


Transmission potential of the entomopathogenic fungi *Isaria fumosorosea* and *Beauveria bassiana* from sporulated cadavers of *Diaphorina citri* and *Toxoptera citricida* to uninfected *D. citri* adults

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Abstract *Diaphorina citri* Kuwayama (Hemiptera: Liviidae) is the vector of the causal agent of Huanglongbing disease, and *Toxoptera citricida* Kirkaldy (Hemiptera: Aphididae) is the vector of the citrus tristeza virus. *D. citri* and *T. citricida* share the same habitat and both are susceptible to *Beauveria bassiana* and *Isaria fumosorosea*. Therefore, the aim of this study was to evaluate the horizontal transmission of *B. bassiana* ESALQ-PL63 and *I. fumosorosea* ESALQ-1296 between *D. citri* cadavers and uninfected *D. citri*

adults, as well as between *T. citricida* cadavers and uninfected *D. citri* adults under laboratory and semi-field conditions. In the laboratory, the presence of one to ten *D. citri* cadavers infected with *B. bassiana* and *I. fumosorosea* in citrus plants resulted in mortality rates of *D. citri* adults ranging from 51.2 to 81.9 % and 36.2 to 68 %, respectively. When *T. citricida* cadavers were used, the mortality rates of uninfected *D. citri* adults ranged from 35.4 to 87.7 % with *B. bassiana* and from 41.7 to 80.4 % with *I. fumosorosea*. The horizontal transmission was also confirmed under semi-field conditions. The results indicate that the control of *D. citri* using *I. fumosorosea* and *B. bassiana* can be promoted through conidial cycling in *T. citricida*, by generating new infection cycles, which may contribute to the overall performance of microbial control in citrus.

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Introduction

Direct and indirect damage caused by pests are important factors that result in citrus yield losses. *Diaphorina citri* Kuwayama (Hemiptera: Liviidae) and *Toxoptera citricida* Kirkaldy (Hemiptera: Aphididae) are vectors of two important diseases: citrus greening, or huanglongbing (HLB) disease, and citrus

tristeza (CTV), respectively (Bové 2006; Michaud 1998). HLB is caused by the bacteria ‘*Candidatus Liberibacter spp.*’, which are transmitted by *D. citri* and affect citrus production in India, China, Indonesia, South Africa and Brazil (Kuchment 2013). These pests are usually controlled through intensive applications of agrochemicals, which may result in side effects such as target pest resurgence, secondary pest outbreaks and phytosanitary product resistance (Gravena 2005).

Entomopathogenic fungi are important natural enemies of pests because they infect a large number of arthropods, and under favorable conditions, they can initiate epizootics in the field (Alves 1998). The pathogenicity of certain fungal species to *D. citri* has been reported, such as *Isaria fumosorosea*, *Beauveria bassiana*, *Hirsutella citriformis*, *Lecanicillium lecanii*, *L. muscarium*, *L. longisporum*, *Metarhizium anisopliae* and *M. brunneum* (Ascomycota: Hypocreales) (Avery et al. 2009; Casique-Valdes et al. 2011; Gandarilla-Pacheco et al. 2013; Hall et al. 2012; Hoy et al. 2010; Hunter et al. 2011; Lezama-Gutiérrez et al. 2012; Meyer et al. 2007, 2008; Padulla and Alves 2009; Pinto et al. 2012; Stauderman et al. 2012; Subandiyah et al. 2000; Yang et al. 2006). The entomopathogenic fungi *I. fumosorosea*, *B. bassiana*, *L. lecanii* and *M. anisopliae* have been reported as pathogens of *T. citricida* (Balfour and Khan 2012; Hernández-Torres et al. 2006; Pick et al. 2012; Poprawski et al. 1999).

The dissemination and transmission of entomopathogenic fungi are important factors for determining their potential as microbial control agents of pests in agroecosystems. An important characteristic of entomopathogenic fungi is the dissemination of conidia in the environment due to both abiotic (wind, rain or dew) and biotic (primary or secondary host, parasites, predators, birds and mammals) factors (Hesketh et al. 2010). The conidia present on an insect can be transferred to the same or different species of insects by horizontal transmission (Harper 1987), and they can develop new infection cycles and natural epizootics in the field (Hajek and St. Leger 1994).

Several studies have confirmed the horizontal transmission of the entomopathogenic fungi *B. bassiana*, *M. anisopliae* and *I. fumosorosea* in agricultural pests through the transmission of conidia between sporulating and uninfected individuals

(Avery et al. 2010; Long et al. 2000; Lopes et al. 2011; Sanjaya et al. 2013; Svedese et al. 2013) as well as between females and males of the same species during copulation (García-Munguía et al. 2011; Quesada-Moraga et al. 2008; Toledo et al. 2007). Avery et al. (2009) demonstrated in laboratory tests that 83 % of *D. citri* adults exposed to yellow cards sprayed with *I. fumosorosea* blastopores became infected and cadavers developed conidia externally. Furthermore, Moran et al. (2011) showed that inoculation of *I. fumosorosea* blastopores on yellow cards resulted in less than 40 % of *D. citri* infected after three weeks of confinement in plastic boxes under semi-field conditions. However, the transmission and dissemination potential of entomopathogenic fungi from sporulating cadavers without introduction of artificial inoculation devices have not been reported in citrus pests.

The aphid *T. citricida* shares the same habitat of young leaves and citrus shoots with *D. citri* (Gravena 2005). Because of its larger size and generally large population densities relative to *D. citri*, the host *T. citricida* may serve as an important source of inoculum in the transmission of *B. bassiana* and *I. fumosorosea* to *D. citri*. Therefore, the objective of the present study was to estimate the horizontal transmission of *I. fumosorosea* (ESALQ-1296) and *B. bassiana* (ESALQ-PL63) between sporulating cadavers and uninfected adult *D. citri* (intraspecific transmission) as well as between sporulating *T. citricida* cadavers and uninfected adult *D. citri* (interspecific transmission) under laboratory and semi-field conditions. *I. fumosorosea* ESALQ-1296 and *B. bassiana* ESALQ-PL63 were selected since they were shown to cause high levels of mortality and sporulation in *D. citri* and *T. citricida* (unpublished data). Also, *B. bassiana* ESALQ-PL63 is registered under the trade name Boveril® (Koppert Brazil), and *I. fumosorosea* ESALQ-1296 is under registration in Brazil.

Materials and methods

Diaphorina citri and *Toxoptera citricida* rearing

Diaphorina citri was reared on *Murraya paniculata* (Rutaceae) seedlings inside cages of 60 × 60 × 50 cm (height, width and depth) with a steel framework and walls covered with voile fabric. *T. citricida*

was reared on *Citrus sinensis* (Rutaceae) plants, and the insects were confined in bags made of voile fabric. Both insects were maintained in a climate-controlled room at $25 \pm 2^\circ\text{C}$, 70–80 % RH and 14 h photophase.

Fungal cultivation

The entomopathogenic fungi *I. fumosorosea* ESALQ-1296 and *B. bassiana* ESALQ-PL63 were obtained from the collection of entomopathogens of the Laboratory of Pathology and Microbial Control of Insects of the “Luiz de Queiroz” College of Agriculture (ESALQ), University of São Paulo (USP), which is located in Piracicaba, São Paulo State, Brazil. The ESALQ-1296 strain was obtained from adults of the whitefly *Bemisia tabaci* biotype B (Hemiptera: Aleyrodidae) collected in Jaboticabal, São Paulo, in 2001. The ESALQ-PL63 strain was isolated from *Atta* sp. (Hymenoptera: Formicidae) collected in Piracicaba, São Paulo, in 1992. The fungi were cultivated in 9 cm-diameter Petri dishes containing Sabouraud dextrose agar medium supplemented with yeast extract (SDYA: 2.5 g l⁻¹ bacteriological peptone, 10 g l⁻¹ dextrose, 2.5 g l⁻¹ yeast extract and 20 g l⁻¹ bacteriological agar) and maintained in an incubator at $26 \pm 1^\circ\text{C}$ with a 12 h photophase for ten days.

Obtaining sporulated cadavers

Inocula of *B. bassiana* (ESALQ-PL63) and *I. fumosorosea* (ESALQ-1296) were produced on Petri dishes containing SDYA and conidia were harvested from the surface by scraping with a metallic spatula. Fungal suspensions at a concentration of 5×10^8 - conidia ml⁻¹ were prepared with Tween 80 (0.01 %) and homogenized with a Vortex® apparatus. Two hundred adults of *D. citri* or *T. citricida* were placed on young shoots of orange plants (1 m height) of the variety “Pera” (20 adults per plant). The plants were sprayed with 400 µl of the conidial suspension of each fungus (five plants per fungus). Spraying was performed with an airbrush-type hand sprayer (SAGYMA®, model SW-168) at 10 PSI pressure. The sprayed plants were maintained in a climate-controlled room at $25 \pm 2^\circ\text{C}$, 70–80 % RH and 12 h photophase. At five days post-application, the dead insects were transferred to 24-well cell culture plates containing cotton moistened with sterile distilled

water (humid chamber) and maintained for three days in an incubator at $26 \pm 1^\circ\text{C}$ with a 12 h photophase to promote sporulation.

Quantification of sporulation

All of the cadavers of *D. citri* or *T. citricida* showed abundant sporulation after pulverization of *I. fumosorosea* ESALQ-1296 and *B. bassiana* ESALQ-PL63. These adults were used to quantify the number of conidia per insect. Five sporulated cadavers of each insect species were chosen at random and placed together in Eppendorf tubes. Then, 1 ml of 0.005 % Tween 80 was added to the tubes, and the material was homogenized with a Vortex® apparatus to facilitate the removal of conidia from the surface of the insects. The total number of conidia was quantified in a Neubauer chamber, and the concentration of conidia per individual was estimated. For each insect species and fungus, four replicates were performed, and each replicate included five sporulated adults ($n = 20$).

Horizontal transmission test under laboratory conditions

The experimental unit consisted of a seedling of *Citrus limonia* measuring 15 cm in height, grown in a pot. The shoot was confined in a 500 ml plastic cup with two openings (45–50 cm²) covered with voile fabric to allow aeration. The cups were maintained in plastic trays to facilitate irrigation. The sporulated *D. citri* or *T. citricida* cadavers were transferred to the plants with the aid of a fine insulin syringe needle, and uninfected adults were transferred with the aid of an aspirator. One, three, five or ten sporulated *D. citri* or *T. citricida* cadavers and 20 uninfected adult *D. citri* were introduced per plant. In the control group, 20 uninfected adult *D. citri* were transferred to the plants without adding any cadavers. All of the treatments were maintained in a climate-controlled room at $25 \pm 2^\circ\text{C}$, 80 \pm 10 % RH and 12 h photophase for ten days. The dead insects were removed daily from the plastic cups to avoid the development of new cycles of infection or sporulation of new cadavers, which would increase the dose of conidia per plant. Two parameters were estimated on the 10th day based on the number of live and dead insects (mortality) and the number of insects that died during the study and sporulated (sporulation). Mortality was measured to

estimate the efficacy of horizontal transmission from sporulated cadavers to uninfected insects, and sporulation was used to confirm that the mortality was caused by the fungal infection and to estimate the potential for new cycles of disease. The experimental design for *D. citri* and *T. citricida* was completely randomized, with at least three replicates per treatment, and all combinations (regarding fungus and cadaver numbers) were run at the same time. The experiment was performed separately for each insect pest and was carried out one time for *T. citricida* and three times on different dates for *D. citri*.

Horizontal transmission test under semi-field conditions

The semi-field assays were performed from March 2013 to April 2013 (*D. citri* → *D. citri*) and April 2013 to March 2014 (*T. citricida* → *D. citri*) (Table 1) at the ESALQ-USP Campus, Piracicaba, São Paulo, Brazil. Orange plants (*Citrus sinensis*) of the variety 'Natal' measuring approximately 55 cm in height were placed on a white plastic tray (43 length × 43 width × 12 depth cm) and covered with a cage constructed of wire and voile fabric (62 × 31 × 31 cm). In this assay, the transmission of the fungi *I. fumosorosea* ESALQ-1296 and *B. bassiana* ESALQ-PL63 from *T. citricida* cadavers and from *D. citri* cadavers to uninfected *D. citri* adults was examined using the same number of insects as the laboratory assays (1:20, 3:20, 5:20 and 10:20

sporulated:uninfected). The evaluations were performed on the 10th day by counting the number of dead insects to calculate the mortality of *D. citri* or *T. citricida* adults. Additionally, the number of infected and adhered insects on the plants was evaluated to estimate the potential of new cadavers for future transmission and dissemination in semi-field conditions. The experimental design was completely randomized with three replicates per treatment, and the experiment was repeated three times. In each experiment, a datalogger (IDEL Manaus HT-4000) was installed inside the cages to record the environmental conditions.

Statistical analysis

Negative binomial generalized linear models were fit to the data on the number of conidia per adult insect (sporulation) with a linear interaction predictor (fungal species × insect species) because the data were overdispersed (Demétrio et al. 2014). When there was a significant difference between treatments, multiple comparisons were performed using the glht function of the multcomp package (Hothorn et al. 2008). The mortality rate and percentage of *D. citri* and *T. citricida* cadavers sporulated with the entomopathogenic fungi were evaluated in the transmission assays performed in the laboratory. In the semi-field assays, the rate of mortality and number of cadavers adhered to the plant were evaluated. As these data consist of proportions and are overdispersed,

Table 1 Environmental conditions inside the cages used in the experiments examining the intraspecific (from *D. citri* to *D. citri*) and interspecific (from *T. citricida* to *D. citri*) horizontal

transmission of entomopathogenic fungi under semi-field conditions in Piracicaba, São Paulo State, Brazil

Treatment	Experiments	Date (month/ day/year)	Mean temp. (min–max) (°C) ^a	Mean RH (min–max) (%) ^b	Rainfall (mm) ^c
From <i>D. citri</i> to <i>D. citri</i>	1	03/15/2013	25.1 (15.4–43.5)	74.7 (34.7–96.8)	15.5
	2	03/27/2013	23.4 (17.0–39.0)	81.6 (40.9–96.7)	129.5
	3	04/12/2013	22.4 (9.8–38.9)	78.4 (34.4–97.2)	35.0
From <i>T. citricida</i> to <i>D. citri</i>	1	04/20/2013	22.1 (10.8–41.4)	74.3 (27.2–96.1)	0
	2	05/03/2013	19.9 (4.7–41.1)	74.3 (25–96.4)	0
	3	03/12/2014	27.78 (18.0–46.7)	70.1 (20.2–98.4)	52.3

^a Temperature (°C) mean (minimum and maximum)

^b RH (%) mean (minimum and maximum)

^c Rainfall during the experimental period (ten days)

quasi-binomial generalized linear models were fit with a logit link, first with different linear predictors per fungal species. Then, nested models with parallel linear predictors, coincident linear predictors and a null model were fit, and the significance of the effects was assessed using analysis of deviance tables. The coincident lines linear predictor was selected, which can be written as $\eta = \beta_0 + \beta_1 x$, where β_0 is the intercept, β_1 is the slope and x is the number of sporulated cadavers. Goodness-of-fit was evaluated using half-normal plots with a simulated envelope (Demétrio et al. 2014). An analysis of deviance was performed to assess the significance of the interaction between the dates and treatments, and if the effect was non-significant ($P > 0.05$), the data from each date could be pooled in the same analysis. All analyses were carried out using the statistical software R (R Development Core Team 2014).

Results

Sporulation of *I. fumosorosea* and *B. bassiana* on *D. citri* and *T. citricida*

The number of conidia produced by the fungi *I. fumosorosea* ESALQ-1296 and *B. bassiana* ESALQ-PL63 on the *D. citri* and *T. citricida* cadavers was evaluated to estimate the amount of inoculum with dispersion potential in the experimental units. The effect of the interaction between fungal species and pest species was significant ($F_{1,12} = 23.86$, $P < 0.001$). The sporulation of *I. fumosorosea* was similar in adult *D. citri* and *T. citricida*. However, *B. bassiana* exhibited greater sporulation in adult *D. citri* than in adult *T. citricida* (Table 2).

Intraspecific horizontal transmission (*D. citri* → *D. citri*) under laboratory conditions

The interaction between the dates and treatments was not significant ($F_{14,48} = 1.302$, $P = 0.241$), and the data from the three dates were therefore pooled in the analysis. There was no difference between the fungal species regarding mortality and sporulation (different vs. parallel linear predictors: $F_{1,68} = 0.044$, $P = 0.834$; parallel vs. coincident linear predictors:

$F_{1,69} = 1.176$, $P = 0.282$ for the mortality data; and different vs. parallel linear predictors: $F_{1,68} = 0.001$, $P = 0.974$; parallel vs. coincident linear predictors: $F_{1,69} = 0.018$, $P = 0.894$ for the sporulation data). Thus, the same regression curve was fit for *B. bassiana* ESALQ-PL63 and *I. fumosorosea* ESALQ-1296. Mortality and sporulation rates increased with the number of introduced cadavers, as indicated by the tests comparing the model with coincident linear predictors and the null model that was significant for both mortality ($F_{1,70} = 15.335$, $P < 0.001$) and sporulation ($F_{1,70} = 14.806$, $P < 0.001$) data. After the introduction of one to ten cadavers sporulated with *I. fumosorosea* and 20 uninfected adult *D. citri*, the mortality rate varied between 36.2 and 68.0 % and sporulation between 34.3 and 67.4 %. However, when one or ten cadavers sporulating with *B. bassiana* were introduced, the mortality rate varied between 51.2 and 81.9 % (Fig. 1a) and sporulation between 45.2 and 75.5 % (Fig. 1b). The treatment in which no cadavers sporulating were introduced (control) exhibited a natural mortality rate of 12.0 % (Fig. 1a), and these insects did not present fungal infections.

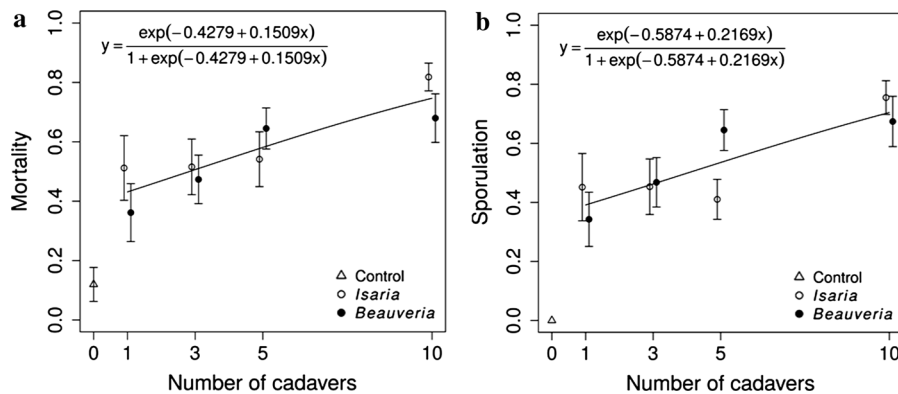
Interspecific horizontal transmission (*T. citricida* → *D. citri*) under laboratory conditions

The two fungal species caused similar mortality and sporulation (different vs. parallel linear predictors: $F_{1,20} = 0.360$, $P = 0.555$; parallel vs. coincident linear predictors: $F_{1,21} = 1.258$, $P = 0.4275$ for the mortality data; and different vs. parallel linear predictors: $F_{1,20} = 1.746$, $P = 0.201$; parallel vs. coincident linear predictors: $F_{1,21} = 2.542$, $P = 0.127$ for the sporulation data). Thus, the same regression curve was fit for *B. bassiana* and *I. fumosorosea*. The tests comparing the model with coincident linear predictors and the null model were significant for both mortality ($F_{1,22} = 19.793$, $P < 0.001$) and sporulation ($F_{1,22} = 4.542$, $P < 0.046$), indicating that mortality and sporulation rates increased with the number of sporulated cadavers (Fig. 2). The *B. bassiana* ESALQ-PL63 isolate caused 35.4 to 87.7 % mortality at ratios of 1:20 to 10:20 (sporulated:uninfected), respectively. The mortality rates observed for *I. fumosorosea* ESALQ-1296 ranged from 41.7 to 80.4 % at ratios of 1:20 to 10:20 (sporulated:uninfected), respectively (Fig. 2a). The percentage of dead

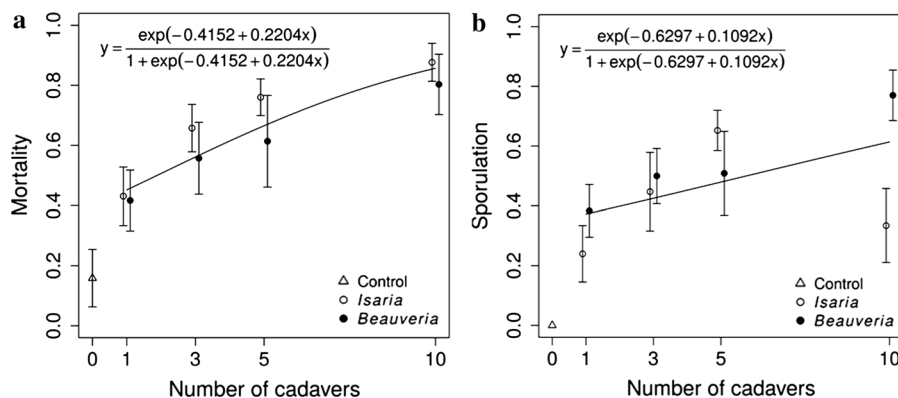
Table 2 Number of conidia produced by *Isaria fumosorosea* ESALQ-1296 and *Beauveria bassiana* ESALQ-PL63 in sporulating cadavers of *Diaphorina citri* and *Toxoptera citricida*

Fungal Species	Means of Conidia number ($\times 10^6$) per cadaver \pm SE ^a	
	<i>Diaphorina citri</i>	<i>Toxoptera citricida</i>
<i>Beauveria bassiana</i> ESALQ-PL63	4.14 \pm 0.44aA	1.06 \pm 0.03bB
<i>Isaria fumosorosea</i> ESALQ-1296	3.02 \pm 0.22aA	3.29 \pm 0.30aA

^a Lower-case letters represent differences within a column (fungi), and upper-case letters represent differences within rows (insect) by multiple comparisons test ($P < 0.05$)

**Fig. 1** Model fit for mortality (a) and sporulation (b) of adult *Diaphorina citri* after ten days of the introduction of different numbers of *Diaphorina citri* cadavers with conidia of *Isaria fumosorosea* and *Beauveria bassiana* at temperature of

25 \pm 2 °C, 80 \pm 10 % RH and 12 h photophase. The same regression curve was fitted for both fungal species. Plotted values represent means while bars represent SE

**Fig. 2** Model fit for mortality (a) and sporulation (b) of adult *Toxoptera citricida* after ten days of the introduction of different numbers of *Toxoptera citricida* cadavers with conidia of *Isaria fumosorosea* and *Beauveria bassiana* at temperature of

25 \pm 2 °C, 80 \pm 10 % RH and 12 h photophase. The same regression curve was fitted for both fungal species. Plotted values represent means while bars represent SE

insects with *I. fumosorosea* and *B. bassiana* spores ranged from 38.3 to 77.0 % and 14.5 to 65.3 %, respectively (Fig. 2b). The treatment in which no

sporulated cadavers were introduced (control) exhibited a natural mortality rate of 15.8 % (Fig. 2a), and these insects did not present fungal infections.

Intraspecific horizontal transmission (*D. citri* → *D. citri*) under semi-field conditions

The analysis of deviance demonstrated no significant interaction between the assays and treatments ($F_{14,48} = 1.37$, $P = 0.21$), and the assays showed the same trend, so only the last one was presented. Similar to the laboratory experiment, no differences were observed in horizontal transmission between the fungal species. Thus, the same regression curve was fit for both *B. bassiana* ESALQ-PL63 and *I. fumosorosea* ESALQ-1296 (different vs. parallel linear predictors: $F_{1,20} = 0.086$, $P = 0.773$; parallel vs. coincident linear predictors: $F_{1,21} = 1.005$, $P = 0.328$). However, the test between the model with coincident linear predictors and the null model was non-significant ($F_{1,22} = 4.317$, $P = 0.051$), so the mortality rate was not influenced by the number of cadavers under semi-field conditions. The mortality rates ranged from 26.7 to 58.3 % for *B. bassiana* and 16.7 to 56.7 % for *I. fumosorosea* (Fig. 3). Although greater variance in the mortality rates within and between replicates was observed, an increase in mortality related to increases in the number of cadavers was observed, which was similar to the results of the experiments conducted under laboratory conditions. After ten days, relatively low percentages of infected *D. citri* adults remained adhered and sporulated on the plants, and the number of adults did not vary significantly according to the number of sporulated cadavers released. The

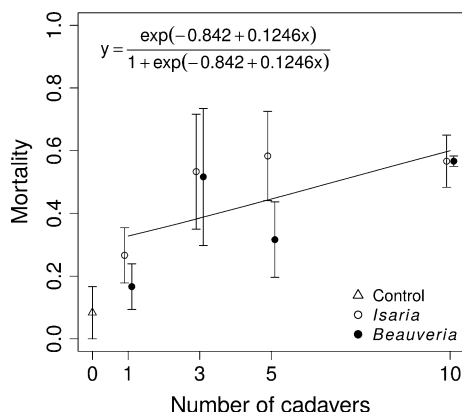


Fig. 3 Model fit for mortality of adult *Diaphorina citri* after ten days of the introduction of different numbers of *D. citri* cadavers with conidia of *Isaria fumosorosea* and *Beauveria bassiana* in semi-field conditions. The same regression curve was fitted for both fungal species. Plotted values represent means while bars represent SE

percentage of dead insects and insects adhered to the plants did not vary significantly as the number of sporulated cadavers released increased ($F_{7,16} = 1.69$, $P = 0.18$). Only 1.7, 3.3, 8.3 and 20.0 % of the dead insects remained adhered to the plant when the fungus *B. bassiana* was used at ratios of 1:20, 3:20, 5:20 and 10:20 (sporulated:uninfected), respectively. However, 5.0, 11.7, 11.7 and 18.3 % of the dead insects remained adhered to the plant when the fungus *I. fumosorosea* was used at ratios of 1:20, 3:20, 5:20 and 10:20 (sporulated:uninfected), respectively. The environmental conditions were characterized by large temperature (9.8–39 °C), RH (34.4–97.2 %) and rainfall ranges (15.5–129.5 mm) during the experimental period (Table 1).

Interspecific horizontal transmission (*T. citricida* → *D. citri*) under semi-field conditions

The analysis of deviance showed no significant interaction between the assays and treatments ($F_{7,32} = 1.25$, $P = 0.31$), and the assays showed the same trend, so only the last one was presented. The introduction of *T. citricida* cadavers sporulated with *I. fumosorosea* ESALQ-1296 and *B. bassiana* ESALQ-PL63 resulted in mortality rates of *D. citri* that ranged from 31.7 to 53.3 % and 15.0 to 38.3 %, respectively (Fig. 4). No differences in horizontal transmission were observed between the fungal species. Thus, the same regression curve was fitted for both *B. bassiana* and *I. fumosorosea* (different vs. parallel linear predictors: $F_{1,20} = 0.113$, $P = 0.741$; parallel vs. coincident linear predictors: $F_{1,21} = 0.654$, $P = 0.428$). Mortality rates were not influenced by the number of cadavers under semi-field conditions, as indicated by the non-significant tests comparing the model with coincident linear predictors and the null model ($F_{1,22} = 0.286$, $P = 0.599$). Although considerable mortality levels caused by the transmission of the entomopathogenic fungi were observed, no significant differences were observed between the treatments. In this experiment, less than 5.0 % of the dead adult *D. citri* were observed adhering to the plants. The environmental conditions were characterized by wide ranges of temperature (4.7–46.7 °C), RH (20.2–98.4 %) and rainfall (0–52.3 mm) during the experimental period (Table 1).

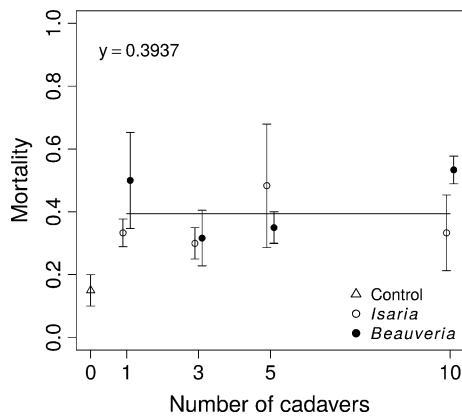


Fig. 4 Model fit for mortality of adult *Diaphorina citri* after ten days of the introduction of different numbers of *Toxoptera citricida* cadavers with conidia of *Isaria fumosorosea* and *Beauveria bassiana* in semi-field conditions. The same regression curve was fitted for both fungal species. All other parameters but the intercept were not significant (test for different linear predictors vs. null model: $F_{3,20} = 0.35$, $P = 0.79$). Plotted values represent means while bars represent SE

Discussion

The two fungal isolates used in the current study, *I. fumosorosea* ESALQ-1296 and *B. bassiana* ESALQ-PL63, were selected based on high mortality (62.8 and 59.7 %, respectively) and sporulation (41.1 and 37.9.3 %, respectively) in adult *D. citri* ten days after treatment with a suspension of 5×10^6 conidia ml^{-1} (unpublished data). Additionally, it was demonstrated that *I. fumosorosea* ESALQ-1296 caused higher mortality (65.4 %) and sporulation (64.2 %) than *B. bassiana* ESALQ-PL63 (mortality of 42.7 % and sporulation of 26 %) in *T. citricida* adults ten days after treatment with a suspension of 5×10^6 conidia ml^{-1} (unpublished data). Therefore, biological control of citrus pests can be enhanced by transmission and dissemination of *B. bassiana* and *I. fumosorosea* between different insect species that share the same habitat, such as *D. citri* and *T. citricida*.

The sporulation of entomopathogenic fungi on a host is an important characteristic because conidia serve as a source of pathogen inocula for horizontal transmission, ecosystem dissemination and infection cycle development (Hesketh et al. 2010). The number of conidia produced by *I. fumosorosea* ESALQ-1296 was similar to the number produced by *B. bassiana* ESALQ-PL63 in *D. citri* cadavers. However,

sporulation in *T. citricida* cadavers was three times higher for the ESALQ-1296 isolate compared with the ESALQ-PL63 isolate. The production of conidia in cadavers is known to vary according to the fungal species, host species, insect size and developmental stage (Sosa-Gómez and Alves 2000; Steinkraus 2006).

The results of the present study demonstrated that considerable horizontal transmission of the fungi *I. fumosorosea* ESALQ-1296 and *B. bassiana* ESALQ-PL63 occurs between *D. citri* individuals (intraspecific). The horizontal transmission potential of both entomopathogenic fungi between *D. citri* individuals was similar, which may be associated with the similar average production of conidia in the cadavers quantified in the study described herein. The increase in the mortality of adult *D. citri* was correlated with the increase in the inoculum source (number of sporulated insects per plant), which may have resulted in greater acquisition and dissemination of the conidia at the site. Using a different method from that used in the present study, Moran et al. (2011) demonstrated the transmission rates of *I. fumosorosea* blastospores impregnated on yellow card between infected and uninfected *D. citri* (1:1.5 treated:healthy ratio) were approximately 25 % after three week, which were lower than rates obtained in the present study when using ten *D. citri* cadavers and 20 healthy adults (1:2 ratio). The horizontal transmission of *B. bassiana* has been shown to be affected by the amount of conidia, density of cadavers, host development stage and the environmental conditions in other agricultural pests (Long et al. 2000).

The present study demonstrated and quantified the potential for interspecific horizontal transmission of the entomopathogenic fungi *B. bassiana* ESALQ-PL63 and *I. fumosorosea* ESALQ-1296 between *T. citricida* cadavers and healthy adult *D. citri*. Although the sporulation of *I. fumosorosea* on *T. citricida* cadavers was greater than that of the *B. bassiana* isolate, the interspecific horizontal transmission observed under laboratory conditions was similar to the intraspecific transmission of both entomopathogenic fungi. This may be associated with factors such as fungal virulence and dissemination potential or host susceptibility.

HLB disease has increased significantly in citrus orchards in recent years. The acceptable level of the HLB insect vector, *D. citri*, is extremely low (near zero). Therefore, insecticides are usually used to control this vector, which has resulted in increased

production costs (FNP 2013). In Brazil, many farmers have ceased control of the vector in older groves, organic groves, and residential groves. The population levels of the aphid *T. citricida*, a vector of CTV, are usually higher than those of *D. citri* because of the use of rootstock resistant to CTV as well as varieties pre-immunized with mild strains of CTV (Meissner Filho et al. 2002). Both insects prefer leaves and green shoot branches (Gravena 2005), and the efficiency of *D. citri* and *T. citricida* control by the entomopathogenic fungi *I. fumosorosea* and *B. bassiana* in citrus groves may therefore be improved due to both intra- and inter-specific horizontal transmission.

Horizontal transmission under semi-field conditions resulted in lower mortality rates of adult *D. citri* compared with the laboratory assays, which may have been caused by the larger size of the experimental units used in the field and environmental conditions and the associated greater temperatures, humidity range, wind and rainfall. Baverstock et al. (2010) indicated that a positive relationship occurs between insect movement and horizontal transmission of entomopathogenic fungi. Therefore, the smaller size of the containers used in the laboratory assays could have increased the direct contact between the insects and cadavers with conidia. The density of individuals may have influenced the transmission and dissemination of conidia. Although wind is an important factor in conidial dissemination, it may have facilitated the dissemination of conidia out of the cages. Despite the occurrence of high rainfall after the release of cadavers into the experimental units, which may have removed the cadavers from the leaves, considerable levels of mortality caused by the fungi were still observed under semi-field conditions.

Diaphorina citri nymphs as well as *T. citricida* wingless adults and nymphs are gregarious (Tsai and Liu 2000; Toledo et al. 2006), and this behavior may favor the transfer of conidia between individuals and increase the efficiency of entomopathogenic fungi in controlling *D. citri* in citrus orchards. The level of horizontal transmission of the entomopathogenic fungi *I. fumosorosea* and *B. bassiana* observed in the present study under laboratory and semi-field conditions depended on the level of sporulation and host density in the field. Sporulation on cadavers is associated with periods of high RH, which occur mostly from November to March in São Paulo, Brazil (Italo Delalibera Junior unpublished data). If other

citrus pest species are affected by the same entomopathogenic fungi and their cadavers exhibit sporulation in the field, the control of such pests could be improved. In abandoned and residential areas, *D. citri* and *T. citricida* densities are high, and after spray application of *I. fumosorosea* and *B. bassiana*, these insects could exhibit sporulation and horizontal transmission, which might increase *D. citri* control in citrus groves. Farmers who carry out intensive management of *D. citri* in Brazil are controlling the pest even in neighboring areas where *D. citri* control has been abandoned, to prevent migration into their groves. We anticipate greater adoption of biological control in these less intensively managed areas, where the host populations are usually greater, which may improve the efficacy of fungal applications for controlling *D. citri* populations in the field. Other factors, such as removal of cadavers by ants, may decrease transmission in the field (Nielsen et al. 2010). In the semi-field experiments, we observed that a low percentage of the dead adult *D. citri* were adhered to the plants, and ants of the genera *Dorymyrmex* and *Brachymyrmex* were observed removing dead insects. For these reasons, long-term field studies are needed to confirm the results obtained here under laboratory and semi-field conditions.

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References

- Alves SB (1998) Controle Microbiano de Insetos, 2nd edn. FEALQ, Piracicaba
- Avery PB, Hunter WB, Hall DG, Jackson MA, Powell CA, Rogers ME (2009) *Diaphorina citri* (Hemiptera: Psyllidae) infection and dissemination of the entomopathogenic fungus *Isaria fumosorosea* (Hypocreales: Cordycipitaceae) under laboratory conditions. Fla Entomol 92:608–618
- Avery PB, Queeley GL, Faull J, Simmonds MSJ (2010) Effect of photoperiod and host distribution on the horizontal transmission of *Isaria fumosorosea* (Hypocreales: Cordycipitaceae) in greenhouse whitefly assessed using a novel model bioassay. Biocontrol Sci Technol 20(10):1097–1111

- Balfour A, Khan A (2012) Effects of *Verticillium lecanii* (Zimm.) Viegas on *Toxoptera citricida* Kirkaldy (Homoptera: Aphididae) and its parasitoid *Lysiphlebus testaceipes* Cresson (Hymenoptera: Braconidae). *Plant Prot Sci* 48(3):123–130
- Baverstock J, Roy HE, Pell JK (2010) Entomopathogenic fungi and insect behavior: from unsuspecting hosts to targeted vectors. *BioControl* 55:89–102
- Bové JM (2006) Huanglongbing: a destructive, newly-emerging, century-old disease of citrus. *J Plant Pathol* 88:7–37
- Casique-Valdes R, Reyes-Martinez Y, Sanchez-Peña SR, Bidochka MJ, Lopez-Arroyo JI (2011) Pathogenicity of *Hirsutella citriformis* (Ascomycota: Cordicipitaceae) to *Diaphorina citri* (Hemiptera: Psyllidae) and *Bactericera cockerelli* (Hemiptera: Trioziidae). *Fla Entomol* 94(3):703–705
- FNP Consultoria & Comércio (2013) Tomada de decisão em laranjais pede cautela. In: AGRANUAL 2013: Anuário da Agricultura Brasileira. São Paulo, pp 249–250
- Demétrio CGB, Hinde J, Moral RA (2014) Models for overdispersed data in entomology. In: Godoy WAC, Ferreira CP (eds) Ecological modelling applied to entomology. Springer, Switzerland, pp 219–259
- Gandarilla-Pacheco FL, Gálan-Wong LJ, Arroyo JI, Rodríguez-Guerra R, Quintero-Zapata I (2013) Optimization of pathogenicity tests for selection of native isolates of entomopathogenic fungi isolated from citrus growing areas of México on adults of *Diaphorina citri* Kuwayama (Hemiptera: Liviidae). *Fla Entomol* 96(1):187–195
- García-Munguía AM, Garza-Hernández JA, Rebollar-Tellez EA, Rodríguez-Pérez MA, Reyes-Villanueva F (2011) Transmission of *Beauveria bassiana* from male to female *Aedes aegypti* mosquitoes. *Parasit Vectors* 4:24–29
- Gravena S (2005) Manual prático de manejo ecológico de pragas dos citros. In: Gravena S (ed) As pragas. Jaboticabal, São Paulo, pp 69–220
- Hajek AE, St. Leger RJ (1994) Interactions between fungal pathogens and insects hosts. *Annu Rev Entomol* 39:293–322
- Hall DG, Hentz MG, Meyer JM, Kriss AB, Gottwald TR, Boucias DG (2012) Observations on the entomopathogenic fungus *Hirsutella citriformis* attacking adult *Diaphorina citri* (Hemiptera: Psyllidae) in a managed citrus grove. *BioControl* 57:663–675
- Harper JD (1987) Applied epizootiology: microbial control of insects. In: Fuxa JR, Tanada Y (eds) Epizootiology of insect diseases. Wiley, New York, pp 473–496
- Hernández-Torres I, López-Arroyo JI, Berlanga-Padilla A, Loera-Gallardo J, Acosta-Díaz E (2006) Efectividad de hongos entomopatógenos y vehículos de aplicación para el control del pulgón café de los cítricos *Toxoptera citricida* Kirkaldy (Homoptera: Aphididae). *Vedalia* 13(1):17–26
- Hesketh H, Roy HE, Eilenberg J, Pell JK, Hails RS (2010) Challenges in modelling complexity of fungal entomopathogens in semi-natural populations of insects. *BioControl* 55:55–73
- Hothorn T, Bretz F, Westfall P (2008) Simultaneous inference in general parametric models. *Biom J* 50:346–363
- Hoy MJ, Singh R, Rogers ME (2010) Evaluations of a novel isolate of *Isaria fumosorosea* for control of the Asian citrus psyllid, *Diaphorina citri* (Hemiptera: Psyllidae). *Fla Entomol* 93:24–32
- Hunter WB, Avery PB, Pick D, Powell CA (2011) Broad spectrum potential of *Isaria fumosorosea* against insect pests of citrus. *Fla Entomol* 94(4):1051–1054
- Kuchment A (2013) The end of orange juice. *Sci Am* 308:52–59
- Lezama-Gutiérrez R, Molina-Ochoa J, Chávez-Flores O, Ángel-Sahagún CA, Skoda SR, Reyes-Martínez G, Barba-Reynoso M, Rebollo-Domínguez O, Ruíz-Aguilar GML, Foster JE (2012) Use of the entomopathogenic fungi *Metarhizium anisopliae*, *Cordyceps bassiana* and *Isaria fumosorosea* to control *Diaphorina citri* (Hemiptera: Psyllidae) in Persian lime under field conditions. *Int J Trop Insect Sci* 32(1):39–44
- Long DW, Groden E, Drummond FA (2000) Horizontal transmission of *Beauveria bassiana* (Bals.) Vuill. *Agric For Entomol* 2:19–32
- Lopes RB, Micherreff-Filho M, Tigano MS, Neves PMOJ, López EL, Fancelli M, Silva JP (2011) Virulence and horizontal transmission of selected Brazilian strains of *Beauveria bassiana* against *Cosmopolites sordidus* under laboratory conditions. *Bull Insectol* 64(2):201–208
- Meissner Filho JO, Soares Filho WS, Velame KVC, Diamantino EP, Diamantino MSAS (2002) Reação de Porta enxerto híbridos ao *Citrus Tristeza Virus*. *Fitopatol Bras* 27:312–315
- Meyer JM, Hoy MA, Boucias DG (2007) Morphological and molecular characterization of a *Hirsutella* species infecting the Asian citrus psyllid, *Diaphorina citri* Kuwayama (Hemiptera: Psyllidae) in Florida. *J Invert Pathol* 95:101–109
- Meyer JM, Hoy MA, Boucias DG (2008) Isolation and characterization of an *Isaria fumosorosea* isolate infecting the Asian citrus psyllid in Florida. *J Invert Pathol* 99:96–102
- Michaud JP (1998) A Review of the literature on *Toxoptera citricida* (Kirkaldy) (Homoptera: Aphididae). *Fla Entomol* 81(1):37–61
- Moran PJ, Patt JM, Cabanillas HE, Adamczyk JL, Jackson MA, Dunlap CA, Hunter WB, Avery PB (2011) Localized autoinoculation and dissemination of *Isaria fumosorosea* for control of the Asian citrus psyllid in south Texas. *Sub Plant Sci* 63:23–35
- Nielsen C, Agrawal AA, Hajek AE (2010) Ants defend aphids against lethal disease. *Biol Lett* 6:205–208
- Padulla LFL, Alves SB (2009) Suscetibilidade de ninfas de *Diaphorina citri* a fungos entomopatogênicos. *Arq Inst Biol* 76:297–302
- Pick DA, Avery PB, Hunter WB, Powell CA, Arthurs S (2012) Effect of *Isaria fumosorosea* (Hymenoptera: Cordycipitaceae) and *Lysiphlebus testaceipes*, (Hymenoptera: Braconidae) on the brown citrus aphid: preliminary assessment of a compatibility study. *Fla Entomol* 95(3):764–766
- Pinto APF, Batista Filho A, Almeida JEM, Wenzel IM (2012) Patogenicidade de *Beauveria bassiana* ao psilídeo *Diaphorina citri* e compatibilidade do fungo com produtos fitossanitários. *Pesqui Agropecu Bras* 47(12):1673–1680
- Poprawski TJ, Parker PE, Tsai JH (1999) Laboratory and field evaluation of hyphomycete insect pathogenic fungi for control of brow citrus aphid (Homoptera: Aphididae). *Environ Entomol* 28:315–321

- Quesada-Moraga E, Martin-Carballo I, Garrido-Jurado I, Santiago-Álvarez C (2008) Horizontal transmission of *Metarhizium anisopliae* among laboratory populations of *Ceratitis capitata* (Wiedemann) (Diptera: Tephritidae). *Biol Control* 47:115–124
- R Development Core Team (2014) R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. ISBN 3-900051-07-0. <http://www.R-project.org/>
- Sanjaya Y, Ocampo VR, Caoili BL (2013) Transmission effect of entomopathogenic fungi on population of *Tetanychus kanzawai* (Kishida) (Tetranychidae: Acarina). *Arthropods* 2(1):36–41
- Sosa-Gómez DR, Alves SB (2000) Temperature and relative humidity requirements for conidiogenesis of *Beauveria bassiana* (Deuteromycetes: Moniliaceae). *An Soc Entomol Bras* 29(3):515–521
- Stauderman K, Avery P, Aristizábal L, Arthurs S (2012) Evaluation of *Isaria fumosorosea* (Hypocreales: Cordycipitaceae) for control of the Asian citrus psyllid, *Diaphorina citri* (Hemiptera: Psyllidae). *Biocontrol Sci Technol* 22(7):747–761
- Steinkraus DC (2006) Factors affecting transmission of fungal pathogens of aphids. *J Invert Pathol* 92:125–131
- Subandiyah S, Nikoh N, Sato H, Wagiman F, Tsuyumu S, Fukatsu T (2000) Isolation and characterization of two entomopathogenic fungi attacking *Diaphorina citri* (Homoptera, Psylloidea) in Indonésia. *Mycoscience* 41:509–513
- Svedese VM, Lima EALA, Figueiredo AL (2013) Horizontal transmission and effect of the temperature in pathogenicity of *Beauveria bassiana* against *Diatraea saccharalis* (Lepidoptera: Crambidae). *Braz Arch Biol Technol* 56(3):413–419
- Toledo FR, Barbosa JC, Yamamoto PT (2006) Distribuição espacial de *Toxoptera citricida* Kirkaldy (Hemiptera: Aphididae) na cultura de citros. *Rev Bras Frutic* 28(2):194–198
- Toledo J, Campos SE, Flores S, Liedo P, Barrera JF, Villaseñor A, Montoya P (2007) Horizontal transmission of *Beauveria bassiana* in *Anastrepha ludens* (Diptera: Tephritidae) under laboratory and field cage conditions. *J Econ Entomol* 100(2):291–297
- Tsai JH, Liu YH (2000) Biology of *Diaphorina citri* (Homoptera: Psyllidae) on four host plants. *J Econ Entomol* 93(6):1721–1725
- Yang Y, Huang M, Andrew G, Beattie C, Xia Y, Ouyang G, Xiong J (2006) Distribution, biology, ecology and control of the psyllid *Diaphorina citri* Kuwayama, a major pest of citrus: a status report for China. *Int J Pest Manag* 52(4):343–352

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