

1 **AQUACULTURE FACILITIES PROMOTE POPULATIONAL STABILITY**
2 **THROUGHOUT SEASONS AND INCREASE MEDUSAE SIZE FOR THE**
3 **INVASIVE JELLYFISH *Cassiopea andromeda***

4 Jorge Thé^{1*}; Hortência de Sousa Barroso¹; Marta Mammone²; Michael Viana¹; Caio
5 Servulo Batista Melo¹; Miguel Mies³; Thomás N. S. Banha³; André C. Morandini^{4,5};
6 Sergio Rossi^{1,2,6}; Marcelo de Oliveira Soares^{1,2,6}

7
8 ¹: Instituto de Ciências do Mar (LABOMAR), Universidade Federal do Ceará (UFC), Fortaleza,
9 Brazil

10 ²: DiSTeBA, Campus Ecotekne, University of Salento, 73100 Lecce, Italy

11 ³: Instituto Oceanográfico, Universidade de São Paulo, São Paulo, Brazil

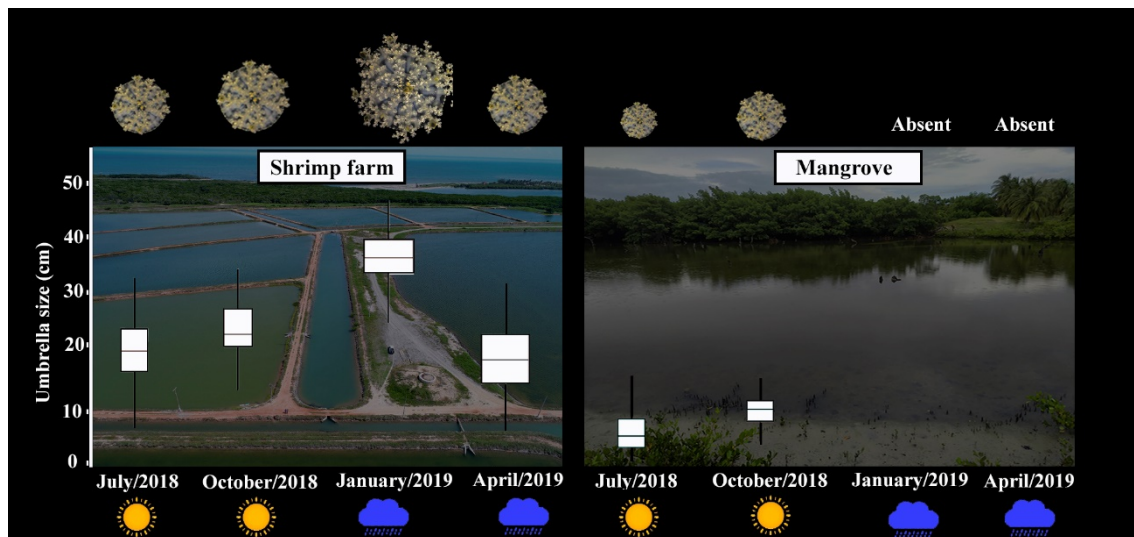
12 ⁴: Departamento de Zoologia, Instituto de Biociências, Universidade de São Paulo (USP), São
13 Paulo, Brazil

14 ⁵: Centro de Biologia Marinha, Universidade de São Paulo, São Sebastião, Brazil

15 ⁶: Institut de Ciència i Tecnologia Ambientals (ICTA), Universitat Autònoma de Barcelona
16 (UAB), Barcelona, Spain.

17 * corresponding author: jorgethe22@gmail.com

18 **GRAPHICAL ABSTRACT**



20 **ABSTRACT**

21 *Cassiopea* jellyfish have successfully invaded several marine ecosystems. We
22 investigated if *Cassiopea andromeda* grows larger (umbrella size) and if their
23 populations are more stable in shrimp farms than in mangroves in the Brazilian
24 coast. Our results show that jellyfish abundance is higher in the shrimp farm during the

25 rainy season and in the mangrove during dry season. The population is stable during
26 both seasons in the shrimp farm, but unstable in the mangroves, as jellyfish are absent
27 during rainy season. Shrimp farm-associated jellyfish are three times larger than those
28 in the mangroves, regardless of season. We recorded the largest (49.2 cm of umbrella
29 diameter) ever *C. andromeda* individual in the shrimp farm. Unlike the mangroves, the
30 shrimp farm provides environmental intra-annual stability that promotes jellyfish
31 growth and population persistence. Therefore, *C. andromeda* populations can be
32 seasonally dynamic and artificial environments such as aquaculture facilities may
33 facilitate the invasion process.

34 Keywords: estuary; mangrove; nutrients; Scyphozoa; shrimp farm; size distribution

35

36 1. INTRODUCTION

37 *Cassiopea* spp. (“upside-down jellyfish”) are mixotrophic, benthic animals that acquire
38 energy by both heterotrophic feeding, and photosynthate translocation performed by
39 their symbiotic dinoflagellates (Symbiodiniaceae) (Ohdera et al., 2018). Because of
40 their trophic plasticity, diversity of reproductive strategies (they display a metagenetic
41 life cycle, with polyps performing asexual reproduction and producing medusae, while
42 medusae reproduce sexually and generate polyps – see Morandini et al., 2016), and
43 resilience to physico-chemical variations, *Cassiopea* spp. have successfully invaded
44 many tropical and subtropical shallow ecosystems worldwide, such as mangroves,
45 seagrass beds and hypersaline estuaries (Stoner et al., 2011; Heins et al., 2015;
46 Morandini et al., 2017). Although there has been some confusion regarding the
47 identification of *Cassiopea* species in the past, it is thought that this genus first invaded

48 the Mediterranean through the Red Sea, before spreading to the Western Atlantic Ocean
49 (Graham and Bayha, 2008; Bayha and Graham, 2014; Morandini et al., 2017).

50 The increase in abundance and size of these invasive jellyfish is usually
51 observed in coastal environments under anthropogenic pressure (Stoner et al., 2011;
52 Zarnoch et al., 2020). Typical examples of artificial causes for intense jellyfish blooms
53 include climate change, eutrophication, overfishing, coastal construction, and
54 introduction of alien species (Purcell et al., 2007; Purcell, 2012; Richardson et al., 2009;
55 Stoner et al., 2016). This occurs because *Cassiopea* spp. populations are largely
56 influenced by intra-annual variations in environmental conditions, such as temperature,
57 salinity and nutrient concentration (Fitt and Costley, 1998; Stoner et al., 2011; 2014;
58 Freeman et al., 2016; Aljbour et al., 2017; 2019).

59 Temperature is a critical factor in *Cassiopea* spp. life cycle and invasion success
60 because it modulates several stages of development for both asexual and sexual
61 reproduction, including strobilation, larval settlement and metamorphosis (Fitt and
62 Costley, 1998; Hofmann et al., 2003; Newkirk et al., 2018). In addition, temperature
63 affects symbiont concentration in the jellyfish tissue because warmer conditions may
64 induce bleaching (McGill and Pomory, 2008; Lampert, 2016; Newkirk et al., 2018;
65 Klein et al., 2019). Salinity is also an important modulator of *Cassiopea* spp.
66 populations because it is a regulatory mechanism for asexual reproduction (Fitt and
67 Costley, 1998; Newkirk et al., 2018). In addition, prolonged exposure to lower salinities
68 may cause high mortality episodes (Klein et al., 2016a). Besides temperature and
69 salinity, variations in other environmental parameters may also play a relevant role in
70 the physiology and ecology of *Cassiopea* jellyfish. For instance, increase in nutrients,
71 chlorophyll-*a* (chl-*a*), and organic matter often enhance both heterotrophic feeding and
72 the photoautotrophy performed by their endosymbionts (Welsh et al., 2009; Stoner et

73 al., 2011; 2016; Lampert, 2016; Freeman et al., 2017; Ohdera et al., 2018; Djeghri et al.,
74 2020). Therefore, fluctuations in all of these parameters due to natural conditions and/or
75 human activities may stimulate either an increase or decrease in *Cassiopea* spp.
76 populations during the invasion process.

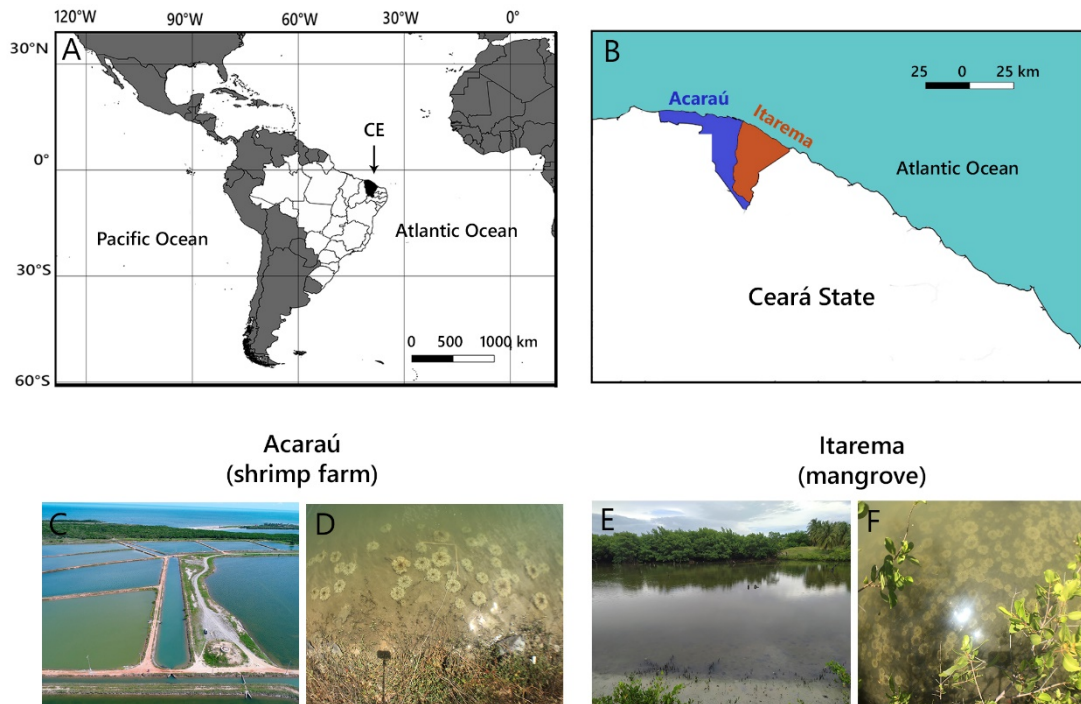
77 Urban development, agriculture, aquaculture activities, and their associated
78 consequences constitute major human pressures to estuaries and mangroves worldwide
79 (Lacerda, 2006; Suárez-Abelenda et al., 2014). These activities produce a large amount
80 of nitrogen and phosphorus, which leads to eutrophication events (Diaz et al., 2017;
81 Barcellos et al., 2019). Recently, it has been detected that the jellyfish *Cassiopea*
82 *andromeda* has invaded shrimp farms on the Brazilian semiarid coast, located in the
83 western Atlantic Ocean (Thé et al., in press). Shrimp farming is an economic activity of
84 great importance worldwide, that is often performed at or near mangroves (Sousa et al.,
85 2006; Nóbrega et al., 2013; Queiroz et al., 2013; FAO, 2018; Barcellos et al., 2019).
86 However, investigations comparing *Cassiopea* spp. non-indigenous populations in
87 aquaculture farms and nearby mangroves have not been conducted. The Brazilian
88 semiarid coast presents high and stable temperatures, oligotrophic waters, and low
89 rainfall rates (Barroso et al., 2018; Fernandez et al., 2019; Soares et al., 2019).
90 Therefore, it provides a unique opportunity to assess the intra-annual variations in size,
91 spatial distribution, abundance and seasonal frequency of the invasive *C. andromeda* in
92 both shrimp farms and mangroves. We investigated *C. andromeda* populations in the
93 semiarid mangrove and shrimp farm facilities to determine differences in medusae
94 umbrella size and populational stability, while also monitoring key environmental
95 conditions to determine the effects of seasonality for both habitats. In this way, *C.*
96 *andromeda* was investigated to determine if it displays larger umbrella size and if
97 populations are more stable in a shrimp farm than in the mangrove.

99 2. MATERIALS AND METHODS

100 2.1. Study area

101 Field surveys were performed in two adjacent habitats on the Brazilian semiarid coast
102 (Ceará state) in the western Atlantic Ocean (Fig. 1A-B): water channels inside a shrimp
103 farm in Acaraú (Fig. 1C-D, 2° 50' 9.00" S/40° 7' 13.30" W); and the mangroves at
104 Itarema (Fig. 1E-F, 2° 53' 2.83" S/39° 54' 42.17" W). Sites were approximately 20 km
105 apart. The shrimp farm has supply channels for pumping seawater from the ocean into
106 the farms. The semiarid mangrove is in a shallow (3-m deep) bar-built estuary. The
107 estuarine lagoon area between the coast and the barrier sandy beach is a protected area
108 of low hydrodynamics.

109 Climate in the area is tropical semiarid, marked by high sea surface temperature,
110 with low intra-annual variability (minimum of 26 °C and maximum of 30 °C), and little
111 rainfall (500–1000 mm.year⁻¹) (Soares et al., 2017). Ninety percent of the rainfall takes
112 place during rainy season in the summer and autumn. In the dry period, because of low
113 rainfall, high irradiance, and higher evaporation rates, a hypersaline condition (as high
114 as 38–63) is common in the shallow-water mangroves and surrounding estuaries
115 (Barroso et al., 2018; Valentim et al., 2018). Four intra-annual surveys were conducted
116 at each of the two habitats on July/2018 and October/2018 (dry season), and
117 January/2019 and April/2019 (rainy season), at a depth of 1–2 m.



118

119 Figure 1. The population structure of the invasive jellyfish *Cassiopea andromeda* was
 120 surveyed at two habitats on the Brazilian coast (Ceará state) (A-B) in the Western
 121 Atlantic Ocean: seawater channels inside a shrimp farm in Acaraú (C-D) and the
 122 adjacent semiarid mangroves at Itarema (E-F).

123

124 2.2. Data collection

125 2.2.1 Environmental parameters

126 A multiparametric probe (AK87, AKSO) was used to measure temperature, salinity and
 127 pH. For both habitats, measurements were performed once during each of the four
 128 surveys, at a depth of 0.5 m. Surface water samples were also collected for
 129 quantification of chlorophyll-a (chl-a) and nutrients. For chl-a analyses, samples were
 130 filtered in glass fiber filters of 0.7 μm (nominal pore size, Macherey-Nagel), and frozen
 131 at -20 $^{\circ}\text{C}$ (Parsons et al., 1984). Chlorophyll-a was extracted with 90% acetone (for 24
 132 hours, in the dark, at 4 $^{\circ}\text{C}$), and readings were performed on a spectrophotometer
 133 (wavelengths of 630, 647, 664, 665 and 750 nm). Absorbance values were converted to
 134 chl-a concentration, according to the equations of Jeffrey and Humphrey (1975).

135 Unfiltered water samples were used for quantification of total nitrogen (TN) and total
136 phosphorus (TP) according to the procedures of UNESCO (1983). Nutrient and chl-a
137 analyses were performed in quadruplicates.

138

139 **2.2.2. Population structure**

140 The unusual lifestyle of *C. andromeda* facilitates investigation and measurements using
141 photographs because, unlike other jellyfish species, they rarely swim and mostly remain
142 relaxed on the benthic surface. To investigate the individual size, abundance, and spatial
143 distribution of *C. andromeda* jellyfish populations in the shrimp farm and semiarid
144 mangrove, we performed three replicated 20-m transects within a single site for each
145 habitat. During each of the four surveys, 40 quadrats (0.5 x 0.5 m) were photographed
146 for each transect. The quadrats were equal (0.5 x 0.5m) with always the same size
147 throughout the sampling along the year. Image J was calibrated photo by photo using
148 the known quadrat sizes (0.5m x 0.5m).

149 *Cassiopea andromeda* individuals were classified as mature or immature based
150 on morphological characteristics and gonad presence (refer to Campbell, 1974 and
151 Schiariti et al., 2012). Ten individuals were collected during each of four surveys at
152 each of the two habitats. We then removed the oral disc and visually assessed the
153 gonadal tissue. Immature specimens displayed indistinguishable gonads, a small number
154 (less than 15) of appendages in the developing oral arms, and a small umbrella size (less
155 than 7.0 cm). Mature specimens presented brown gonads with visible gametes, a higher
156 number of oral appendages (> 15) and measured 7.0 cm or more in umbrella diameter.
157 Histological preparations to discern between different developmental stages of the
158 gonads were not made.

159

160 **2.3. Data sampling**

161 The presence of *C. andromeda* in the shrimp farm and semiarid mangrove was
162 quantified by analyzing the photoquadrats on ImageJ 1.51k software. The mean size of
163 the umbrella of *C. andromeda* specimens at both sites was also calculated with the use
164 of ImageJ 1.51k.

165 The spatial distribution of *C. andromeda* in the shrimp farm and mangrove was
166 also quantified using the Morisita index to determine if upside-down jellyfish display a
167 random, uniform or aggregated (patchy) distribution in both habitats (Morisita, 1959;
168 Amaral et al., 2014; Hayes and Castillo, 2017). The Morisita index ($I\delta$) was applied to
169 the populations in shrimp farm and mangroves assessed during each of the four surveys,
170 according to the formula (Equation 1).

$$171 \quad I\delta = n \times \Sigma X^2 - N/N(N-1), \quad (\text{Eq. 1})$$

172 where n: number of sample units; N: total number of individuals counted in all n sample
173 units; ΣX^2 : sum of the square of the number of individuals per sample unit. If dispersion
174 is random, $I\delta = 1.0$; if it is perfectly uniform, $I\delta = < 1.0$; and the aggregate pattern is
175 given by $I\delta > 1.0$, with the maximum aggregation being $I\delta = n$ (when all individuals are
176 found in 1 sample unit).

177

178 **2.4. Data and statistical analyses**

179 To determine if jellyfish umbrella size varies seasonally for mangrove versus shrimp
180 farms, as well as between seasons (January, April, July, and October), a two-way
181 ANOVA and post-hoc comparisons (Tukey's HSD) were conducted. All analyses were

182 conducted using R software (version 3.1.1), with the 'vegan' package (Oksanen et al.,
183 2016). To describe the spatial and temporal intra-annual variability of the abiotic factors,
184 and environmental variables (temperature, salinity, pH, chl-a, TN and TP), a principal
185 component analysis (PCA), and a cluster analysis were performed. The environmental
186 data were transformed and standardized for further use in the PCA analysis. The
187 grouping analysis was based on Euclidean distance, and complete linkage, with the data
188 previously transformed by $\log(x + 1)$. The statistical software PAST v. 3.20 (Hammer
189 et al., 2001) was used for PCA analysis, and the software PRIMER v. 6.0 (Clarke and
190 Gorley, 2006) was used for cluster analysis and dendrogram construction. The
191 significance level for all statistical analyses was considered at $p < 0.05$.

192

193 **3. RESULTS**

194 **3.1. Environmental data**

195 Salinity and pH values varied similarly in both habitats, with minimum values in the
196 rainy period (April/19) and maximum values in the dry period (October/2018) (Table
197 1). In the dry period (July and October), and January, waters were hypersaline, both in
198 the shrimp farm and semiarid mangroves. Temperatures reached slightly higher values
199 in the mangroves than in the shrimp farm (34 °C and 31.5 °C, respectively) (Table 1).
200 Nutrients (TN and TP) in the shrimp farm were unexpectedly lower than in the
201 mangroves (Table 1). The exception was the April/2019 (rainy season) survey, where
202 similar values of TP and TN were found in the shrimp farm and mangrove. A similar
203 intra-annual trend was also observed for chl-a (Table 1).

204

205

206

207 Table 1. Six environmental parameters (temperature, salinity, pH, total nitrogen-TN,
 208 total phosphorus-TP and chlorophyll-*a*) were evaluated in water channels inside a
 209 shrimp farm, and on semiarid mangroves containing invasive *C. andromeda* jellyfish
 210 populations. Both habitats are located on the tropical Brazilian coast in the Western
 211 Atlantic Ocean. The parameters were evaluated during dry (July 2018 and October
 212 2018) and rainy (January 2019 and April 2019) seasons.

Site	Month /Year	C°	Salinity	Chlorophyll- <i>a</i> ($\mu\text{g L}^{-1}$)	pH	TN (μM)	TP (μM)
Shrimp farm	Jul/2018	27.8°	39.9	1.70	7.9	18.38 $\pm 1,73$	1,05 \pm 0,043 213 214
	Oct/2018	31.5°	46.2	0.76	8.4	13.30 $\pm 0,94$	0,31 \pm 0,046 215 217
	Jan/2019	31.1°	43.2	1.46	8.2	15.70 $\pm 0,30$	0,31 \pm 0,046 218
	Apr/2019*	28.6°	31.6	13.2	8.0	42.78 $\pm 0,61$	1,81 \pm 0,112 219 220
Mangrove	Jul/2018	31.7°	38.9	5.08	8.0	52.84 $\pm 3,71$	2,76 \pm 0,094 221 222
	Oct/2018	31.8°	46.9	3.26	7.9	42.28 $\pm 2,29$	1,87 \pm 0,046 223 224
	Jan/2019	34°	44.5	2.18	8.3	64.70 $\pm 0,80$	2,13 \pm 0,230 225
	Apr/2019*	30.4°	24.4	2.45	8.1	42.06 $\pm 0,84$	1,79 \pm 0,032 226 227

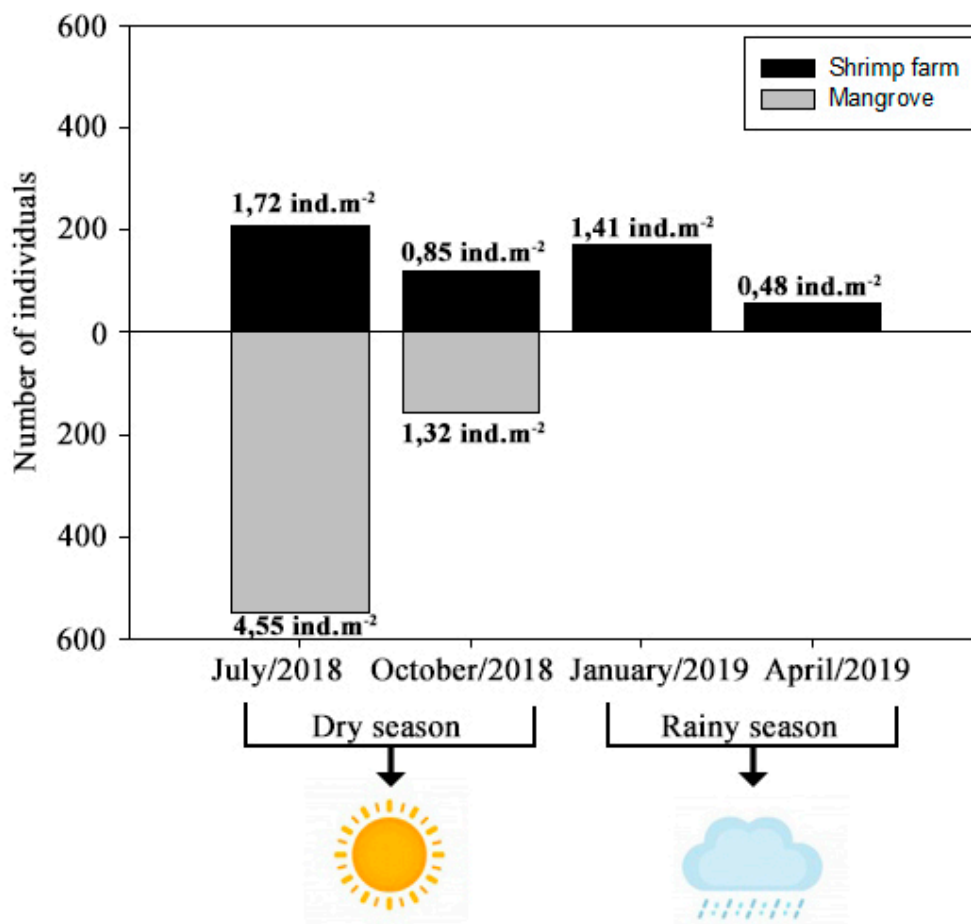
228

229 * Most intense rainy period in the last 11 years old (FUNCEME 2019).

230

231 3.2. Population structure

232 A greater number of jellyfish was found in the mangroves during dry season (Fig. 2). A
 233 total of 546 individuals were counted in the mangroves on July/2018, and 158 for
 234 October/2018; 207 and 119 were counted for the shrimp farm on those dates,
 235 respectively. However, zero jellyfish were detected in the semiarid mangroves during
 236 the rainy season, while the shrimp farm recorded 170 and 58 individuals for
 237 January/2019 and April/2019, respectively (Fig. 2) (XXX). For both habitats,
 238 populations display an aggregated spatial distribution pattern (Table 2).



239

240 **Figure 2.** Number of individuals and population density (individuals per m²) of the
 241 invasive jellyfish *Cassiopea andromeda* evaluated during the dry and rainy seasons in a
 242 shrimp farm and mangroves, both located on the tropical Brazilian coast (Western
 243 Atlantic Ocean).

244

245

246

247 Table 2. The Morisita Index ($I\delta$) was applied to investigate whether *Cassiopea*
 248 *andromeda* populations display a random ($I\delta = 1.0$), uniform ($I\delta < 1.0$) or aggregated
 249 ($I\delta > 1.0$) distribution pattern in a shrimp farm and in a semiarid mangrove, both located
 250 in the tropical Brazilian coast (Western Atlantic Ocean). (-): no jellyfish detected.

Sites	Month/Year	$I\delta$	Distribution
Shrimp farm	July/2018	3.6	Aggregated
	October/2018	1.71	Aggregated
	January/2019	1.98	Aggregated
	April/2019	2.73	Aggregated
Mangrove	July/2018	1.22	Aggregated
	October/2018	2.04	Aggregated
	January/2019	-	-
	April/2019	-	-

251

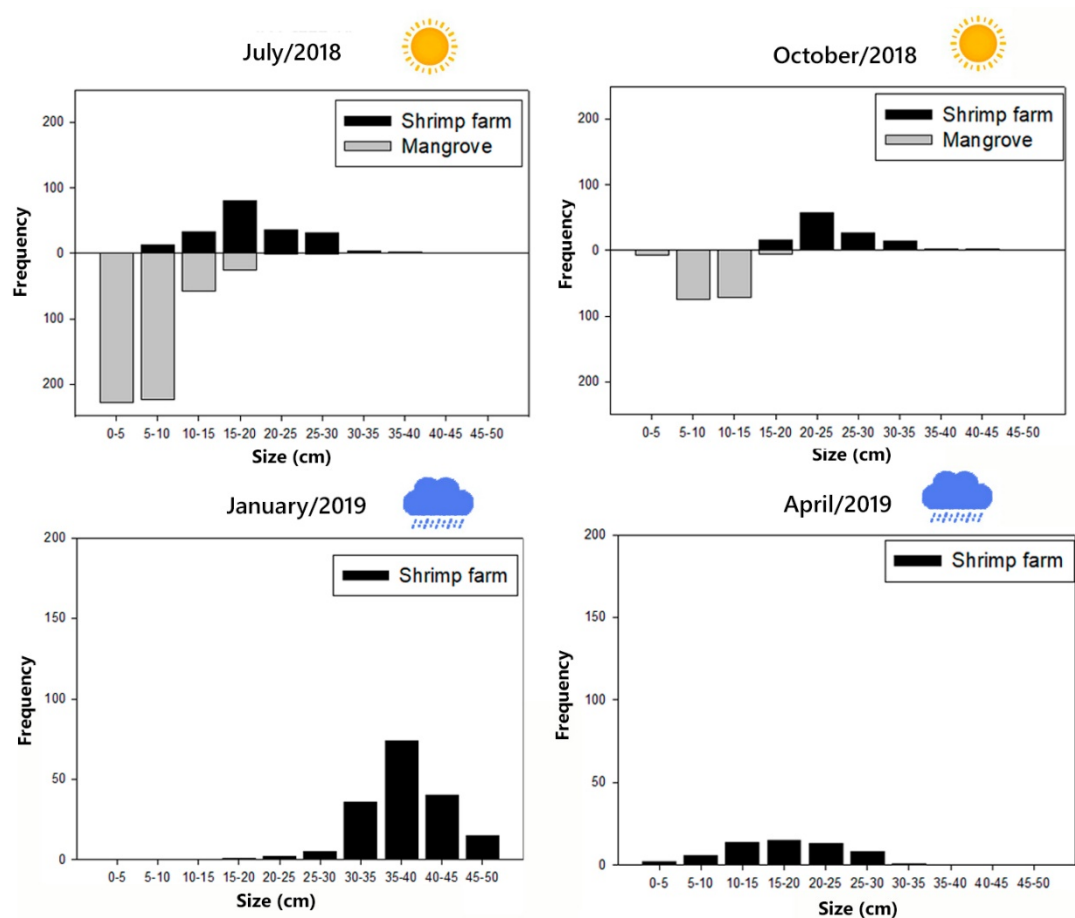
252

253 *Cassiopea andromeda* umbrella size was 8.2 ± 3.4 cm in the mangroves, and
 254 24.7 ± 5.8 cm in the shrimp farm (Figs. 3 and 4). The results of Two-Way ANOVA on
 255 umbrella size showed significant differences between the mangrove and shrimp farm
 256 ($df=1$; $F=4679.49$; $p < 0.0016$), as well as between the months ($df=3$; $F=517.68$; $p <$
 257 0.0016), as well as identified significant interactions between these two factors ($df=1$;
 258 $F=10.12$; $p < 0.0015$). The Tukey multiple comparisons of means between the months
 259 (January, April, July, October) showed significant differences in all comparisons; the
 260 only exception is between April and July in the shrimp farm when the umbrella
 261 diameter showed similar mean sizes ($F=4.08$; $p=0.1239$) (Figura 4). The smallest size
 262 class (0–7 cm) was recorded only in the dry period, and for the mangrove only (Fig. 3).
 263 This pattern explains the abundance of small-sized jellyfish (Fig. 3) in this estuarine

264 **habitat** during the dry period (Fig. 2). Smaller jellyfish were present during dry and
265 rainy seasons at the shrimp farm. Larger jellyfish were predominant during both seasons
266 at the shrimp farm (Fig. 3). Adult jellyfish were only seen in the semiarid mangroves in
267 the dry period (Figs. 3 and 4).

268

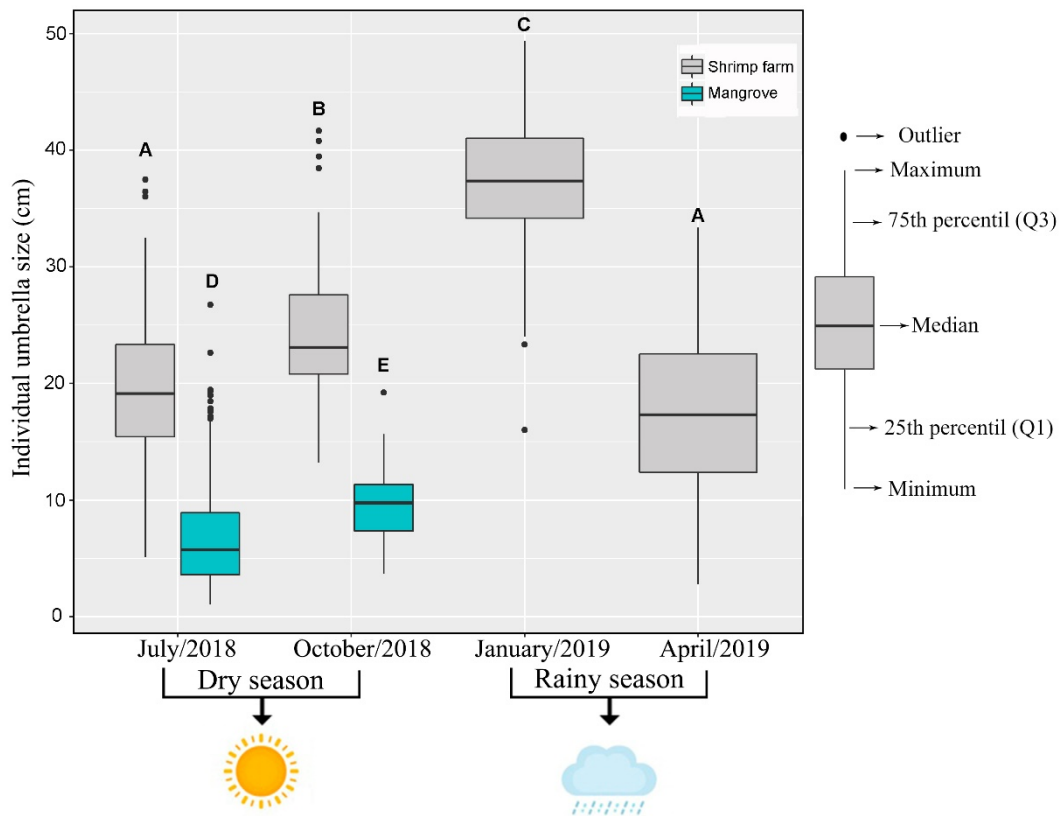
269



270

271 **Figure 3.** Size class frequency of the umbrella diameter (cm) of the invasive jellyfish
272 *Cassiopea andromeda* in a shrimp farm and in semiarid mangroves located on the
273 tropical Brazilian coast in the Western Atlantic Ocean. Data were produced during both
274 dry and rainy seasons.

275



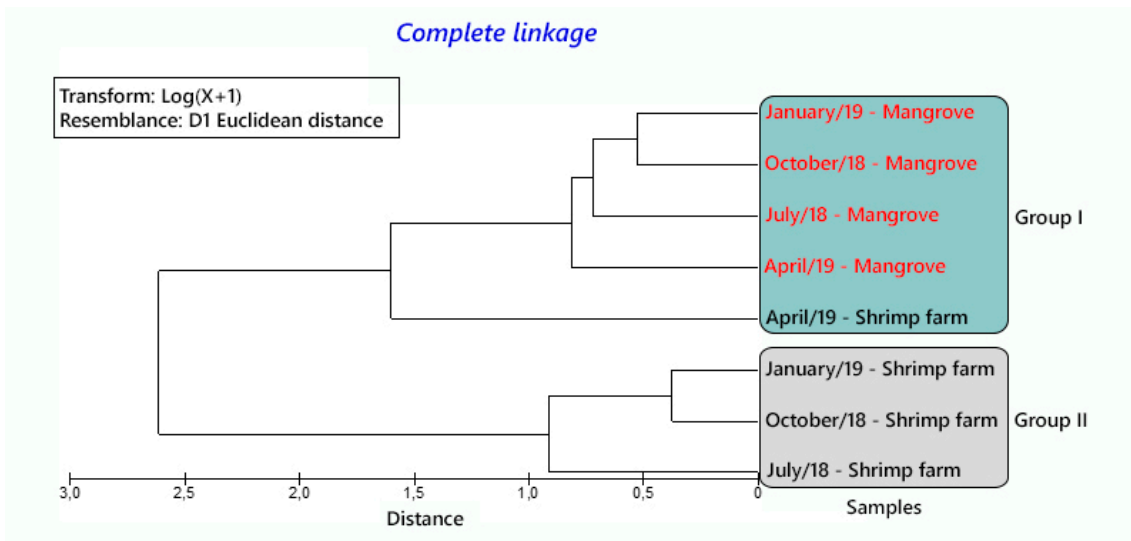
276

277 **Figure 4.** Seasonal variation in the umbrella size of the invasive jellyfish *Cassiopea*
 278 *andromeda* in a shrimp farm and in semiarid mangroves located on the tropical
 279 Brazilian coast in the Western Atlantic Ocean. Data were produced during dry
 280 (July/2018 and October/2018) and rainy seasons (January/2019 and April/2019).
 281 Different letters above the box plots indicate statistically different groups assessed with
 282 the use of a two-way analysis of variance and Tukey's multiple comparison test.

283

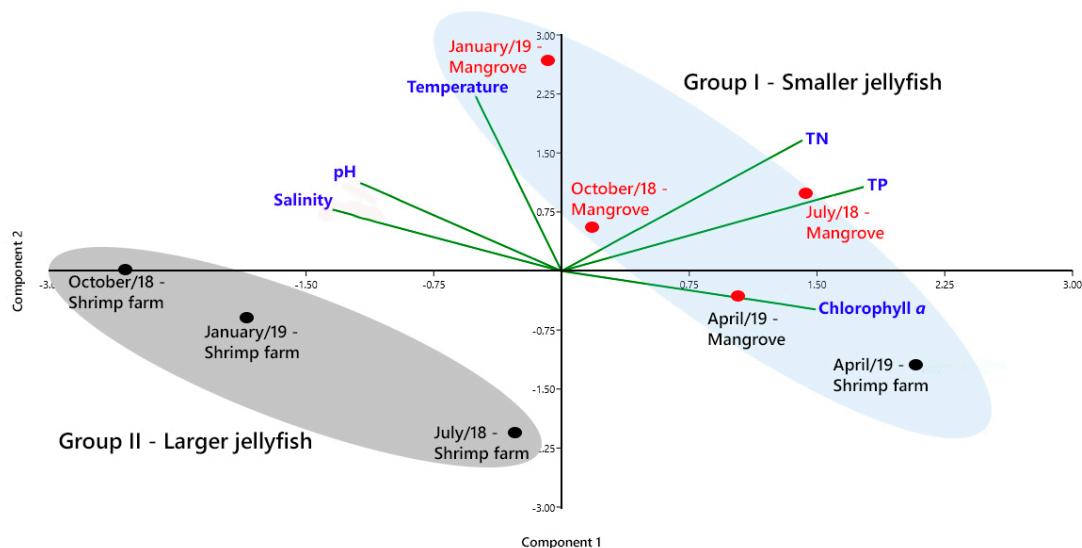
284 Cluster analysis and PCA revealed two distinct groups (Figs. 5 and 6). In the
 285 PCA, the first two axes explained most of the variation in the environmental data (76%)
 286 (Fig. 5). Axis 1 (42%) was negatively correlated with temperature ($r = -0.15$), pH ($r = -$
 287 0.35), and salinity ($r = -0.41$), and positively correlated with TN ($r = 0.42$), TP ($r =$
 288 0.53) and chl-a ($r = 0.45$). On the other hand, axis 2 (34%) was negatively correlated
 289 with chl-a ($r = -0.14$), and positively correlated with temperature ($r = 0.67$), salinity ($r =$
 290 0.23), TN ($r = 0.50$), TP ($r = 0.32$) and pH ($r = 0.33$). Group I was characterized by
 291 higher values of TN, TP and chl-a, and generally smaller jellyfish. Group II presented

292 the exact opposite trend, and was composed of shrimp farm samples only (Figs. 5 and
293 6).



294
295

296 **Figure 5.** Cluster analysis performed on six environmental parameters (temperature,
297 salinity, pH, total nitrogen, total phosphorus and chlorophyll-*a*) measured at a shrimp
298 farm and adjacent semiarid mangroves containing invasive *Cassiopea andromeda*
299 populations. Both sites were located on the tropical Brazilian coast in the Western
300 Atlantic Ocean.



301

302 **Figure 6.** Principal component analysis (PCA) analysis of spatial and temporal variation
303 in environmental parameters (temperature, salinity, pH, total nitrogen-TN, total
304 phosphorus-TP and chlorophyll-*a*), and the umbrella size of the invasive jellyfish
305 *Cassiopea andromeda* in a shrimp farm and semiarid mangroves located on the tropical
306 Brazilian coast (Western Atlantic Ocean).

307

308 4. DISCUSSION

309 This work investigated if the invasive jellyfish *C. andromeda* is more adapted to
310 the artificial shrimp farm environment than their typical mangrove habitat by assessing
311 if they present more stable populations and grow to larger umbrella sizes. Our findings
312 show that *C. andromeda* populations are similarly abundant throughout rainy and dry
313 seasons in the shrimp farm. For the mangroves, however, while populations are larger
314 during dry season, they become absent during rainy season. Therefore, shrimp farm
315 populations are relatively stable, while mangrove populations are seasonal and
316 dominated by smaller jellyfish. As for size, shrimp farm jellyfish grew three times
317 larger than those in the Brazilian semiarid mangroves, regardless of season.

318 The main reason for the population stability in the Brazilian shrimp farm is the
319 stability of the physico-chemical environment. Mangroves are relatively unstable
320 ecosystems (Hogarth, 2015), while shrimp farms are usually kept under controlled and
321 stable conditions in order to optimize shrimp production (Yu et al., 2006). The semiarid
322 mangroves were associated with a higher fluctuation in salinity, and reached a low value
323 of 24.4 during the rainy season, which is much lower than the optimum range of 35–45
324 for *Cassiopea* jellyfish (Kristensen and Ypma, 1971; Klein et al., 2016b). This
325 coincided with the disappearance of the jellyfishes in the mangrove, and with the high
326 volume of rainfall (546 mm month⁻¹), which was the highest for the area in the past
327 decade (FUNCEME, 2019). The rain could have transported the jellyfish in the
328 mangrove downstream, but the study area is a bar-built and closed estuarine ecosystem
329 with no connection to rivers. Therefore, transport of jellyfish was prevented and this
330 very likely caused relevant mortality associated with low salinity. However, *Cassiopea*
331 polyps are more resistant and likely to survive, which could explain repopulation of
332 medusae in the dry season.

333 The hypersaline condition (32-47) in the mangroves during dry season and in the
334 shrimp farm during both seasons was likely favorable for population maintenance.
335 Temperature also plays a major part, as our results show that it was lower in the shrimp
336 farm, not exceeding 32 °C, and usually stable with low intra-annual variation (28.6 to
337 31.5 °C). This temperature range is considered ideal for the growth and development of
338 *Cassiopea* spp. (Fitt and Costley, 1998; Klein et al., 2016; Newkirk et al., 2018; Aljbour
339 et al., 2019). However, during the rainy season in the mangrove, temperature reached
340 values higher than 34°C which is the threshold for bleaching and thermal stress in the
341 jellyfish (McGill and Pomory, 2008). Higher temperatures such as these have been
342 found to increase jellyfish mortality rates (Fitt and Costley, 1998; Klein et al., 2016b;
343 Newkirk et al., 2018; Aljbour et al., 2019).

344 The populations of *C. andromeda* in the shrimp farm and semiarid mangroves
345 are composed only of females (Thé et al., in press). This suggests that they have a low
346 genetic variability (Morandini et al. 2017) that is associated with a higher susceptibility
347 to negative impacts provoked by environmental instability (Le Roux and Wieczorek,
348 2008). A similar case was reported for *C. andromeda* in Cabo Frio (subtropical Brazil),
349 which had a population composed only of males that disappeared after changes in
350 temperature and salinity (Morandini et al., 2017). Therefore, the mangrove physico-
351 chemical instability associated with the synergistic effects of salinity reduction and
352 temperature increase during rainy season may have contributed to the disappearance of
353 jellyfish. The aggregated distribution and low mobility of *C. andromeda* emphasizes the
354 importance of abiotic parameters, which are often key drivers for this type of
355 disappearance, and spatial distribution pattern (Legendre and Fortin 1989).

356 Our findings also show that *C. andromeda* grows to larger sizes in the shrimp
357 farm throughout all seasons. This can be attributed to environmental stability and

358 organic matter input. Nutrient and chl-a concentration were, unexpectedly, higher in the
359 mangrove than at the shrimp farm. A possible explanation for higher growth in less
360 eutrophic conditions is that nutrient concentrations in the semiarid mangroves may have
361 been excessive. Total phosphorus (TP) in the shrimp farm and mangroves ranged from
362 0.3–1.8 and 1.8–2.8 μM , respectively. Stoner et al. (2011) suggested that the highest
363 density of *Cassiopea* spp. is found when TP is lower than 1.5 μM , and there is evidence
364 that nutrient intake is reduced when TP exceeds 2.0 μM (Todd et al., 2006).

365 It is likely that organic matter availability is likely a much stronger predictor for
366 jellyfish fitness and growth than nutrients because *Cassiopea* spp. are efficient
367 osmotrophs (McGill and Pomory, 2008; Welsh et al., 2009). Brazilian shrimp farms are
368 notoriously intense producers of dissolved and particulate organic matter (Sousa et al.,
369 2006; Venekey and Melo, 2016), which would promote a constant, aseasonal, and
370 intense heterotrophic behavior in *Cassiopea* spp. The importance of heterotrophy
371 (predation and/or osmotrophy) has been widely shown for several cnidarians, including
372 reef-building corals (Grottoli et al., 2006; Houlbrèque and Ferrier-Pagès, 2009; Mies et
373 al., 2018), pelagic jellyfish (McCloskey et al., 1994; Purcell et al., 2007; 2012), and also
374 *Cassiopea* spp. (Pierce et al., 2005; McGill and Pomory, 2008; Banha et al., submitted).
375 While the photoautotrophy performed by the symbionts may provide the host with a
376 significant amount of carbon (Muscatine and Porter, 1977), photosynthetic products
377 present a typically low nitrogen and phosphorus content, which are essential for protein
378 synthesis (Battey and Patton, 1986; Ferrier-Pagès et al., 2003). Therefore, besides
379 physicochemical stability, heterotrophic feeding may explain why the *C. andromeda* in
380 the shrimp farm are among the largest ever recorded at 49 cm (of umbrella), unlike their
381 typical size of less than 15 cm worldwide (Niggl and Wild, 2010; Stoner et al., 2011,
382 2014a,b; Zarnoch et al., 2020). However, a more robust experimental approach,

383 possibly using stable isotopes or fatty acid markers, is required for confirming the
384 relevance of heterotrophic input.

385 Research on upside-down jellyfish (*Cassiopea* spp.) has increased in recent
386 years because it is considered a model organism for invasion, symbiosis, and climate
387 change investigations (Odhera et al., 2018; Djeghri et al., 2019). Within these contexts,
388 the major contribution from our findings is showing that nearshore aquaculture facilities
389 provide stable conditions that stimulate *Cassiopea* growth at population and individual
390 levels, thus aiding the invasion process. In addition, we present baseline information on
391 the seasonal dynamics of *Cassiopea* populations, and also highlight that *Cassiopea* may
392 also serve as a relevant model organism for trophic investigations. Nonetheless, given
393 the inherent variability in jellyfish populations, more intensive and long-term ecological
394 research programs are required to understand the spatial and temporal dynamics of the
395 invasion processes.

396

397 5. ACKNOWLEDGEMENTS

398 JT thanks CAPES MSc. scholarship. ACM was supported by FAPESP (2015/21007-9) and
399 CNPq (309440-2019-0). This is a contribution of NP-BioMar USP. MOS thanks CNPq
400 Research Productivity Fellowship, INCT Ambtropic, CAPES-PRINT Program and FUNCAP
401 (Chief-Scientist Program). This paper from our research group (number 4) celebrates the
402 beginning of the United Nations (UN) Decade of the Ocean Science for Sustainable
403 Development (2021–2030). We hope that this decade will provide a ‘once in a lifetime’ global
404 opportunity to create a new science-based foundation to society, across the science-policy
405 interface, to strengthen the management of our oceans and coasts for the benefit of humankind
406 and all marine species.

407

408

409 6. REFERENCES

410 Aljbour SM, Zimmer M, Kunzmann A (2017) Cellular respiration, oxygen consumption, and
411 trade-offs of the jellyfish *Cassiopea* sp. in response to temperature change. *Journal of Sea*
412 *Researc* 128, 92–97. Doi: [dx.doi.org/10.1016/j.seares.2017.08.006](https://doi.org/10.1016/j.seares.2017.08.006)

413
414 Aljbour SM, Zimmer M, Al-Horani FA, Kunzmann A (2019) Metabolic and oxidative stress
415 responses of the jellyfish *Cassiopea* sp. to changes in seawater temperature. *Journal of Sea*
416 *Researc* 145, 1–7. Doi: doi.org/10.1016/j.seares.2018.12.002

417
418 Amaral MK, Netto OS, Lingnau C, Figueiredo AF (2014) Evaluation of the Morisita index for
419 determination of the spatial distribution of species in a fragment of araucaria forest. *Applied*
420 *Ecology and Environmental Research* 13, 361-372. Doi: [10.15666/aeer/1302_361372](https://doi.org/10.15666/aeer/1302_361372)

421
422 Banha TNS, Mies M, Güth AZ, Pomory CM, Sumida PYG (2020) Juvenile *Cassiopea*
423 *andromeda* (Cnidaria: Scyphozoa) medusae are resistant to multiple thermal stress events and
424 rely more on heterotrophy than on the symbiotic relationship with Symbiodiniaceae for growth.
425 Submitted

426 Barcellos D, Queiroz HM, Nóbrega GN, Filho RLO, Santaella ST, Otero XL, Ferreira TO
427 (2019)
428 Phosphorus enriched effluents increase eutrophication risks for mangrove systems in
429 northeastern Brazil. *Marine Pollution Bulletin* 142, 58–63. Doi:
430 doi.org/10.1016/j.marpolbul.2019.03.031

431
432 Barroso HS, Tavares TCL, Soares MO, Garcia TM, Rozendo B, Vieira ASC, Viana B, Pontes
433 TM, Ferreira TJ, Filho JP, Schettinie CAF, Santaella ST (2018) Intra-annual variability of
434 phytoplankton biomass and nutrients in a tropical estuary during a severe drought *Estuarine,*
435 *Coastal and Shelf Science* 213, 283–293. Doi: [org/10.1016/j.ecss.2018.08.023](https://doi.org/10.1016/j.ecss.2018.08.023)

436
437 Battey J., Patton JS (1986) Glycerol translocation in *Condylactis gigantea*. *Marine Biology*,
95:37–46. Doi: [10.1007 / BF00447483](https://doi.org/10.1007/BF00447483)

438 Bayha KM, Graham WM (2014) Nonindigenous Marine Jellyfish: Invasiveness, Invasibility,
439 and Impacts. In: K. A. Pitt & C. H. Lucas (Eds), *Jellyfish Blooms*. Springer Netherlands,
440 Dordrecht, pp. 45-77.

441
442 Campbell RD (1974) Cnidaria. In: A. C. Giese, J.S. Pearse (Ed), *Reproduction of marine invertebrates,*
443 *Volume I: Acoelomate and pseudocoelomate metazoans*. Academic Press, pp. 133-199.

444
445 Clarke KR, Gorley RN (2006) PRIMER v6: User Manual/Tutorial. PRIMER-E Ltd., Plymouth
446 Marine Laboratory, Plymouth

447
448 Diaz R, Selman M, Chique-Canache C (2017) Global eutrophic and hypoxic coastal systems:
449 eutrophication and hypoxia–nutrient pollution in coastal waters. In: WRI Policy Note. World
450 Resources Institute.

451
452 Djeghri N, Pondaven P, Stibor H, Dawson MN (2019) Review of the diversity, traits, and
453 ecology of zooxanthellate jellyfishes. *Marine Biology* 166, 1-19. Doi:
454 <https://doi.org/10.1007/s00227-019-3581-6>

455
456 Djeghri N, Stiborb H, Lebeauc O, Pondaven P (2020) $\delta^{13}\text{C}$, $\delta^{15}\text{N}$, and C:N ratios as nutrition
457 indicators of zooxanthellate jellyfishes: insights from an experimental approach. *Journal of*
458 *Experimental Marine Biology and Ecology* 522, 151257. Doi:
459 <https://doi.org/10.1016/j.jembe.2019.151257>

460

- 461 FAO (2018) The state of world fisheries and aquaculture meeting the sustainable development
462 goals. FAO, Rome.
463
- 464 Fernandez GB, Rocha TB, Barboza EG, Dillenburg SR, Camara Rosa MLC, Angulo RJ, Souza
465 MC, Oliveira LHS, Dominguez JML (2019) Natural Landscapes Along the Brazilian Coastline.
466 In The Physical Geography of Brazil, Chapter 10, 199-218. Springer, Cham.
- 467 Ferrier-Pagès C, Witting J, Tambutté E, Sebens KP (2003) Effect of natural zooplankton
468 feeding on the tissue and skeletal growth of the scleractinian coral *Stylophora pistillata*. Coral
469 Reefs 22, 229–240. Doi: 10.1007/s00338-003-0312-7
- 470
- 471 Fitt WK, Costley K (1998) The role of temperature in survival of the polyp stage of the tropical
472 rhizostome jellyfish *Cassiopea xamachana*. Journal of Experimental Marine Biology and
473 Ecology 222, 79–91. Doi: [https://doi.org/10.1016/S0022-0981\(97\)00139-1](https://doi.org/10.1016/S0022-0981(97)00139-1)
474
- 475 Freeman CJ, Stoner EW, Easson CG, Matterson KO, Baker D M (2016) Symbiont carbon and
476 nitrogen assimilation in the *Cassiopea*–*Symbiodinium* mutualism. Marine Ecology Progress
477 Series 544, 281–286. Doi: 10.3354/meps11605
- 478
- 479 Freeman CJ, Stoner EW, Easson CG, Matterson KO, Baker DM (2017) Variation in $\delta^{13}\text{C}$ and
480 $\delta^{15}\text{N}$ values suggests a coupling of host and symbiont metabolism in the *Symbiodinium*-
481 *Cassiopea* mutualism. Marine Ecology Progress Series 571, 245–251. Doi:
482 <https://doi.org/10.3354/meps12138>
483
- 484 FUNCEME (2019) “Fundação Cearense De Meteorologia E Recursos Hídricos.” *Postos*
485 *Pluviométricos*. [http://www.funceme.br/app/calendario/produto/municipios/maxima/diario?data](http://www.funceme.br/app/calendario/produto/municipios/maxima/diario?data=hoje)
486 [=hoje](http://www.funceme.br/app/calendario/produto/municipios/maxima/diario?data=hoje)
- 487 Graham WM, Bayha KM (2008) Biological Invasions by Marine Jellyfish. In: Nentwig W.
488 (eds) Biological Invasions. Ecological Studies (Analysis and Synthesis), vol 193. Springer,
489 Berlin, Heidelberg. https://doi.org/10.1007/978-3-540-36920-2_14
- 490 Grottoli AG, Rodrigues LJ, Palardy JE (2006) Heterotrophic plasticity and resilience in
491 bleached corals. Nature 440, 1186-9. Doi: 10.1038/nature04565
- 492 Hammer Ø, Harper DAT, Ryan PD (2001) Past: paleontological statistics software pack-age for
493 education and data analysis. Paleontol Electron 4, 1–9.
- 494
- 494 Hayes JJ, Castillo O (2017) A New Approach for Interpreting the Morisita Index of
495 Aggregation through Quadrat Size. International Journal of Geo-Informationc 6, 296.
496 Doi: 10.3390/ijgi6100296
- 497
- 497 Heins A, Glatzel T, Holst S (2015) Revised descriptions of the nematocysts and the asexual
498 reproduction modes of the scyphozoan jellyfish *Cassiopea andromeda* (Forskål, 1775).
499 Zoomorphology 134. Doi: 10.1007/s00435-015-0263-x
- 500
- 500 Hofmann DK, Fitt WK, Fleck J (1996) Checkpoints in the life-cycle of *Cassiopea* spp.: control
501 of metagenesis and metamorphosis in a tropical jellyfish. The International journal of
502 developmental biology 40, 331-8.
- 503
- 503 Hogarth PJ (2015) The biology of mangroves and seagrasses. 3rd edition. Oxford University
504 Press, Oxford.
- 505
- 505 Houlbrèque F, Ferrier-Pagès C (2009) Heterotrophy in tropical scleractinian corals. Biological
506 Reviews 84, 1–17. Doi: 10.1111/j.1469-185X.2008.00058.x

507

508 Jeffrey SW, Humphrey GF (1975) New spectrophotometric equations for determining
509 chlorophyll a, b, c1 and c2 in higher plants, algae and natural phytoplankton. *Biochemie*
510 *Physiologie der Pflanz* 167, 191-194. Doi: [https://doi.org/10.1016/S0015-3796\(17\)30778-3](https://doi.org/10.1016/S0015-3796(17)30778-3)

511

512 Klein SG, Pitt KA, Carroll AR (2016a) Reduced salinity increases susceptibility of
513 zooxanthellate jellyfish to herbicide toxicity during a simulated rainfall event.
514 *Environmental Pollution* 209, 79–86. Doi: [10.1016/j.envpol.2015.11.012](https://doi.org/10.1016/j.envpol.2015.11.012)

515

516 Klein SG, Pitt KA, Carroll AR (2016b) Surviving but not thriving: inconsistent responses of
517 zooxanthellate jellyfish polyps to ocean warming and future UV-B scenarios. *Science Report* 6,
518 28859. Doi: [10.1038/srep28859](https://doi.org/10.1038/srep28859) 1

519 Klein SG, Pitt KA, Lucas CH, Hung S-H, Schmidt-Roach S, Aranda M and Duarte CM (2019)
520 Night-Time Temperature Reprieves Enhance the Thermal Tolerance of a Symbiotic
521 Cnidarian. *Frontiers Marine Science* 6, 453. Doi: [10.3389/fmars.2019.00453](https://doi.org/10.3389/fmars.2019.00453)

522 Kristensen I, Ypma, L (1971) Ecology of the medusa *Cassiopea xamanchana* in Curaçao.
523 *Studies on the Fauna of Curaçao and other Caribbean Islands* 38, 110 - 115.

524

525 Lacerda LD (2006) Input of nitrogen and phosphorus to estuaries of northeastern Brazil from
526 intensive shrimp farming. *Brazilian Journal of Aquatic Science and Technology* 10, 13–27.
527 Doi: <http://dx.doi.org/10.14210/bjast.v10n2.p13-27>

528

529 Lampert KP (2016) “*Cassiopea* and its zooxanthellae,” in *The Cnidaria, Past, Present and*
530 *Future* eds S. Goffredo and Z. Dubinsky (Berlin: Springer), 415–423. Doi: [10.1007/978-3-319-](https://doi.org/10.1007/978-3-319-31305-4_26)
531 [31305-4_26](https://doi.org/10.1007/978-3-319-31305-4_26)

532

533 Legendre P, Fortin MJ (1989) Spatial pattern and ecological analysis. *Vegetatio*, London 80,
534 107-138.

535

536 Le Roux J, Wicczorek AM (2008) Molecular systematics and population genetics of biological
537 invasions: towards a better understanding of invasive species management. *Annals of Applied*
538 *Biology* 154, 1–17. Doi: [10.1111/j.1744-7348.2008.00280.x](https://doi.org/10.1111/j.1744-7348.2008.00280.x)

539

540 McCloskey LR, Muscatine L, Wilkerson FP (1994) Daily photosynthesis, respiration, and
541 carbon budgets in a tropical marine jellyfish (*Mastigias* sp.). *Marine Biology* 119, 13–22. Doi:
542 <https://doi.org/10.1007/BF00350101>

543

544 McGill CJ, Pomory CM (2008) Effects of bleaching and nutrient supplementation on wet
545 weight in the jellyfish *Cassiopea xamachana* (Bigelow) (Cnidaria: Scyphozoa). *Marine*
546 *and Freshwater Behaviour and Physiology* 41, 179–189. Doi: [10.1080/10236240802369899](https://doi.org/10.1080/10236240802369899)

547 Mies M, Güth AZ, Tenório AA, Banha TNS, Waters LG, Polito PS, Taniguchi S, Bicego MC,
548 Sumida PYG (2018) In situ shifts of predominance between autotrophic and heterotrophic
549 feeding in the reef-building coral *Mussismilia hispida*: an approach using fatty acid trophic
550 markers. *Coral Reefs* 37, 677-689. Doi: [10.1007/s00338-018-1692-z](https://doi.org/10.1007/s00338-018-1692-z)

551 Morandini AC, Schiariti A, Stampar SN, Maronna MM, Straehler-Pohl I, Marques AC (2016)
552 Succession of generations is still the general paradigm for scyphozoan life cycles. *Bulletin of*
553 *Marine Science* 92(3), 343–351. Doi: doi.org/10.5343/bms.2016.1018

554

555 Morandini AC, Stampar SN, Maronna MM, Silveira FL (2017) All non-indigenous species were
556 introduced recently? The case study of *Cassiopea* (Cnidaria: Scyphozoa) in Brazilian waters.
557 Journal of the Marine Biological Association of the United Kingdom 97, 321–328. Doi:
558 10.1017/S0025315416000400

559

560 Morisita M (1959) Measuring of the Dispersion of Individuals and Analysis of the
561 Distributional Patterns. Memories of Faculty Science, Kyushu University, Series E. Biology 2,
562 215-235.

563

564 Muscatine L, Porter J (1977) Reef Corals: Mutualistic Symbioses Adapted to Nutrient-Poor
565 Environments. BioScience 27, 454-460. Doi:10.2307/1297526

566

567 Newkirk CR, Frazer TK, Martindale MQ (2018) Acquisition and proliferation of algal
568 symbionts in bleached polyps of the upside-down jellyfish, *Cassiopea xamachana*. Journal of
569 Experimental Marine Biology and Ecology 508, 44–51. Doi:
570 doi.org/10.1016/j.jembe.2018.08.010

571

572 Niggli W, Wild C (2010) Spatial distribution of the upside-down jellyfish *Cassiopea* sp. within
573 fringing coral reef environments of the Northern Red Sea: implications for its life cycle.
574 Helgoland Marine Research Online First. Doi 10.1007/s10152-009-0181-8

575

576 Nóbrega GN, Ferreira TO, Romero RE, Marques AG, Otero XL (2013) Iron and sulfur
577 geochemistry in semi-arid mangrove soils (Ceará, Brazil) in relation to seasonal changes and
578 shrimp farming effluents. Environmental Monitoring and Assessment 185, 7393-407. Doi:
579 10.1007/s10661-013-3108-4.

580 Ohdera AH, Abrams MJ, Ames CL, Baker DM, Suescún-Bolívar LP, Collins AG, Freeman CJ,
581 Gamero-Mora E, Goulet TL, Hofmann DK, Jaimes-Becerra A, Long PF, Marques AC, Miller
582 LA, Mydlarz LD, Morandini AC, Newkirk CR, PutriSastia P, Samson JE, Stampar SN,
583 Steinworth B, Templeman M, Thomé PE, Vlok M, Woodley CM, Wong JCY, Martindale MQ,
584 Fitt WK, Medina M (2018) Upside-Down but Headed in the Right Direction: review of the
585 Highly Versatile *Cassiopea xamachana* System. Frontiers in Ecology and Evolution 6, 1-
586 15. Doi: 10.3389/fevo.2018.00035

587 Oksanen J., Blanchet FG, Kindt R, Legendre P, Minchin PR, O'hara RB, Simpson GL, Solymos
588 P, Stevens MHH, Wagner H (2016) vegan: Community Ecology Package. R package version
589 2.0-10. Available at: <http://CRAN.R-project.org/package=vegan>, <http://www.cran.r-project.org>, <https://github.com/vegandevs/vegan>.

590

591 Parsons TR, Maita Y, Lalli CM (1984) Manual of chemical and biological methods for seawater
592 analysis. Oxford, Pergamon Press

593

594 Pierce J (2005) A system for mass culture of upside-down jellyfish *Cassiopea* spp as a potential
595 food item for medusivores in captivity. International Zoo Yearbook 39, 62–69. Doi:
596 <https://doi.org/10.1111/j.1748-1090.2005.tb00005.x>

597

598 Purcell JE, Uye S, Lo W-T (2007) Anthropogenic causes of jellyfish blooms and their direct
599 consequences for humans: a review. Marine Ecology Progress Series 350, 153–174. Doi:
600 10.3354/meps07093

601

602 Purcell JE (2012). Jellyfish and ctenophore blooms coincide with human proliferations and
603 environmental perturbations. Annual Review of Marine Science 4, 209–235. Doi:
604 10.1146/annurev-marine-120709-142751

605

- 606 Queiroz L, Rossi S, Meireles J, Coelho C (2013) Shrimp aquaculture in the federal state of
607 Ceará, 1970–2012: trends after mangrove forest privatization in Brazil. *Ocean & Coastal*
608 *Management* 73, 54–62. Doi: [dx.doi.org/10.1016/j.ocecoaman.2012.11.009](https://doi.org/10.1016/j.ocecoaman.2012.11.009)
- 609 Richardson AJ, Bakun A, Hays GC, Gibbons MJ (2009) The jellyfish joyride: causes,
610 consequences and management responses to a more gelatinous future. *Trends in Ecology &*
611 *Evolution*. 24, 312–322. Doi: [10.1016/j.tree.2009.01.010](https://doi.org/10.1016/j.tree.2009.01.010)
- 612 Schiariti A, Christiansen E, Morandini AC, Silveira FL, Giberto DA, Mianzan HW (2012)
613 Reproductive biology of *Lychnorhiza lucerna* (Cnidaria: Scyphozoa: Rhizostomeae): Individual
614 traits related to sexual reproduction. *Marine Biology Research* 8, 255-264. Doi:
615 [10.1080/17451000.2011.616897](https://doi.org/10.1080/17451000.2011.616897)
- 616
- 617 Soares M, Rossi S, Martins F, Carneiro P (2017) The forgotten reefs: Benthic assemblage
618 coverage on a sandstone reef (Tropical South-western Atlantic. *Journal of the Marine Biological*
619 *Association of the United Kingdom* 97, 1585-1592 Doi: [10.1017 / S0025315416000965](https://doi.org/10.1017/S0025315416000965)
- 620
- 621 Soares MO, Teixeira CEP, Ferreira SMC, Gurgel ALAR, Paiva BP, Menezes MOB, Davis M,
622 Tavares TCL (2019) Thermal stress and tropical reefs: mass coral bleaching in a stable
623 temperature environment? *Marine Biodiversity* 49, 2921–2929. Doi: [Doi.org/10.1007/s12526-](https://doi.org/10.1007/s12526-019-00994-4)
624 [019-00994-4](https://doi.org/10.1007/s12526-019-00994-4)
- 625
- 626 Sousa OV, Macrae A, Menezes FG, Gomes NC, Vieira RH, Mendonça-Hagler LC (2006) The
627 impact of shrimp farming effluent on bacterial communities in mangrove waters, Ceará, Brazil.
628 *Marine Pollution Bulletin* 12, 1725-34. Doi: [10.1016/j.marpolbul.2006.07.006](https://doi.org/10.1016/j.marpolbul.2006.07.006)
- 629
- 630 Stoner EW, Layman CA, Yeager LA, Hassett HM (2011) Effects of anthropogenic disturbance
631 on the density and size of epibenthic jellyfish *Cassiopea* spp. *Marine Pollution Bulletin* 62,
632 1109–1114. Doi:[10.1016/j.marpolbul.2011.03.023](https://doi.org/10.1016/j.marpolbul.2011.03.023)
- 633
- 634 Stoner EW, Yeager LA, Layman CA (2014a). Effects of epibenthic jellyfish, *Cassiopea* spp., on
635 faunal community composition of Bahamian seagrass beds. *Caribbean Naturalist* 12, 1-10.
636
- 637 Stoner EW, Yeager LA, Sweatman JL, Sebilian SS, Layman CA (2014b). Modification of a
638 seagrass community by benthic jellyfish blooms and nutrient enrichment. *Journal of*
639 *Experimental Marine Biology and Ecology*, 461, 185-192. Doi:
640 <https://doi.org/10.1016/j.jembe.2014.08.005>
- 641 Stoner EW, Sebilian SS, Layman CA (2016) Comparison of zooxanthellae densities from
642 upside-down jellyfish, *Cassiopea xamachana*, across coastal habitats of The Bahamas. *Revista*
643 *de Biologia Marina Y Oceanografia* 51, 203-208. Doi: [10.4067/S0718-19572016000100022](https://doi.org/10.4067/S0718-19572016000100022)
- 644 Suárez-Abelenda M, Ferreira TO, Camps-Arbestain M, Rivera-Monroy VH, Macías F, Nóbrega
645 GN, Otero XL (2014) The effect of nutrient-rich effluents from shrimp farming on mangrove
646 soil carbon storage and geochemistry under semi-arid climate conditions in northern Brazil.
647 *Geoderma* 213, 551–559. Doi: <https://doi.org/10.1016/j.geoderma.2013.08.007>
- 648
- 649 Thé J, Gamero-Mora E, Silva MVC, Morandini AC, Rossi S, Soares MO (2020) Non-
650 indigenous upside-down jellyfish *Cassiopea andromeda* in shrimp farms (Brazil). Submitted
651
- 652 Todd BD, Thornhill DJ, Fitt WK (2006) Patterns of inorganic phosphate uptake in *Cassiopea*
653 *xamachana*: a bioindicator species. *Marine Pollution Bulletin* 52, 515–521. Doi:
654 <https://doi.org/10.1016/j.marpolbul.2005.09.044>

655 Unesco 1983. Chemical methods for use in marine environmental monitoring.
656 Intergovernmental Oceanographic Commission. Manuals and guides. 53p.

657

658 Valentim SS, Menezes MOB, Teixeira CEP (2018) Seasonally Hypersaline Estuaries in
659 Semiarid Climate Regions: an Example from the Northeast Brazil. *Journal of Coastal Research*
660 85, 6-10. Doi: <https://doi.org/10.2112/SI85-002.1>

661

662 Venekey V, Melo TPG (2016) Nematodes as indicators of shrimp farm impact on an amazonian
663 estuary (Curuçá, Pará, Brazil). *Brazilian Journal of Oceanography* 64, 75-87. Doi:
664 <https://dx.doi.org/10.1590/S1679-87592016108206401>

665 Welsh DT, Dunn RJK, Meziane T (2009) Oxygen and nutrient dynamics of the upside down
666 jellyfish (*Cassiopea* sp.) and its influence on benthic nutrient exchanges and primary
667 production. *Hydrobiologia* 635, 351–362. Doi: 10.1007/s10750-009-9928-0

668

669 Yu R, Leung PG, Bienfang P (2006) Optimal production schedule in commercial shrimp
670 culture. *Aquaculture* 254, 426-441. Doi: <https://doi.org/10.1016/j.aquaculture.2005.11.022>.

671

672 Zarnoch CB, Hossain N, Fusco E, Alldred M, Hoellein TJ, Perdikaris S (2020) Size and density
673 of upside-down jellyfish, *Cassiopea* sp., and their impact on benthic fluxes in a Caribbean
674 lagoon. *Marine Environmental Research* 154, 104845. Doi: [https://doi.org/10.1016/](https://doi.org/10.1016/j.marenvres.2019.104845)
675 [j.marenvres.2019.104845](https://doi.org/10.1016/j.marenvres.2019.104845).

676