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## 7 **How important is budding speciation for comparative studies?**

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13 **Running title:** BUDDING SPECIATION IN COMPARATIVE METHODS

14 **Abstract:** The acknowledgment of evolutionary dependence among species has fundamentally  
15 changed how we ask biological questions. Phylogenetic models became the standard approach for  
16 studies with three or more lineages, in particular those using extant species. Most phylogenetic  
17 comparative methods (PCMs) translate relatedness into covariance, meaning that evolutionary  
18 changes before lineages split should be interpreted together whereas after the split lineages are  
19 expected to change independently. This clever realization has shaped decades of research. Here we  
20 discuss one element of the comparative method often ignored or assumed as unimportant: if nodes  
21 of a phylogeny represent the dissolution of the ancestral lineage into two new ones or if the ancestral  
22 lineage can survive speciation events (i.e., budding). Budding speciation is often reported in  
23 paleontological studies, due to the nature of the evidence for budding in the fossil record, but it is  
24 surprisingly absent in comparative methods. Here we show that many PCMs assume that divergence  
25 happens as a symmetric split, even if these methods don't explicitly mention this assumption. We  
26 discuss the properties of trait evolution models for continuous and discrete traits and their adequacy  
27 under a scenario of budding speciation. We discuss the effects of budding speciation under a series  
28 of plausible evolutionary scenarios and show when and how these can influence our estimates. We  
29 also propose that long-lived lineages that have survived through a series of budding speciation  
30 events and given birth to multiple new lineages can produce evolutionary patterns that challenge our

31 intuition about the most parsimonious history of trait changes in a clade. We hope our discussion  
32 can help bridge comparative approaches in paleontology and neontology as well as foster awareness  
33 about the assumptions we make when we use phylogenetic trees.

34

35 **Keywords:** budding speciation; diversification; fossil record; model adequacy; phylogenetic  
36 comparative methods; speciation mode.

37       Phylogenetic trees are the main representation of evolutionary relationships among lineages  
38 and stand as a symbol of evolutionary thought. However, their relatively simple structure is not  
39 capable of informing us about all aspects of evolution. Processes such as horizontal gene transfer,  
40 hybridization, and introgression can produce complex evolutionary relationships that challenge the  
41 explanatory power of bifurcating phylogenies (Philippe and Douady, 2003; Mallet et al., 2016;  
42 Bastide et al., 2018). Interestingly, the graphical representation of bifurcating trees could even  
43 influence how we think (Baum et al., 2005). A node connecting one ancestral branch to two new  
44 ones can suggest evolutionary histories much simpler than what we observe in nature and potentially  
45 downplay important aspects of macroevolution. For instance, if read literally, bifurcating  
46 phylogenies can be seen as speciation events that happened due to the split of an ancestral lineage  
47 into two new ones, coincident with the extinction, or dissolution, of the ancestral lineage (Meier and  
48 Willmann, 2000; Bokma, 2008). This view makes the concept of a lineage synonymous with a  
49 branch of a phylogenetic tree. However, evidence from empirical systems shows the mode of  
50 speciation can be varied and often complex (Rosenblum et al., 2012), including frequent instances of  
51 budding speciation (Wagner, 1998; Funk and Omland, 2003; Gottlieb, 2004; Crawford, 2010;  
52 Anacker and Strauss, 2014; Otero et al., 2019; Patsis et al., 2021).

53       The paleontological literature often adopts the representation of budding trees (e.g., Raup  
54 and Gould, 1974; Raup, 1985; Foote, 1996; Wagner, 1998; Benton and Pearson, 2001; Silvestro et  
55 al., 2018) that inform which branches are new lineages and which are the continuation of the  
56 ancestral lineage. Budding is recognized in the fossil record as a cladogenetic event in which a new  
57 lineage appears as a branching of an older lineage that can still be found after the speciation event  
58 (e.g., Foote, 1996). Although budding speciation is commonly reported in paleontology, it is rarely  
59 incorporated into phylogenetic comparative models (PCMs). Bokma and colleagues have developed  
60 a series of PCMs incorporating budding and the effect of punctuated equilibrium (Bokma 2002;

61 Bokma, 2008; Matilla and Bokma, 2008; Monroe and Bokma, 2009; Bokma, 2010; Jansen et al.,  
62 2022). Unfortunately, these and other similar methods (Bartoszek, 2014; Bartoszek, 2020, Pagel et  
63 al., 2022) have not been widely used in the PCM literature, perhaps due to the perception that  
64 bifurcating molecular phylogenies show no evidence of budding speciation. Here we bring a  
65 different, and perhaps controversial, point of view; that budding speciation can be common, that it  
66 might affect inferences of trait evolution, that it can be detected in molecular phylogenies, and it  
67 should be considered in PCMs even when there is no information from the fossil record evidencing  
68 its role in the diversification of the group.

69         There has not been a consensus about the effect of budding speciation on our estimates of the  
70 tempo and mode of trait evolution using PCMs. Bokma (2008) implemented a trait evolution model  
71 to estimate the contribution from cladogenetic and anagenetic changes, however, there is no  
72 investigation of the impact of the cladogenetic process in inferences using PCMs that do not  
73 accommodate such effects. De Lisle and colleagues (2021) explored the effect of shifts in the  
74 adaptive optima on extinction rates using a model based on population-level dynamics and showed  
75 that lineages would rarely survive peak shifts and those occupying stable optima are expected to  
76 have a higher chance of survival. In turn, Duchon and colleagues (2021) studied how cladogenesis  
77 changes the average species phenotype using individual-level simulations and showed that new  
78 lineages budding off from an ancestral population can show a significant phenotypic deviation due  
79 to neutral processes (see Gaborieau et al., 2023 for a similar approach). Combined, these results  
80 point to the idea that long-lived evolutionary lineages might occupy stable optima (Eldredge and  
81 Gould, 1972; Gould and Eldredge, 1993; also see Goldberg and Foo, 2020) whereas new lineages  
82 may bud off with distinct phenotypes, due to the effect of cladogenesis on the species trait and  
83 selection to occupy a distinct region of the morphospace (Eldredge and Gould, 1972; Gould and  
84 Eldredge, 1993; Bokma, 2008; Matilla and Bokma, 2008).

85         Here we discuss in which circumstances budding speciation can affect our estimates of the  
86 tempo and mode of trait evolution. More specifically, we review the properties of PCMs of trait  
87 evolution that dictate whether or not budding speciation can influence our conclusions. In our view,  
88 most PCMs of trait evolution were developed with the strong assumption that speciation is  
89 symmetric, lineages cannot continue after speciation events, and there is no effect of budding in  
90 molecular phylogenies. Here we discuss how budding speciation can bias our estimates using

91 simulations as an argumentative guide to our narrative. Distinct from previous studies (Matilla and  
92 Bokma, 2008; Silvestro et al., 2018; De Lisle et al., 2021; Duchen et al., 2021; Crouch et al., 2021  
93 among others), our discussion focuses on cases in which budding speciation is not considered when  
94 using PCMs.

95

96

## 97 **What is budding and how to recognize it?**

98         Budding is defined as a speciation event in which the new species co-occurs in time with its  
99 direct ancestral lineage (Foote, 1996), meaning the ancestral lineage continues to exist after  
100 speciation. We use the term progenitor lineage to help differentiate the parent from the daughter  
101 lineage (see Gottlieb, 2004). A progenitor lineage is a lineage that has given birth to one or more  
102 new lineages through budding. Budding speciation is inherent in Mayr's (1942) concept of  
103 speciation via peripatric speciation. Also, as discussed by Eldredge and Gould (1972) and Grant  
104 (1981), drift likely has an important impact on small founder populations and, naturally, will have  
105 consequences for trait evolution (see De Lisle et al., 2021). Perhaps due to its intrinsic role in  
106 diversification, the neontological literature has attributed different nomenclature to what is  
107 fundamentally budding speciation. In this section, we discuss how budding speciation has been  
108 recognized in the literature, which patterns might be the result of past budding events, and how  
109 budding can be detected using data from extant species. The reader will note that we attribute a  
110 variety of processes to the effect of budding speciation. Indeed, one of the main goals of this  
111 discussion is to bring awareness to the role of budding speciation in studies of macroevolution and  
112 how it connects to multiple patterns we observe in phylogenetic trees.

113         In the absence of the fossil record, budding has been recognized as a new lineage formed  
114 within or at the edge of the ancestral lineage (Anacker and Strauss, 2014) or as a biological cause of  
115 paraphyletic species (Funk and Omland, 2003; also see Fig. S1). This has also been associated with  
116 the hypothesis of Punctuated Equilibrium (Eldredge and Gould, 1972; Gould and Eldredge, 1993),  
117 since peripheral populations can become isolated and show fast trait evolution due to selection  
118 towards a new adaptive peak and/or the effects of drift in small populations, leading to speciation  
119 (Mayr 1942; Simpson, 1944; Eldredge and Gould, 1972; Grant, 1981; Gould and Eldredge, 1993;  
120 Bokma, 2008; Matilla and Bokma, 2008; De Lisle et al., 2021). Budding speciation is considered to

121 be opposed by bifurcation—the split of an ancestral lineage into two new ones (Fig. S1). Hagen and  
122 colleagues (2015), for example, utilized the term *symmetrical* speciation to capture the role of  
123 allopatric speciation and opposed it to *asymmetrical* speciation which represents peripatric  
124 speciation with the continuation of the progenitor species—thus, budding. Although these authors  
125 use distinct nomenclature, each is an example of budding. The paleontological literature suggests  
126 budding is a common evolutionary pattern and some argue it represents the majority of speciation  
127 events observed in deep time. For example, Wagner (1998) used budding speciation to estimate a  
128 phylogeny of hyenas that implied reduced stratigraphic debt (i.e., less ghost lineages) when  
129 compared with alternative trees, indicating a better fit to the fossil record. Aze and colleagues (2011)  
130 reconstructed a large phylogeny of macroperforate foraminifera in which most cladogenetic events  
131 were recognized as budding through analysis of morphological characters. Bapst and Hopkins  
132 (2017) applied an explicit probabilistic model to date a phylogeny of trilobites and also show that  
133 budding events are often supported by the fossil record. Similarly, Parins-Fukuchi (2021) re-  
134 evaluated the diversification of hominins and suggests the occurrence of budding speciation events.  
135 In contrast, there is little to no mention of budding speciation in the neontological literature which,  
136 in our view, creates an undesirable disconnect between paleontology and neontology (but see  
137 Silvestro et al., 2018). One could argue that this absence is due to the impossibility of detecting  
138 budding using molecular phylogenies, however, as we discuss below, we disagree with this  
139 sentiment.

140       If budding speciation is frequent, we expect to recover recent events of budding using  
141 molecular data. When a new lineage buds off from its ancestral lineage, the progenitor species  
142 becomes paraphyletic (Funk and Omland, 2003; see Fig. S1B). The advantage of neontological data  
143 is that molecular phylogenies can show evidence of budding independent of the use of  
144 morphological divergence to estimate the tree, which is necessary to both estimate phylogenies and  
145 detect speciation based on fossil remains (Foote, 1996; Wagner, 1998; Bapst, 2013). If the new  
146 lineage maintains cohesion and does not go extinct (see De Lisle et al., 2021) or is not reabsorbed  
147 via hybridization with the progenitor (Taylor et al., 2006; Richmond and Jockush, 2007; Behm et al.,  
148 2010; Lackey and Boughman, 2017), gene flow among the populations of the progenitor should  
149 complete sorting and the progenitor and daughter lineages will eventually become sister species in  
150 estimated molecular phylogenies—erasing the signal of budding. Thus, budding speciation can be

151 detected using molecular phylogenies, but its signal disappears over time whereas, in the fossil  
152 record, the information is preserved if the record is reasonably complete. Otero and colleagues  
153 (2019) show an interesting case in *Iberodes* plants which underwent two events of budding within  
154 the last 5 million years. In both instances, the new lineage evolved distinct morphological and  
155 ecological traits (Otero et al., 2019). *Iberodes* has inland and coastal species and the potential  
156 change in selective pressure together with the peripheral distribution of the younger coastal lineages  
157 likely were key factors for budding divergence. Similarly, Papuga and colleagues (2018) show  
158 peripheral plant populations that have lower niche breadth (i.e., are more specialized) than central  
159 populations as well as divergence in ecological traits (i.e., soil parameters), both factors that can  
160 cause budding by ecological speciation. Strong evidence for budding speciation was also detected  
161 from molecular phylogenies by Baldwin (2005), showing that *Layia glandulosa* (a Compositae  
162 plant) is the progenitor species for *L. discoidea*. Anacker and Strauss (2014) tested 71 sister pairs  
163 and demonstrated that young divergences frequently show overlapping and asymmetrical ranges—  
164 another indication of budding speciation. This asymmetry was not detected among older clades,  
165 suggesting the signal of budding on the geographic distribution of sister pairs is lost as lineages get  
166 older. Furthermore, taxonomic revisions that re-name paraphyletic species into several new species  
167 also erase the signal of budding.

168       If budding is frequent, and we suspect it is, it can be an important factor in understanding  
169 trait evolution because peripheral populations can show distinct mean phenotypic values (Papuga et  
170 al., 2018) and divergence through budding can generate new lineages with distinct average  
171 phenotypes (Gottlieb, 2004; Duchen et al., 2021; Gaborieau et al., 2023) and evolutionary  
172 trajectories (De Lisle et al., 2021). If we assume molecular phylogenies are literal bifurcating trees,  
173 despite the evidence for budding speciation, then PCMs might be based on inadequate assumptions.  
174 In the next two sections, we visit the most popular PCMs and discuss scenarios in which the  
175 presence of budding would, or would not, affect our estimates.

176

### 177 **When budding doesn't matter**

178       Raup (1985) stated that budding should not influence estimates of net diversification rate  
179 because the addition or subtraction of lineages at any given time would be perceived similarly if we  
180 represent a phylogeny either by budding or bifurcation. This question has been revisited by Bapst

181 and Hopkins (2017) and Crouch et al. (2021), both showing that budding can change divergence  
182 time estimation and alter estimates of the accumulation of lineages through time (also see Wagner,  
183 1998). Thus, budding should not influence the net diversification rate only if the true dated tree is  
184 known, otherwise, changes in divergence time estimation can potentially impact estimates of  
185 diversification down the line.

186

187         With respect to models of trait evolution, budding should not influence our estimates if  
188 changes happening at any point in time, and at any branch of the phylogeny, are independent of the  
189 prior history of the lineage and their ancestors. Two important simplifications were introduced when  
190 Felsenstein described the method of independent contrasts (1981; 1985); trait changes happen  
191 independently in each branch of the phylogeny and evolutionary changes at each point along a  
192 branch are independent and identically distributed (iid). Most models of trait evolution share these  
193 assumptions (see review in O'Meara, 2012; Pennell and Harmon, 2013). However, few, if any,  
194 PCMs were created with the intent to accurately describe evolution in a mechanistic way, and the  
195 use of simplifications does not mean we assume evolution follows these rules.

196         If models of trait evolution that assume a homogeneous process across all branches of the  
197 tree are adequate representations of macroevolution, the incorporation of budding speciation will not  
198 change our estimates. This is because differentiating lineages in the phylogenetic tree will have no  
199 influence on the underlying model—lineages become effectively interchangeable. However, this is  
200 not the trend that we are currently observing in PCM development. Extensions allowing  
201 heterogeneity in the process, often associated with some predictor, have been shown to better  
202 capture the variation of empirical data (e.g., Eastman, 2011; Rabosky et al., 2014; Uyeda and  
203 Harmon, 2014; Caetano et al., 2018; Pagel et al., 2022). More recently, studies have demonstrated  
204 that rate heterogeneity should be taken into account even when no a priori predictors are present  
205 (e.g., Rabosky and Goldberg, 2015; Beaulieu and O'Meara, 2016; Caetano et al., 2018; May and  
206 Moore, 2020). Development of more adequate models often means the increase in model complexity  
207 to reflect the dynamic nature of macroevolution and, as a result, hint that the condition of  
208 homogeneous and memoryless evolutionary changes with interchangeable lineages—under which  
209 budding would not matter—is unlikely across the tree of life. Below we discuss how budding could  
210 be generating heterogeneity in the phylogenetic history of phenotypes and in which ways the results

211 affect our conclusions about trait evolution.

212

### 213 **When budding matters**

214 Budding is expected to be important in any evolutionary scenario in which the identity of  
215 evolutionary sister lineages is relevant. This might be the case if lineage age influences the tempo  
216 and/or mode of trait evolution (Hagen et al., 2018; Goldberg and Foo, 2020) or if the age of  
217 competing lineages is important to predict their competitive strength and/or risk of extinction (Ezard  
218 et al., 2011; Rosenblum et al., 2012; Carrillo et al., 2020; Januario and Quental, 2021). Although  
219 there are other evolutionary processes under which the identity of lineages might be important, here  
220 we focus on these two scenarios for simplification. In contrast, there are special cases that generate  
221 heterogeneity in trait evolution but under which budding likely is not relevant. For example, if shifts  
222 in rates of trait evolution are due to abiotic causes equally affecting all lineages concurrent with the  
223 event, such as response to climatic changes or mass extinctions, then, everything else being equal,  
224 we would expect responses to be independent of lineage identity (e.g., Clavel and Morlon, 2017).  
225 Below we enumerate scenarios in which we argue that budding speciation could influence our  
226 conclusions about the tempo and mode of trait evolution when using PCMs.

227

#### 228 *1) When evolutionary changes are concentrated at or near lineage origination*

229 The central distinction between budding and bifurcation is the age contrast between  
230 progenitor and daughter lineage immediately after divergence. The daughter species will usually  
231 have a smaller population size and geographic distribution (Foote et al., 2007; Liow and Stenseth,  
232 2007) and might undergo quick phenotypic change as they move towards a new adaptive peak  
233 (Eldredge and Gould, 1972; Gould and Eldredge, 1993; Hunt et al., 2008; De Lisle et al., 2021). In  
234 contrast, progenitor lineages might show a slowdown in trait evolution due to prolonged time under  
235 a stable adaptive zone (Goldberg and Foo, 2020; De Lisle et al., 2021). If lineage age is related to  
236 the tempo of trait evolution, such that younger lineages are expected to show faster rates of trait  
237 change, we would expect relatively more evolution to happen in a daughter lineage when compared  
238 to its progenitor. Thus, the disparity between two descendants of a budding node in a phylogenetic  
239 tree should not be attributed to equal amounts of change at each branch because budding suggests  
240 evolution will be concentrated in the daughter lineage (Fig. 1 top left panel).



241

242 2) *When daughter lineage survival depends on being ecologically different from its progenitor*

243 The asymmetry in age generated by budding speciation can influence the competitive  
244 strength of daughter lineages relative to their progenitors and, as a result, also the extinction risk of  
245 the younger lineage (Ezard et al., 2011; Rosenblum et al., 2012; Carrillo et al., 2020; Januario and  
246 Quental, 2021). Progenitor lineages are expected to have larger population sizes and geographic  
247 ranges (Anacker and Strauss, 2014) which, everything else being equal, improves their chance of  
248 survival in interspecific competition with newly formed species. When competition between  
249 progenitor and daughter lineages is present, daughter lineages that have lived enough to be sampled,  
250 either in the fossil record or still living today, are expected to be sufficiently distinct from their  
251 progenitors to have escaped competitive exclusion (De Lisle et al., 2021). Of course, competition is  
252 not exclusive to budding. However, budding could potentially intensify the effect of interspecific  
253 competition, and eventually increase heterogeneity in trait evolution.

254 Although we predict an intensifying effect of budding speciation on interspecific  
255 competition, nature of other interactions might be more complex. Nuismer and Harmon (2015)  
256 demonstrated mathematically the effect of the mode of trait evolution and phylogenetic diversity  
257 (PD) in the outcome of interspecific interactions in communities of closely related taxa. They  
258 showed that PD is a good predictor of interspecific interactions if these are dependent on phenotypic  
259 matching, such as competition, with more closely related lineages showing stronger interspecific  
260 interactions. Budding could change the relationship between PD and expected trait similarity,  
261 because long-lived progenitor species would accumulate fewer evolutionary changes than expected  
262 under a homogeneous trait evolution model, such as Brownian motion, causing the role of the  
263 phylogeny as a predictor to become less prevalent. In contrast, Nuismer and Harmon (2015) show  
264 that under stabilizing coevolution the phylogeny is a poor predictor of interactions, and we do not  
265 expect that budding would influence this result.

266

267 **How do budding speciation and lineage-age-dependent processes influence estimates of trait**  
268 **evolution?**

269 We use simple simulations to illustrate different scenarios in which budding speciation  
270 should impact trait evolution and, more importantly, discuss if these deviations hinder our

271 understanding of phenotypic evolution using phylogenetic trees. We explored the impact of budding  
272 on the parameter estimates and adequacy of PCMs for continuous and discrete traits. We also  
273 investigated how likely is budding speciation to produce erroneous estimates of ancestral states. We  
274 focused our attention on phylogenies of extant species, which are most often estimated using  
275 molecular data, and in the absence of fossil tips.

276

### 277 *Simulation of trait evolution under budding*

278 We simulated 50 phylogenetic trees using a constant rate birth-death model ( $\lambda = 0.2$ ,  $\mu = 0.1$ )  
279 with root age set to 40 My and excluding all extinct lineages. To reduce variation in tree size we  
280 used rejection sampling to keep only phylogenies with 250 to 350 extant lineages. We used the same  
281 pool of 50 trees to perform all simulation replicates and conducted pairwise comparisons across  
282 scenarios. We simulated budding speciation using an independent binary variable to control the  
283 presence of budding on each node of the tree. As a result, long-lived progenitor lineages are  
284 produced by random events of successive budding events. We produced four scenarios of budding  
285 speciation, with frequencies of 0%, 25%, 50%, and 100% of the nodes.

286 We simulated a single continuous trait using a Brownian motion model ( $\sigma^2 = 0.2$ ) and a  
287 discrete trait with three states using an equal rates Markov model (transition rates = 0.02). To  
288 emulate a scenario of fast evolution in younger lineages we introduced a rate slowdown process.  
289 Relative rates of change, for both continuous and discrete traits, varied along the branches of the tree  
290 following a scaling factor ( $s$ ) computed as a function of lineage-age ( $a$ ), such that

$$291 \quad s(a_i) = \frac{1}{\exp(za_i)} \quad (1)$$

292 where  $a_i$  is the average lineage-age at time interval  $i$  (i.e., lineage-age at the midpoint of the time  
293 interval  $i$ ) and  $z$  is the parameter controlling the rate slowdown. Note that progenitor lineages can  
294 span multiple branches of the tree (see Fig. 1). In order to compute  $a$ , we divided the branches of the  
295 tree into  $i$  time intervals of length  $1 \times 10^{-3}$  of the tree height. At lineage-age of 0 My, for instance,  $s$  is  
296 equal to 1 and it decays as a function of  $z$  (Fig. 1). We simulated three scenarios of lineage-age  
297 dependent rates of trait evolution: a mild effect ( $z = 0.042$ ); a medium effect ( $z = 0.279$ ); and a  
298 strong effect ( $z = 0.925$ ). The parameter values were chosen to produce a rate reduction of 10%,  
299 50%, and 90% of the base rate when a lineage becomes 2.5 My old, respectively. Because the base  
300 rate is scaled by  $s$ , which depends on the lineage age and the budding history of each phylogeny, the

average rate of trait evolution for each phylogeny ( $\bar{r}$ ) can vary across replicates. We standardized across the tree (for both discrete and continuous traits) to differentiate the rate slowdown of each lineage from the confounding effect of an overall change in the average rate across the phylogeny.

For that, we computed the weighted average as

$$\bar{r} = \sum_i r s_{(a_i)} t_i / \sum_i t_i \quad (2)$$

where  $r$  is the base rate of trait evolution (i.e., the  $\sigma^2$  for the BM model and the transition rate for the equal rates Markov model), and  $s_{(a_i)}$  is the slowdown scale factor at a time interval  $t_i$  (see Equation (1)). Then we chose  $r$  values that minimized the distance of  $\bar{r}$  among replicates.

We also explored the effect of cladogenetic changes on discrete traits (associated or not with budding). Cladogenetic changes were simulated as a change with equal probability to any state immediately after speciation. When budding is present, cladogenetic changes were restricted to daughter lineages whereas it could happen to either or both lineages in the absence of budding. We also explored a scenario in which cladogenetic changes restricted to daughter lineages (thus dependent on budding) produce convergence among all daughters of the same long-lived progenitor lineage (see examples in Figs. 2 and 3). A detailed report of the simulation is available in the Supplementary Materials (<https://doi.org/10.5061/dryad.qbzkhl8kw>).

### *Evaluating model adequacy and errors in ancestral estimation*

We used the method described by Pennell and colleagues (2015) to evaluate the adequacy of PCMs for continuous traits in the presence of budding speciation and lineage-age-dependent rates of trait evolution. This method computes a pool of summary statistics (see Table 1 in Pennell *et al.*, 2015) and compares each with an expected distribution estimated from the data. If the model is adequate, the observed summary statistics should fall around the mean of the null distribution whereas values outside the 95% highest density interval indicate that the PCM is inadequate. To evaluate the effect of budding in the ancestral estimation of discrete states we used an index of how incorrect the estimate at a node is with respect to the true history of the trait. We measured the highest marginal probability among all states excluding the true state for the nodes as an estimate of “wrongness”. This metric reflects how likely the state of a node would be estimated as the wrong ancestral state. Note that this is distinct from uncertainty because wrongness is maximized when we have certainty of the ancestral state but it is incongruent with the true (simulated) history.

331 Uncertainty is a lesser problem than wrongness because we will not, or at least should not, support  
332 or refute evolutionary hypotheses based on uncertain estimates. Wrongness, on the other hand, can  
333 result in misleading interpretations. We then used linear mixed models to test for the association  
334 between node age and wrongness across all simulation scenarios and selected the best model using  
335 the Akaike Information Criterion (AIC).

336

337 *Effects of budding and lineage-age dependent processes on the adequacy of continuous trait*  
338 *evolution models*

339       After simulating continuous traits under 12 scenarios, varying the strength of the lineage-age  
340 dependent process and the frequency of budding speciation, we estimated parameters for single rate  
341 Brownian motion (Felsenstein, 1973), variable rate BM (Eastman et al., 2011), single optimum  
342 Ornstein-Uhlenbeck (Butler and King, 2004), and Early-Burst (Harmon et al., 2010) models as  
343 implemented in *geiger* (Pennell et al., 2014). Note that none of those models is the true model that  
344 generated the data. Our goal is to evaluate which is the preferred model among the suite of PCMs  
345 most used in the literature and to better understand the potential effects of budding speciation on our  
346 inferences. We also hope that this simple illustration through the use of simulations motivates  
347 further research in model development. Overall, variable rate BM models showed significant  
348 improvement in model adequacy under budding.

349       Model adequacy tests for a homogeneous rate BM model (Pennell et al., 2015) detected a  
350 negative slope of the linear fit between node depth and the size of the phylogenetic independent  
351 contrasts ( $S_{\text{hgt}}$ ) indicating that larger trait changes are more frequently detected closer to the tips  
352 (Fig. S3). However, a variable rate BM model does not show evidence for such deviation (Fig. S3),  
353 suggesting this is an effect of underestimating the rate variation introduced by budding (which  
354 introduces heterogeneity in a different way than the variable rate BM models). A regression of  
355 phylogenetic independent contrasts (PICs) and their expected variance ( $S_{\text{var}}$ ) shows that nodes  
356 connected by short branches are associated with more trait change (Fig. S2), independently of  
357 whether the fitted BM model was homogeneous or not. Inadequacy of  $S_{\text{var}}$  is expected due to the  
358 concentration of rates early in the history of lineages and the stronger effect of slowdown on the  
359 more longevous lineages when compared with short-lived ones—a pattern that is expected under  
360 budding speciation and punctuated equilibrium. The deviances for  $S_{\text{hgt}}$  and  $S_{\text{var}}$  are only detectable

361 when the rate slowdown is strong, meaning that a relaxed rates model (Eastman et al., 2011) seems  
362 to be able to adequately describe trait variation if lineage-age effects are mild, but not if they are  
363 strong. Inadequacies in both  $S_{\text{hgt}}$  and  $S_{\text{var}}$  point to trait changes concentrated close to the tips, which  
364 is expected since molecular trees have an accumulation of nodes near the present, some of these  
365 generated by budding, producing new lineages with higher rates of trait evolution. Model  
366 inadequacy could be an artifact of unobserved speciation events deeper in the tree but results remain  
367 constant when we replicate analyses including extinct lineages. Deviations of  $S_{\text{hgt}}$ ,  $S_{\text{var}}$ , and  $C_{\text{var}}$   
368 (coefficient of variation of PICs, a measure of rate heterogeneity) get stronger as the intensity of the  
369 lineage-age-dependent slowdown factor increases. On the other hand, changing the frequency of  
370 budding speciation, while controlling for the strength of the slowdown factor, did not change the  
371 patterns of model adequacy across all summary statistics we investigated (Figs. S2-4). We did not  
372 verify any deviation from the remaining summary statistics adopted by Pennell and colleagues  
373 (2015). In summary, if budding speciation is frequent and there is strong age-dependent trait  
374 evolution (punctuated equilibrium representing an extreme version of this), current inference  
375 methods would have trouble adequately capturing patterns of trait evolution.

376         With respect to the support for alternative trait evolution models (i.e., Brownian motion,  
377 Ornstein-Uhlenbeck, and Early-Burst) as a function of lineage-age dependent rate variation, the OU  
378 model shows a marked increase in AIC weights in response to stronger slowdown factors (Figs. S5  
379 and S6). In the majority of cases, the phylogenetic half-life was estimated to be multiple times  
380 longer than the age of the clade (40 My) indicating very weak attraction towards the evolutionary  
381 optimum (Cooper et al. 2016). Average phylogenetic half-life estimated across replicates was only  
382 shorter than clade age when the strongest slowdown factor was applied (Fig. S7). This means that an  
383 OU process is only supported when progenitor lineages are practically in stasis whereas virtually all  
384 trait evolution is concentrated on the origination of new lineages following budding speciation. In  
385 other words, if budding speciation produces a pattern congruent with Punctuated Equilibrium we  
386 expect support for OU models. These results remain constant regardless of the frequency of budding  
387 speciation used to simulate the data (Fig. S4) or the inclusion of extinct lineages.

388         When we simulate continuous traits under a lineage-age dependent process, the BM model  
389 with varying rates adequately describes most characteristics of the data but fails to capture the  
390 concentration of trait changes on shorter branches ( $S_{\text{var}}$ ). If lineage-age dependent processes happen

391 in nature, our results reinforce the cautionary note that parameter estimates can be more informative  
392 than model choice alone (Cooper et al. 2016). Although the majority of scenarios supported OU  
393 models, only the strongest case of rate slowdown resulted in relatively short phylogenetic half-life  
394 values. Fortunately, the deviance of the slope of absolute contrasts as a function of their expected  
395 variance ( $S_{\text{var}}$  - Pennel et al. 2015) can help to detect a concentration of trait change towards shorter  
396 branches, even when controlled for rate variation, which is one of the expectations of lineage-age-  
397 dependent rates of evolution. Those simulations are far from being comprehensive (our goal was not  
398 to be exhaustive but to provide examples to support our narrative), but they emphasize the potential  
399 effects of not explicitly considering budding speciation, in particular when age-dependent trait  
400 evolution is present. Future simulation studies should more deeply focus on the different aspects  
401 superficially touched here as well as on others not discussed.

402

#### 403 *Effects of budding speciation and lineage-age dependent processes on ancestral estimation*

404       Here we investigate how budding affects our estimations of ancestral state for discrete traits.  
405 As expected, all fitted models show a strong positive association between node age and wrongness  
406 (Fig. 4), meaning that ancestral estimation of nodes closer to the root of the tree is more likely to be  
407 misleading. The best-ranked linear mixed model using AIC (Table S1) suggests that budding  
408 speciation has a significant effect on wrongness when cladogenetic trait changes are also present  
409 (see example in Fig. 3). Without cladogenetic changes, there is no detectable difference between the  
410 null model (homogeneous rates and bifurcating speciation) and the model with budding speciation  
411 (Fig. 4). Budding associated with cladogenetic changes increases the chance of misleading ancestral  
412 state estimation, especially for younger nodes. This result is somewhat unexpected and important  
413 because younger nodes are often expected to have more information than older ones (Schultz et al.,  
414 1996; Boyko and Beaulieu, 2021).

415

#### 416 **Can budding help us understand complex histories of trait evolution?**

417       Here we used simulations to exemplify the effect of budding speciation in PCMs of trait  
418 evolution. Our initial results show that budding has an impact but does not completely hinder the  
419 utility of the most popular models of trait evolution. Some of the effects we report here, for the most  
420 part, can be translated as heterogeneity in trait evolution among lineages. Different from other

421 sources of heterogeneity (e.g., Uyeda and Harmon, 2014; Boyko and Beaulieu, 2021), budding  
422 produces variation dependent on lineage identity, requiring the identification of progenitor lineages  
423 potentially comprising multiple contiguous branches of a phylogeny. From this perspective, we are  
424 optimistic about incorporating budding speciation into PCMs, and we hope our initial discussion on  
425 the subject motivates further research on how we can properly incorporate budding speciation and  
426 age-dependent trait evolution in PCMs. In fact, it is plausible that a portion of the intrinsic  
427 heterogeneity captured by rate-varying models, such as hidden rates models for discrete traits  
428 (Beaulieu and O’Meara, 2016; Caetano et al., 2018; Boyko and Beaulieu, 2021) and those applying  
429 reversible jump MCMC for continuous traits (Eastman et al., 2011; Rabosky et al., 2014; Uyeda and  
430 Harmon, 2014), is due to the effect of budding speciation.

431         Distinct from scenarios in which some predictor trait is responsible for rate shifts, budding is  
432 expected to affect trait evolution dependent on the mode of speciation. This introduces a  
433 complication because we need to reconstruct the budding history of lineages, which does not easily  
434 leave a trace on molecular phylogenies (e.g., it needs samples of multiple populations of recently  
435 diverged lineages). One potential solution is to use data augmentation (e.g., Quintero and Landis,  
436 2020) to co-estimate budding history and trait evolution model parameters using simulations. This  
437 approach could be challenging because both the frequency of budding speciation and the location of  
438 the progenitor lineages would need to be sampled. However, our inability to pinpoint the location of  
439 progenitor lineages should not be used as an argument for ignoring its effect on trait evolution.  
440 Today we have several PCMs that are able to recover the signal of shifts in the tempo and mode of  
441 trait evolution without a priori hypotheses (Eastman, 2011; Rabosky et al., 2014; Uyeda and  
442 Harmon, 2014; Pagel et al., 2022) and, more importantly, there is evidence that such methods  
443 improve model adequacy (Rabosky and Goldberg, 2015; Beaulieu and O’Meara, 2016; Caetano et  
444 al., 2018). We suggest that budding speciation should be considered as a confounding factor akin to  
445 rate heterogeneity, which needs to be taken into account when estimating the history of trait  
446 evolution using molecular phylogenies—even if budding is not the focus of the study.

447         Another challenge is that progenitor lineages can produce scenarios incongruent with the  
448 most parsimonious history for a trait (e.g. Figs. 2 and 3). For example, ancestral estimates of the  
449 scenario shown in Figure 3 in the absence of fossil information would suggest, with confidence, that  
450 the trait history is due to convergence. This is a scenario in which PCM estimates can conflict with

external evidence of homology. For instance, Pyron (2015) discusses the inference of multiple transitions from viviparity back to oviparity in snakes, based on PCMs, despite the external evidence based on development and physiology against it (Griffith et al., 2015). Pyron (2015) suggests that comparative approaches should not ignore external evidence but also that findings from phylogenetic inferences should be further investigated integratively. However, an unlikely ancestral reconstruction of parity might simply mean that the model is inadequate. For instance, budding speciation could help explain oviparous lineages nested deep into viviparous clades as descendants of long-lived progenitor lineages (see discussion in Pyron 2015). The budding speciation scenario would require many additional evolutionary transitions, but it would support the extensive knowledge about genetics, development, and physiology of snakes (see discussion in Griffith et al., 2015). In our view, when there is a clash between model estimates based on projections into millions of years in the past and biological knowledge, it is wise to review our models and ponder which important processes the model might be failing to capture, including the possibility of budding speciation.

465

#### 466 **Closing remarks**

Budding speciation might indirectly or directly impact both estimates of lineage diversification and trait evolution under PCMs widely used in the literature. Our results suggest that ignoring budding speciation when age-dependent trait evolution operates might lead to incorrect inferences such as inferring the wrong ancestral state for younger nodes. We also suggest that it might be possible, although challenging, to incorporate budding speciation into PCMs for both discrete and continuous traits. The introduction of budding speciation in comparative approaches, however, depends on the departure from the parsimony paradigm which we suspect is a barrier to the development of macroevolutionary models that can fully integrate external biological information about trait evolution. When we intuitively imagine a parsimonious trait history, we are doing so independently of what is known about the evolutionary history of the system. Reflecting on the role of comparative approaches and recognizing their limitations, especially when testing scenarios of complex trait evolution, is key to the development of alternative models that help the study of macroevolution to become a more integrative endeavor. The incorporation of budding speciation is one example of the direction we can take in improving comparative studies, and we



481 hope our discussion motivates researchers to explore further some of these possibilities.

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493

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496

497 **Conflict of interest:** The authors have no conflicts of interest to declare.

498

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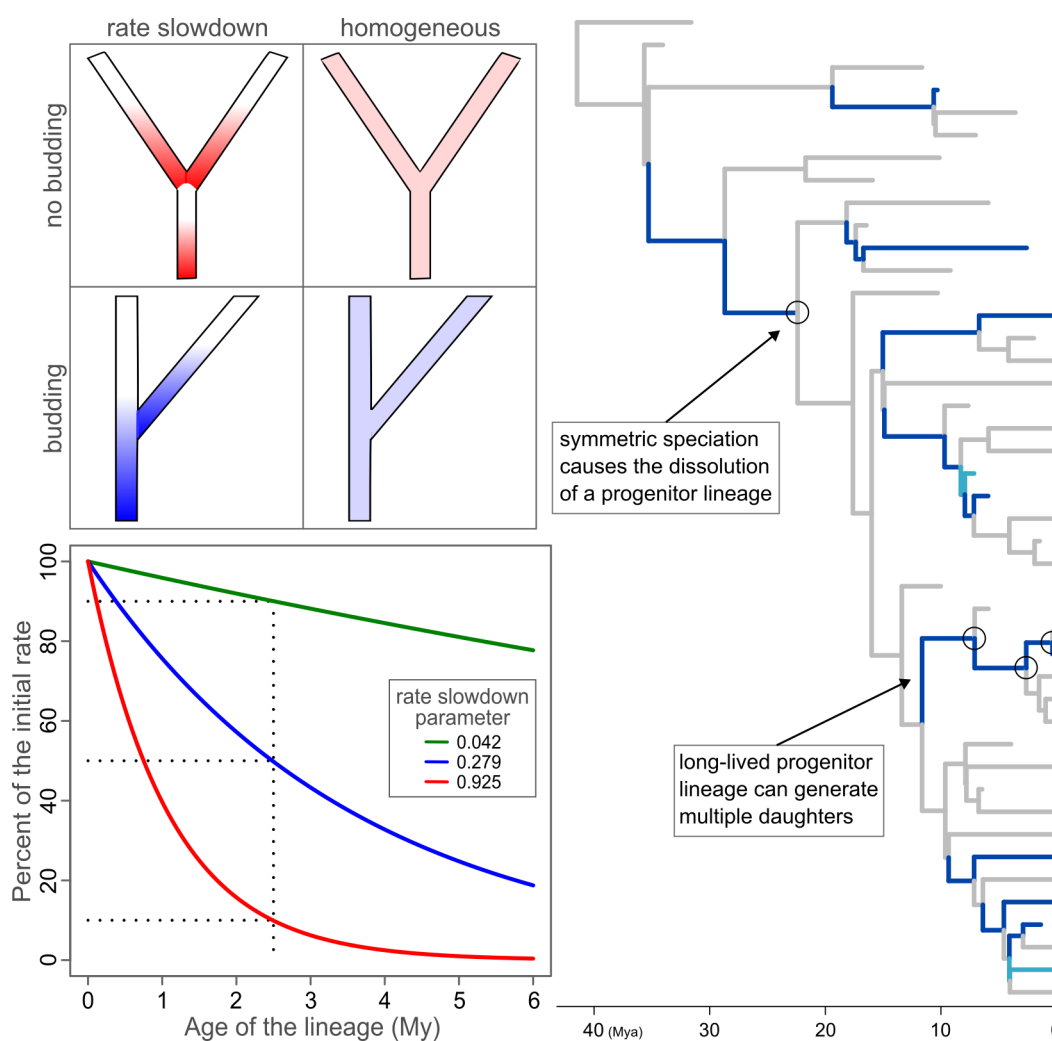
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703 **FIGURES:**

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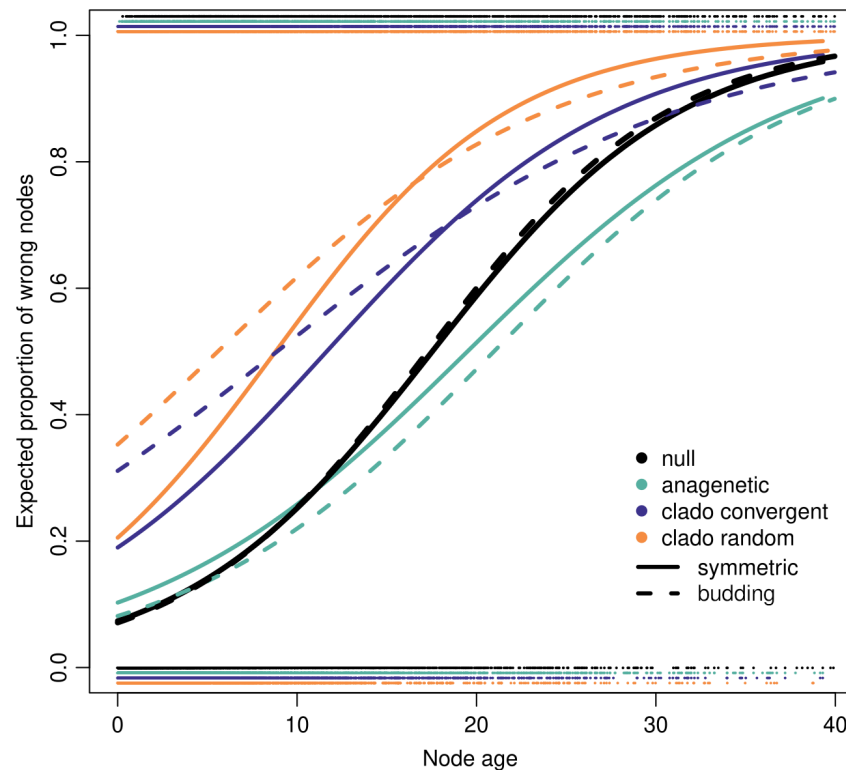
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707 **Figure 1:** Approach used to illustrate and simulate budding speciation and trait evolution under

708 lineage-age dependent rates. Top-left: Interaction between age-dependent trait evolution and  
709 speciation with and without budding. Stronger shades of color indicate faster rates of trait change.  
710 Bottom-left: Rate slowdown as a function of lineage age under the three treatments used. Right:  
711 Example of budding history with 20 extant species showing progenitor lineages in light and dark  
712 blue.  
713

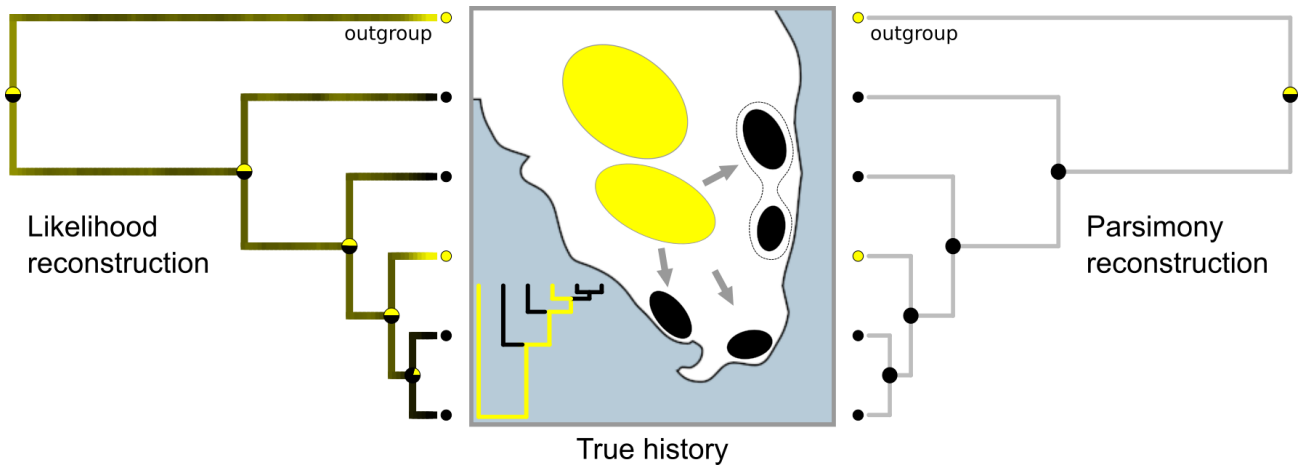


714  
715 **Figure 2:** Conflict between ancestral state estimation and the true history of lineage evolution under  
716 budding. New species appeared through budding and converged three times into a different niche,  
717 associated with the coastal distribution (black). The progenitor lineage is the sister to the outgroup  
718 and carries the inland distribution (yellow). Neither likelihood nor parsimony recovered the true  
719 history of the lineages. Left: Marginal ancestral state estimation for the best fit Markov model.  
720 Branches are painted following a posterior distribution of 100 stochastic maps. Right: Most  
721 parsimonious ancestral estimation following Sankoff's (1975) algorithm. Center: True history of the  
722 trait. Phylogeny in the bottom left shows the continuation of the progenitor lineage through multiple  
723 budding events.





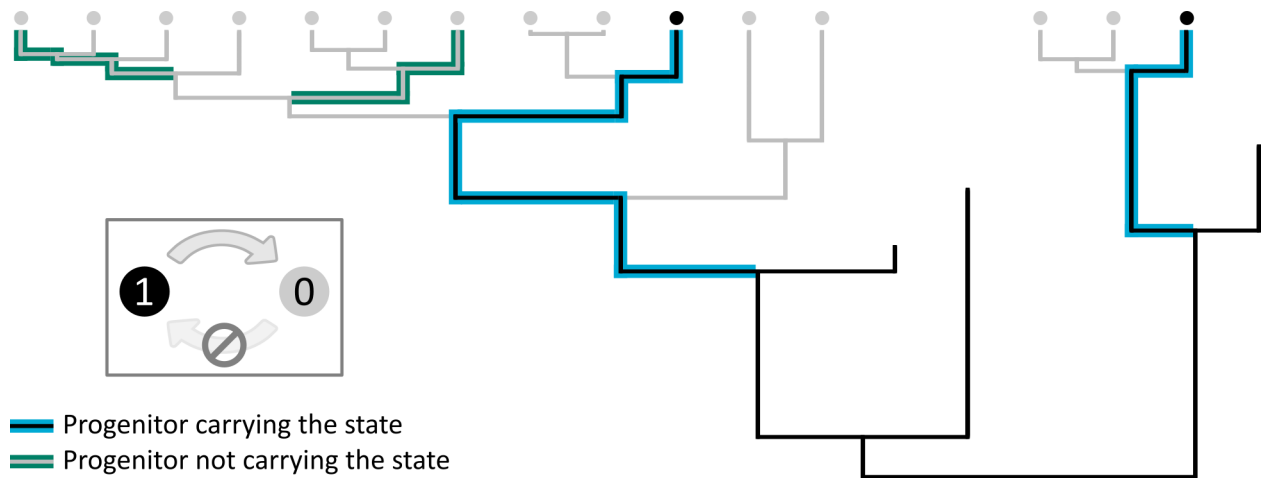
725



726

727 **Figure 3:** Evolution of a truly irreversible trait. Long-lived progenitor lineages carry the ancestral  
 728 state whereas daughter lineages have lost the trait independently several times. Fossil lineages show  
 729 evidence of trait homology. PCMs in the absence of the fossil record would wrongly estimate two  
 730 independent origins of the "black" trait.

731



**Figure 4:** Relationship between the presence of nodes with wrongly reconstructed states and node age. Nodes were considered wrongly reconstructed if the marginal ancestral state probability for any state distinct from the correct state were  $> 0.5$  at that node. Logistic regressions were performed with the pooled results from the 50 simulation replicates per group and independent for each type of node and study group (see Table S1). Dashed lines represent nodes with budding speciation whereas solid lines show nodes without budding. Dots on the top and bottom represent pooled nodes across simulation replicates correctly and incorrectly estimated, respectively.