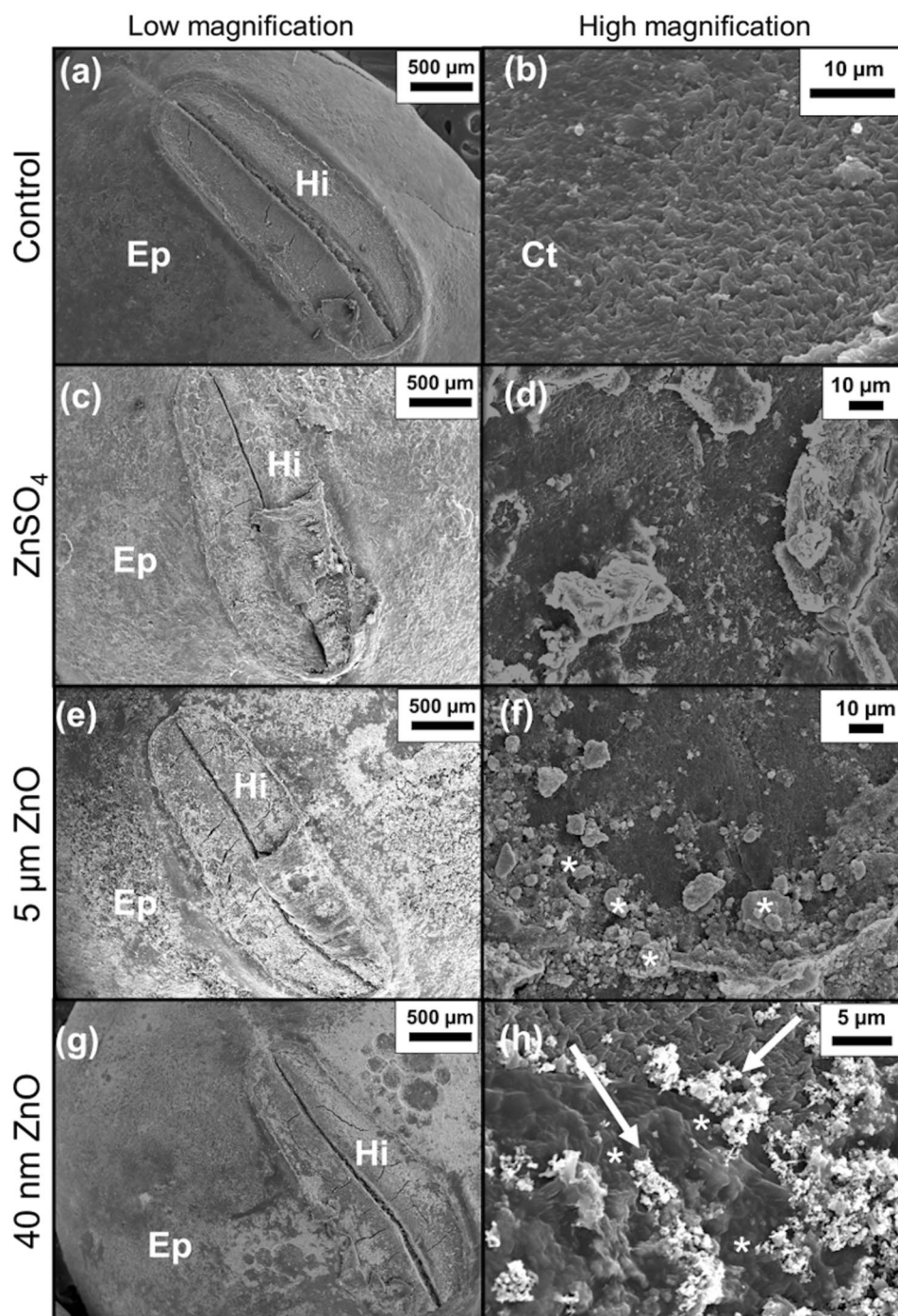
3.3 Chemical speciation of Zn coated on soybean seeds

Figure 3 presents normalized XANES spectra recorded at the hilum of coated and control soybean seeds. Additionally, it shows the spectra for pristine compounds used for seed coating (referred as standards). Since the spectral shape is a fingerprint of the sample chemical environment, it allows inferring whether Zn was biotransformed in the seeds. The control seed spectra clearly diverged from those acquired on coated seeds. However, the XANES spectra for ZnSO_4 and ZnO coated seeds were similar to their respective pristine compounds. Hence, it indicated that the major fraction of Zn was not biotransformed while adhered to the soybean seed coat.

3.4 Effects of seed coating on soybean germination and seedling development

Seedling development was affected by the seed coating with Zn sources. Figure 4a shows both nano- and micrometric ZnO, increased by ca. 40% of germinated

Fig. 1 Electron micrographs at low and high magnification of control soybean seed (a–b), and seeds coated with ZnSO_4 (c–d), $5\ \mu\text{m}$ ZnO (e–f), and $40\ \text{nm}$ ZnO (g–h). Note the rugulose cuticle morphology of the control sample (a–b) and the thick coverage caused by ZnSO_4 (c–d). The points presented with (*) indicates the presence of ZnO clusters (e–h). The arrows show the deformation of the cuticle. Ep—epidermis; Hi—hilum



seeds, comparing with the control group. The opposite was observed with the ZnSO_4 . Figure S5 presents data for an independent biological replicate where a similar pattern was found. Besides, the length of roots whose seeds were coated with ZnSO_4 was lower compared to those coated with ZnO and the control group (Fig. 4b). Albeit, the root length of plants whose seeds were exposed to nanometric ZnO was slightly shorter than those coated with micrometric ZnO . On the other hand,

the length of the shoots does not statistically differ as an effect of the seed treatments. Figure S6 presents pictures of the seedlings. The seed coating influenced the architecture of roots and shoots. Seeds coated with ZnO sources sprout seedlings with more developed roots and shoots, compared to the control. Conversely, the treatment with ZnSO_4 was detrimental to these parameters. These factors were quantified and are presented in Fig. S7—it shows root and shoots average mass gain per

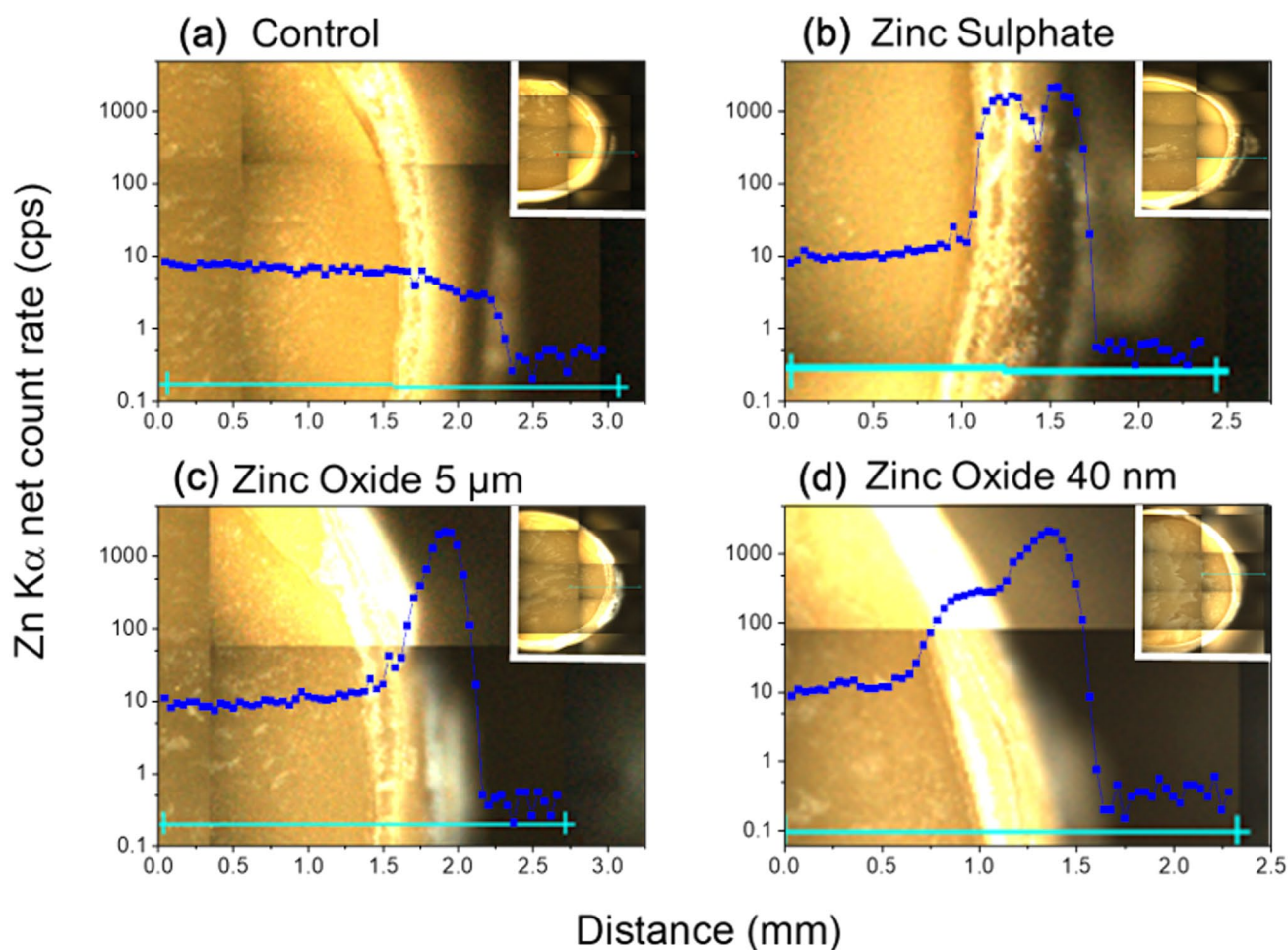


Fig. 2 XRF linescans showing the Zn distribution in soybean seeds coated with water (control) **(a)**, ZnSO_4 **(b)**, $5\ \mu\text{m}$ ZnO **(c)**, and $40\ \text{nm}$ ZnO **(d)** at $4\ \text{g}$ of Zn per kg of seeds. Regardless of the Zn source,

the number of counts was higher at the seed coat and hilum region. Co—cotyledons; Te—tegument

plant. Seedlings treated with ZnO performed better than control and those coated by ZnSO_4 .

3.5 Zinc mass balance

Table 1 shows the comparison between Zn content applied and found on soybean seeds and its respective losses during coating. The coating process yield, i.e., the fraction of Zn effectively transferred to the seed during the coating procedure, followed the order ZnSO_4 (89%) > nano ZnO (79%) > micrometric ZnO (48%). The fraction of Zn that was not transferred to the seed remained adsorbed on the walls of vials, pipette pointer, and the in the bag used in the processing.

Table 2 presents the Zn content balance in plants sprout from coated seeds. We determined the partitioning in roots, shoots, seed coats, and soil 8 days after sowing. Regardless of the source used to prime the seeds, the major fraction of Zn (> 80%) was found in soil, only ca. 15%

of the Zn was in tissues. It is worth noting that plants that received ZnSO_4 presented more Zn (12%) in shoot than those coated with nano- and micrometric ZnO (ca. 4%). On the other hand, Zn from oxide sources remained mostly in the seed coat than in living tissues. Table S1 presents the mass balance in milligrams (mg), and S2 shows that independent experiments found similar patterns.

3.6 Probing the presence and availability of Zn in soil

Figure 5a shows the experimental setup built to evaluate the presence of Zn in the rhizosphere of soybean plants whose seeds were coated with Zn sources. Plant roots and soil were probed in situ by $\mu\text{-XRF}$. Zinc was detected in most of the investigated root tissue and soil points (Fig. 5b). The Zn counts in the different probed points did not follow a normal distribution; hence, it was not possible to compare the average Zn counts using ANOVA. On the

Fig. 3 Zn K-edge XANES spectra recorded at the seed coat of control soybean seeds and seeds coated with ZnSO_4 , 40 nm ZnO and 5 μm ZnO at 4 g of Zn per kg of seeds. The control group was exposed only to deionized water. The blue circle indicates where the X-ray beam was probed. There is no evidence of Zn biotransformation on the seed surface

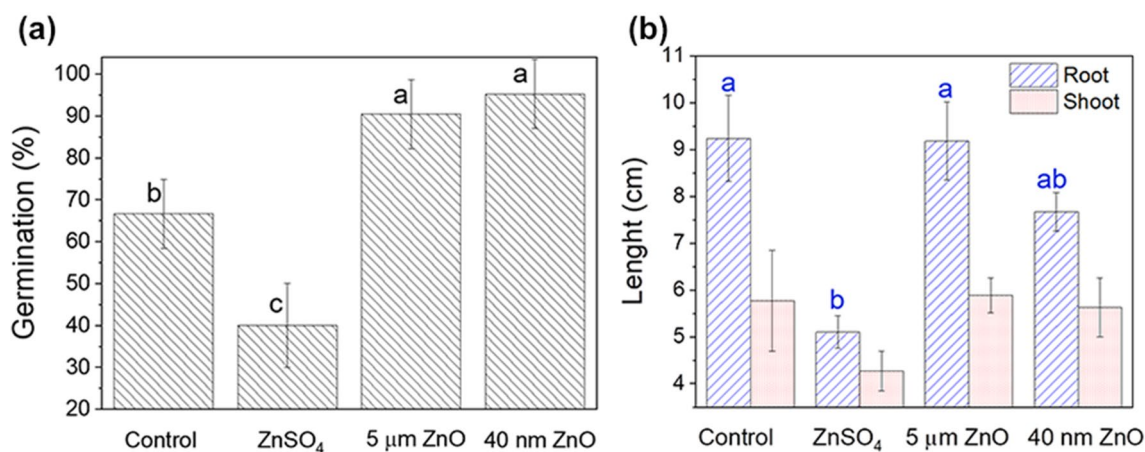
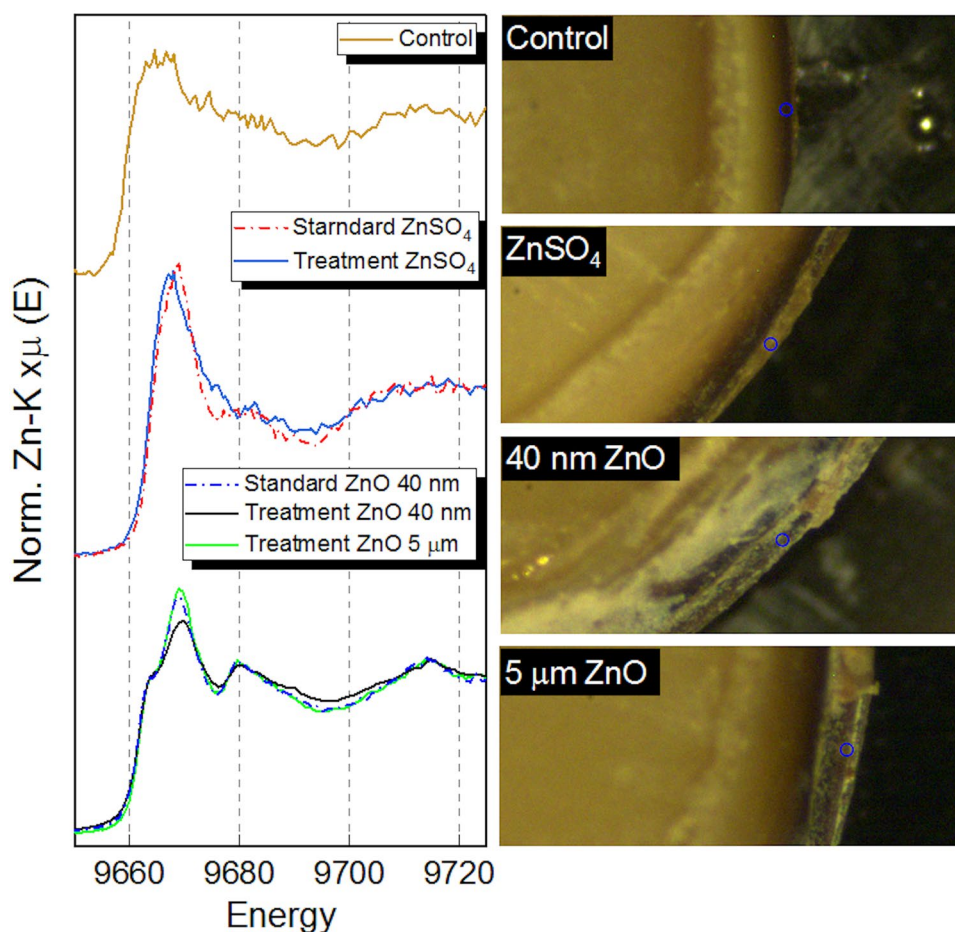


Fig. 4 Germination percentage (a) and root/shoot length (b) of soybean plants exposed to seeds coated with ZnSO_4 (c–d), 5 μm ZnO (e–f), and 40 nm ZnO (g–h) at 4 g of Zn per kg of seeds. The data were subjected to one-way ANOVA and Tukey post-compari-

son test with a 95% confidence interval. The degrees of freedom, p value, and F are **a** (11, <0.001, 25.38) and **b** (11, 0.04, 10.29—root; 11, 0.113, 2.75—shoot). Note that variations on soybean shoot length (b) does not present statistical significance

Table 1 Zinc losses during the seed coating with ZnSO_4 , 5 μm ZnO, and 40 nm ZnO at 4 g of Zn per kg of seeds

	Applied (%)	Recovered (%)	Process losses (%)
Control	–	–	–
ZnSO_4	100	89	11
5 μm ZnO	100	49	51
40 nm ZnO	100	80	20

The control group was exposed only to deionized water. The losses occurred due to the transference of zinc to the materials used in the process, i.e., bags, vials, and pointers. It represents the total zinc content of each tissue from twenty-one samples

Table 2 Zinc mass balance in the root, shoot, seed coat, and soil of germinated seeds coated with ZnSO_4 , 5 μm ZnO, and 40 nm ZnO at 4 g of Zn per kg of seeds

	Seeds (%)	Shoot (%)	Root (%)	Coat (%)	Soil (%)
Control	100	–	–	–	–
ZnSO_4	100	13	0	3	84
5 μm ZnO	100	4	–	10	85
40 nm ZnO	100	4	0	11	85

The control group was exposed only to deionized water. It represents the total zinc content of each tissue from twenty-one samples

other hand, the median number of counts for the probed points (Fig. 5b) was higher in plants and soil that received the treatments. Thus, the Zn applied via the seed coat was transferred to plant tissue and rhizosphere soil.

Figure 6 presents the availability of zinc on the soil by DTPA extraction 15 days after sowing. Regardless of the Zn source (despite the statistical differences between them), a minor fraction (4 to 13%) of the Zn remained available for plants.

4 Discussion

The chosen priming method was a proxy of the actual approach carried out by end-user farmers and in industrial seed treatment [27–30]. Such as in practical seed treatment, the procedure intends to transfer the coating to the seed surface rather than triggering seed germination during priming. Therefore, the amount of water and time of soaking was smaller and shorter, respectively, than those commonly reported in the literature [26, 32–34]. Additionally, an imbibed seed presents reduced mechanical resistance and is more prone to the attack of pathogens.

The μ -XRF linescans revealed that the seed coat prevented the Zn movement towards the cotyledon.

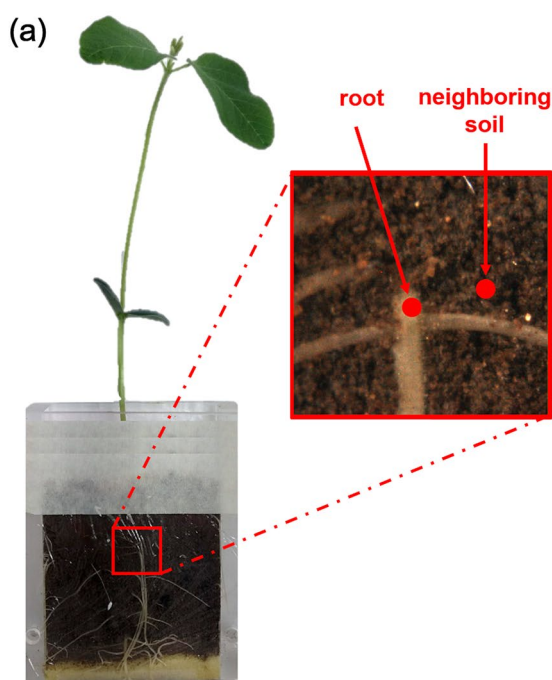
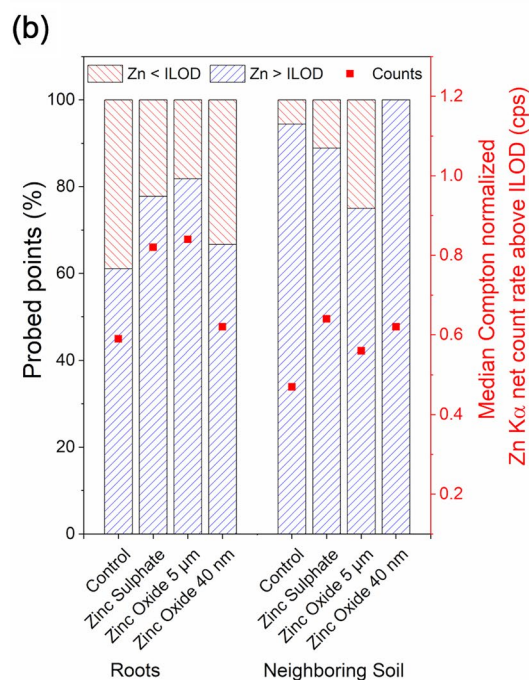


Fig. 5 Detailed view of rhizobox where XRF spectra were collected at soybean roots and neighboring soils (a). The presence or absence of Zn expressed as a percentage of the probed points ($n=18$ for the controls and ZnSO_4 , and $n=12$ for micro and nano),



and on the left axis is shown the median normalized Zn number of counts found above the ILOD while the amount expressed as cps is shown in the right axis (b). The amount of Zn was higher in the points probed in primed samples

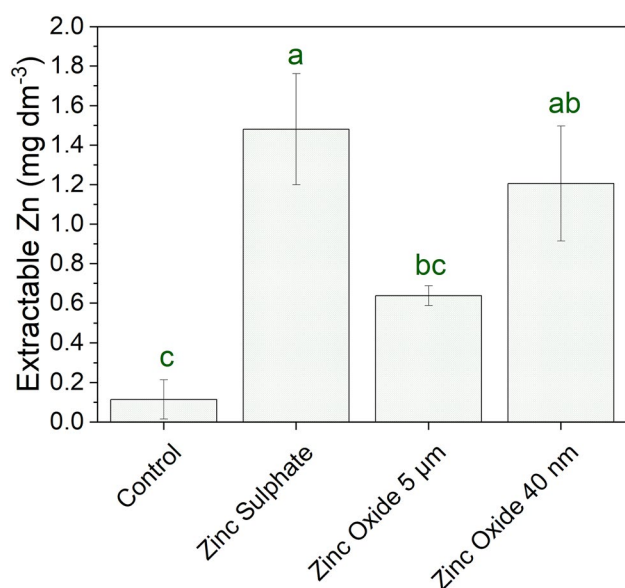


Fig. 6 DTPA extracted Zn in soils cultivated with primed ZnSO_4 , 5 μm ZnO, and 40 nm ZnO and control seeds. The data were subjected to one-way ANOVA plus a Turkey post-comparison test with a 95% confidence interval. The degrees of freedom, p -value, and F are 10, 0.001, and 18.54

According to the literature, the seed coat acts as a barrier for seed protection, due to its lignified tissues (e.g., sclerenchyma) [35], while the hilum is the porous tissue responsible for the imbibition processes [36]. Similar trends were found by μ -XRF maps of common bean (*Phaseolus vulgaris*) seeds soaked with CuSO_4 and 25 nm CuO at 1000 mg Cu L⁻¹; 40 nm CuO at 5000 mg Cu L⁻¹; 20 nm Fe_3O_4 and Fe_3O_4 at 1000 mg Fe L⁻¹; and ZnSO_4 and 20, 40, and 60 nm ZnO at 5000 mg L⁻¹. In all cases, the applied compounds remained on the seed coat, mainly concentrated in the hilum region [26, 33, 37]. These results are reinforced by the SEM images, where the coated ZnSO_4 formed a thick coverage on the soybean seed tegument, whereas for nano- and micro-ZnO dispersion, it was surrounding the hilum in the form of clusters.

Additionally, XANES revealed that Zn was not bio-transformed by the coating process. These results differ from those obtained for common beans (*Phaseolus vulgaris*), which showed biotransformation of the coated Cu, Fe, and Zn [26, 33, 37]. Furthermore, Cu XANES speciation in *Arabidopsis thaliana* seeds soaked in CuO nanoparticles at 20 and 50 mg L⁻¹ revealed Cu mainly as CuO, but also small fractions as $\text{Cu}_2(\text{OH})\text{PO}_4$, Cu acetate, and Cu_2O [38]. This contrast might be related to the time of soaking and water content employed in the coating process. Another possible explanation to the non-transformation relies on the soybean cuticle composition that

may interfere in the Zn absorption. Further studies are requested to clarify this topic.

The mass balance reveals that most of Zn at the coated seeds is transferred to the soil after sowing. Therefore, both ZnSO_4 and ZnO may modify the soil pH and concentration gradient microenvironment around the seeds [39]. Our study shows that ZnSO_4 resulted in lower germination percentage and constrained plant development. Similar results were found for rice (*Oryza sativa* L.) [40] and common bean (*Phaseolus vulgaris*) [26]. Higher doses of ZnSO_4 are reported to cause phytotoxicity due to salt accumulation on cell walls and induce several modifications to plant metabolism and growth. Also, phytotoxicity was found in radish (*Raphanus sativus*), rape (*Brassica napus*), ryegrass (*Lolium perenne*), lettuce (*Lactuca sativa*), corn (*Zea mays*), and cucumber (*Cucumis sativus*) seeds soaked in 35 nm Zn and 20 nm ZnO at 2000 mg L⁻¹ for 2 h, in which the half-maximal inhibitory concentration (IC_{50}) on seed germination and root growth was ca. 50 mg L⁻¹ for radish and roughly 20 mg L⁻¹ for rape and ryegrass [22]. On the other hand, our study showed that ZnO micro- and nanocoated seeds exhibited germination percentage and seedling development higher than those exposed to ZnSO_4 and the control group. Likewise, peanut (*Arachis hypogaea*) seeds soaked during 3 h in 25 nm ZnO at 1000 mg L⁻¹ presented a higher germination ratio, higher stem length, and root growth compared to those exposed to ZnSO_4 [19].

The mass balance showed that an important fraction of coated Zn remained adhered to the seed coat after germination. In addition, the Zn content in the shoots exposed to both ZnO sources is around four-fold lower than those exposed to ZnSO_4 . This result suggests that the solubility of Zn source is a factor limiting Zn uptake by plants. It was also reported Zn and Cu increase in maize (*Zea mays*) root and shoot after seed coating with CuCO_3 and ZnO at 0.14, 0.28, 0.56, 1.12, and 2.24 mg seed⁻¹ [29] and 0.55, 1.10, 2.20, 4.40, and 8.80 mg seed⁻¹ [41], respectively.

Conversely, although we found higher nano- and micro-metric ZnO source losses during the coating process compared to ZnSO_4 , the germination and plant development of the former seed treatments were higher than the latter one. Then, it is possible to assume herein that the agronomic efficiency of the process will not be directly proportional to the mass of coating material adhered to the seed.

Despite the increase in the total Zn content in soil due to the coating input, the latter contribution represents only around 5 μg of Zn per kg of soil. In this case, the Zn might remain in the rhizosphere. The μ -XRF intensities recorded at the roots and neighboring soil showed that the seed coating enriches the rhizosphere with Zn (Fig. 5).

We found higher DTPA extractable Zn fractions for soils that received seeds coated with ZnSO_4 and nanometric ZnO. It is known that charges in oxisols are highly

dependent on functional groups in organic matter and the surfaces of Fe, Al, and Mn (hydr)oxides [42]; such charges can immobilize Zn^{2+} . It has been previously reported that due to its higher surface-to volume ratio, nano-ZnO is rapidly transformed in soil [43], which may imply on its higher availability than micrometric ZnO source.

5 Conclusions

This study showed that the three zinc sources at 4 mg g^{-1} affected soybean seed coating germination, as well as soybean early-seedling development. Regardless of the source, the ZnO seed treatment boosted the germination by ca. 40%, compared to the control; it yielded longer roots with more developed architecture, compared to the control. The opposite was observed for ZnSO_4 , which clearly provided a toxic environment.

Counterintuitively, Zn seems not to cross the seed coat and enrich the cotyledons. It remained on the seed coat in their pristine chemical form. Besides, only a small fraction of the Zn applied to seeds was transferred to the seedling tissues. Most of it remained adhered on the seed coat or was transferred to the soil after sowing.

In addition to physiological pieces of evidence, Zn fertilization can be discussed thanks to in situ microprobe X-ray fluorescence. Although the Tukey test has not shown statistical difference from control, the average Zn was higher in roots and soil of rhizoboxes that received the treated seed. Hence, the Zn employed in the treatment is either transferred during the imbibition or it provides a fertile microenvironment that favors the initial seedling development. Perhaps, one of the leading advantages seed coatings consists in delivering the desired nutrient in the soil region where the roots will grow. In this context, ZnO treatments represent a better alternative since they release the Zn ions slower than the ZnSO_4 . Complementary studies comprising the plant life cycle are necessary to investigate the impact on productivity. This could be accomplished using Zn stable or radioactive isotopes.

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Author contributions GSM performed the seed treatment, XRF and XANES experiments and treated the data. ESR processed the XANES data, assisted the XRF experiments and discussion. JPRM performed and discussed the SEM analysis and manuscript writing.

EA participated in XRF quantitative analysis and manuscript writing. MC performed the DTPA extraction and contributed to the discussion of the results. GSM organized the content and structure of the manuscript and wrote the text. HWPC designed and coordinated the study, assisted the writing, and reviewed the whole paper.

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Compliance with ethical standards

Conflict of interest On behalf of all authors, the corresponding author states that there is no conflict of interest.

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