

Review

Advances in Fungal Peptide Vaccines

Leandro B. R. Da Silva ^{1,2}, Carlos P. Taborda ^{1,3}  and Joshua D. Nosanchuk ^{2,*} 

¹ Instituto de Ciencias Biomedicas, Departamento de Microbiologia, Universidade de Sao Paulo, Sao Paulo 05508-000, Brazil; leandrobr87@usp.br (L.B.R.D.S.); taborda@usp.br (C.P.T.)

² Departments of Medicine (Division of Infectious Diseases) and Microbiology and Immunology, Albert Einstein College of Medicine, New York, NY 10461, USA

³ Instituto de Medicina Tropical de Sao Paulo, Laboratorio de Micologia Medica, Departamento de Dermatologia, Faculdade de Medicina, Universidade de Sao Paulo, Sao Paulo 05403-000, Brazil

* Correspondence: josh.nosanchuk@einsteinmed.org; Tel.: +1-718-430-2993

Received: 9 June 2020; Accepted: 22 July 2020; Published: 25 July 2020



Abstract: Vaccination is one of the greatest public health achievements in the past century, protecting and improving the quality of life of the population worldwide. However, a safe and effective vaccine for therapeutic or prophylactic treatment of fungal infections is not yet available. The lack of a vaccine for fungi is a problem of increasing importance as the incidence of diverse species, including *Paracoccidioides*, *Aspergillus*, *Candida*, *Sporothrix*, and *Coccidioides*, has increased in recent decades and new drug-resistant pathogenic fungi are emerging. In fact, our antifungal armamentarium too frequently fails to effectively control or cure mycoses, leading to high rates of mortality and morbidity. With this in mind, many groups are working towards identifying effective and safe vaccines for fungal pathogens, with a particular focus of generating vaccines that will work in individuals with compromised immunity who bear the major burden of infections from these microbes. In this review, we detail advances in the development of vaccines for pathogenic fungi, and highlight new methodologies using immunoproteomic techniques and bioinformatic tools that have led to new vaccine formulations, like peptide-based vaccines.

Keywords: fungal; vaccine; peptides; *paracoccidioides*; *sporothrix*; *aspergillus*; *candida*; *coccidioides*; dendritic cells

1. Introduction

Although preceded by variolation in China as early as the 15th century, the development of standardized vaccines began in the 18th century [1]. From the beginning, vaccine development has been focused on using whole attenuated or inactivated microorganisms or fractions of microbes [1]. Remarkably, these methodologies are still routinely used in modern vaccinology. More recently, with advances in technology and a more detailed understanding of immunology, newer innovative methods are being applied for vaccine development. Our current vaccine armamentarium includes vaccines against diverse lethal viral and bacterial diseases, but there is no vaccine against a fungal disease. The absence of mass market appeal has been presented as the major obstacle in fungal vaccine development [2–6]. However, this situation is changing rapidly as the incidence of invasive mycoses has increased with the rising numbers of individuals with increased risk for fungal disease, including cancer patients receiving chemotherapy, bone marrow transplant recipients, individuals with acquired immune deficiency syndrome (AIDS), individuals treated with immune function inhibitors, and others patients with different types of immunosuppression [7–9]. Additionally, certain fungi, such as *Candida auris*, an emerging multi-drug resistant pathogen, have received significant global media attention. Importantly, systemic mycoses are among the leading causes of death and morbidity in the USA, totaling more than 1.6 million deaths [10,11], with annual costs over \$7.2 billion dollars [12–14].

Hence, the increased incidence of mycoses, the emergence of multi-drug resistant species, and the rising costs associated with combatting these diseases have recently led to significantly greater visibility of fungal diseases worldwide.

The global burden of invasive mycoses is massive. For example, there are at least ~250,000 cases of invasive aspergillosis, 3,000,000 cases of chronic pulmonary aspergillosis, ~223,100 cases of cryptococcal meningitis, ~700,000 cases of invasive candidiasis, ~500,000 cases of *Pneumocystis jirovecii* pneumonia, ~100,000 cases of disseminated histoplasmosis, ~10,000,000 cases of fungal asthma, and ~1,000,000 cases of fungal keratitis annually [15–17]. Why, with the increase in the number of cases and the immense medical need, have more than two centuries of vaccine research failed to produce a single therapeutic or prophylactic vaccine for a mycosis? Vaccines, in addition to preventing lethal diseases, improve people's quality of life [18,19], which is particularly notable for fungal diseases that frequently require protracted durations of antifungal drugs that have diverse toxicities and costs. Therefore, we will briefly discuss general issues in fungal vaccine development and then we will focus on some novel and conceptual advances in the field of peptide vaccines against fungal infections, which may simplify and accelerate the achievement of a safe and effective antifungal vaccine that is effective in both immunocompetent and immunologically suppressed individuals.

2. Fungal Vaccine: Some Challenges

One of the first and most difficult challenges in working with fungal vaccines is the need to determine the target population in which a vaccine is applied, as many invasive mycoses have a predilection for causing disease in immunosuppressed individuals [7–9]. Specific knowledge is required regarding the type of protective response necessary to combat a specific fungus and then there is a need to translate this information into a formulation that remains safe and effective in an immunocompromised host [20,21].

Another obstacle on the pathway to developing vaccines against fungal infection is the complexity of the fungal cell. Fungi are eukaryotic, and pathogenic species have marked differences and similarities with human cells. Fungal cells have a double layer of protection: an inner plasma membrane and an outer cell wall [22]. The plasma membrane is a phospholipid bilayer that may vary in composition, due to the presence of specific fungal sterols in different species. Ergosterol, which is similar to human cholesterol, is particularly important for membrane fluidity and it is essential for viability [23]. The cell wall is generally organized as a scaffold of carbohydrate polymers to which a variety of proteins and other components are added, creating a strong but elastic structure [22]. Although there are diverse variations in polysaccharide composition across species, there are conserved components, such as a core of branched β -1,3-glucan-chitin [24]. Thus, the fact that fungi have preserved compounds in both the cell wall and plasma membrane makes it theoretically possible to develop a universal vaccine, where the presence of a common antigen among closely-related and/or disparate pathogens could be used to protect against different mycosis or even disease caused by others microorganisms [25,26]. For example, a β -glucan laminarin has demonstrated protection against infection by *Candida* and *Aspergillus* species by means of growth-inhibiting antibodies, particularly when conjugated with the diphtheria toxoid CRM197 carrier protein [27–29].

An interesting and intriguing aspect of fungal vaccines is the apparent existence of two major immunological mechanisms for achieving protection. The immune responses that have received the most study for fungal infection are a Th1 and/or Th17-based response and antibody-mediated immunity. Although both immune processes cooperate for the final protective outcome, the mechanisms are different. In particular, Th1 and/or Th17 immune response mediate protection indirectly, promoting an inflammatory response with recruitment of soluble (antimicrobial peptides, cytokines, chemokines) and cellular (macrophages, neutrophils) effectors that are responsible for the elimination or control of the fungal cells at the site of infection [30]. In contrast, antibodies can mediate protection not only by classical opsonization and complement activation, but also by direct neutralization of factors such as adhesins or enzymes, which are a critical step for infection, fungal growth, inhibiting fungal escape

from host immunity, or even directly killing the fungus [31–33]. Additionally, antibody binding to the fungal cell surface can directly regulate biological processes in the bound cells [34–36]. Recently, Boniche et al., (2020) described the approaches in immunotherapy against systemic mycosis using antibodies and the importance of this method for prospectively protecting immunocompromised host with defective cellular effectors.

Vaccine protection starts with the injection of an antigen source (live, inactivated, subunits, nucleic acids) that will be taken up/recognized by professional antigen-presenting cells (APCs) (macrophages or dendritic cells). After antigen phagocytosis, the APCs migrate to lymphoid organs where they interact with and present the antigen to lymphocytes. These lymphocytes are activated upon recognizing the antigen and by concomitant receipt of appropriate co-stimulatory signals, and the activated lymphocytes then produce a cell specific immune response. Activated B cells produce antibodies (IgG, IgM, IgA, IgE) that can target the invading fungal cells or otherwise modify immune cell responses. T-Cells are the major representative of cell-mediated immunity; activated Cytotoxic T-lymphocytes (CTLs, also called cytotoxic T-cells) can directly kill fungi, and T-helper cells (Th1-type) activate macrophages to enhance their capacity to kill intracellular pathogens. Furthermore, some of the B- and T-cells maintain themselves for many years as memory B- and T-cells such that they can rapidly activate and clonally expand when they encounter specific fungal antigens in the future and effectively combat the invading fungus [37–39]. A vaccine can be comprised by a live attenuated or inactivated microorganism or by one or more antigens. Antigens may be derived from the microbe, such as nucleic acids, proteins, carbohydrates or polysaccharides, and their efficacy can be enhanced by using targeted components of these structures, which is the case with peptide vaccines.

3. Fungal Peptide Vaccine

As discussed, immunization or vaccination using live-attenuated or inactivated pathogens (virus, bacteria, fungal, etc.) have been used for the induction of antigen-specific responses to protection against subsequent experimental infections. However, whole microbes contain thousands of distinct antigens and many are unnecessary for the induction of protective immune responses; moreover, some may induce unwanted responses, such as allergenic and/or reactogenic responses. These concerns have led to studies of subunits, such as a protein, from pathogens as vaccine candidates [40,41]. However, proteins are also relatively large and display many antigenic epitopes, which can also lead to adverse activities along with the induction of protective immune response. Therefore, peptide vaccines have been explored for their ability to induce desirable T cell and B cell-mediated immune response to highly defined, specific epitopes [42].

The first indication that a peptide vaccine could modify host-pathogen interactions arose from studies on tobacco mosaic virus in 1963, when Anderer demonstrated that conjugation of a hexapeptide derived from viral coat protein and coupled with bovine serum albumin could induce neutralizing antibodies to the intact virus [43]. Subsequently, Anderer demonstrated that synthetic tri-, tetra-, penta-, or hexapeptides also effectively generated neutralizing antibodies [44]. However, the work by Langebeheim and colleagues (1976) was the first to demonstrate that synthetic peptides derived from the coat protein of bacteriophage MS2 could induce antibodies that were as effective as those generated against the intact protein for neutralizing the bacteriophage. These discoveries together with ongoing technological advances, particularly in refining techniques for sequencing proteins and synthesizing peptides [45], spurred a marked increase in experimental peptide vaccine research in the 1980s [46]. Studies to date have demonstrated that engineered peptide vaccines can generally be considered as safe and cost effective when compared to conventional vaccines. However, the peptide's small size means that they are weakly immunogenic, such that they require transport molecules, which have the dual roles of serving as an adjuvant and promoting chemical stability [47].

Currently, the PubMed database (NCBI) contains thousands of reports on clinical studies of peptide vaccines for therapeutic or prophylactic use for diseases such as HIV, hepatitis C virus (HCV), hepatitis B virus (HBV), cytomegalovirus (CMV), influenza, tuberculosis, malaria, pneumonia, genital herpes,

and cancer, among others. However, it is also possible to find hundreds of studies of peptide vaccines for the treatment of fungal infection such as coccidioidomycosis, histoplasmosis, sporotrichosis, paracoccidioidomycosis, blastomycosis, aspergillosis, cryptococcosis, candidiasis, and other mycoses. The focus in this review is on new studies on engineering-based peptides for the treatment of fungal infections that are particularly due to *Paracoccidioides*, *Aspergillus*, *Candida*, *Sporothrix*, and *Coccidioides*. Table 1 provides an overview of the data from the major references detailed in this review.

Table 1. New vaccine proposals.

Fungi (Reference)	Vaccine (Peptide/Protein/Chimeric)	Immune Response	Results
<i>Paracoccidioides</i> [48–58]	Peptide vaccine (P10) P10 primary DC P10 primary monocyte derived-DC	CD4 ⁺ Th1 cell	Protection against i.t challenge, reduction of fungal burden both in immunosuppressed and immunocompetent mice, and efficacy of DNA vaccine; all tests were performed in animal models
	Prediction of sequence of epitopes from extracellular antigens	Potential to stimulate the immune response mediated by B cells and antibodies.	N.A.
<i>Aspergillus</i> [59,60]	Peptides from the protein Asp f1	Th1 cell	Peptides from Asp f1 stimulate production of Th1 cytokines.
<i>Candida</i> [33,61–84]	Fab and Met6 Peptides	Antibody	Mice immunized with either the Fba or Met6 peptide-DC vaccine had improved survival and reductions in fungal burdens in an immunosuppressed mouse model of disseminated candidiasis.
	14-mer Fab peptide conjugated each mimotopes from Met6 (PS2, PS31, PS28, PS55 and PS76) and	Specific antibody response	The peptides mimotopes induced a specific antibody response, and immunization with three of the peptide conjugate vaccines protected against disseminated candidiasis.
	18 peptides used to construct a multivalent recombinant protein	N.A.	N.A./requires specific HLA haplotypes to bind these particular peptide epitopes
	Recombinant protein (NDV-3 and NDV-3A)	B and T cells	Tested in Phase 1b/2a; one intramuscular dose was safe and NDV-3A was immunogenic and reduced frequency of recurrent vulvovaginal candidiasis (RVVC)
	Recombinant protein (NDV-3 and NDV-3A)	Antibodies and CD4 ⁺ Th1 Cell	Vaccinated mice were protected against lethal <i>C. auris</i> infection.
	Peptides (ZR1, ZR3, ZR3, ZR4, ZR5, ZR6, ZR7, ZR8)	CD4 ⁺ T cell	ZR3, ZR4 and ZR8 promoted cell proliferation in vitro. ZR8 induced IFN- γ , IL-17A and IL-1 β , and showed protection against <i>S. brasiliensis</i> infection
<i>Sporothrix</i> [85–87]	Phage displaying of the peptide KR	Th1 and Th17 cell and humoral immune response	Immunization with recombinant phage increased the survival rate of <i>S. globosa</i> infected mice.
<i>Coccidioides</i> [88–96]	Peptides from the protein Pep1	-	Induced IFN- γ production when exposed to lymphocytes.
	Peptides from the proteins Amn1 and Plb	-	Induced IFN- γ production by T-cells

Table 1. Cont.

Fungi (Reference)	Vaccine (Peptide/Protein/Chimeric)	Immune Response	Results
	recombinant T cell epitope-based vaccine (rEBV)	Th1, Th2, and Th17 cells	Mice immunized with rEBV had increased IFN- γ and IL-17 production, and they had significant reductions in fungal burden and prolongation of survival compared to nonvaccinated mice.
	Recombinant chimeric polypeptide vaccine (rCpa1)	Th1 and Th17 cell	rCap1 vaccination generated high levels of IL-17 in T-cell recall assays, earlier lung infiltration by activated Th1 and Th17, and increased the survival rates.
	Ag2/PRA-cDNA transfected DC	T cell	Vaccinated mice had lower fungal burdens and increased amounts of IFN- γ
	Ag2/PRA primary DC	T cell and IgG isotypes	Vaccinated mice did not show any illness or detectable injury and the immunization effectively induced IFN, IL-4 and IL-17 production
Pan fungal [97]	Calnexin peptide Recombinant calnexin (rCalnexin)	CD4 ⁺ Th1 and Th17 cells	rCalnexin formulated in GP reduced lung and spleen CFU in mice infected with <i>B. dermatitidis</i> or <i>Coccidioides posadasii</i> and prolonged survival. Calnexin peptide plus LPS delivery by i.v. route improved the expansion of calnexin-specific T cells.
Chimeric antigen receptor (CAR) T-cell therapy [98–100]	D-CAR T-cells	-	D-CAR ⁺ T-cells controlled the <i>Aspergillus</i> infections in the presence of immunosuppressive drugs

LPS, Lipopolysaccharide; CFU, colony form unit; i.v, intravenous; rEBV, bacterium-expressed recombinant epitope-based vaccine; rCpa1, recombinant chimeric polypeptide vaccine; DC, dendritic cell; RVVC, recurrent vulvovaginal candidiasis; VVC, vulvovaginal candidiasis; GPs, yeast cell wall-derived glucan particles; N.A., not analyzed.

3.1. Paracoccidioidomycosis (PCM)

Remarkably, fungal peptide vaccines are most advanced for the treatment of paracoccidioidomycosis, which is a neglected fungal disease restricted to Latin America [48]. Among the most promising treatments for Paracoccidioidomycosis is the vaccine using peptide 10 (P 10). This peptide was mapped based on the sequence of gp43, the main diagnostic antigen of *P. brasiliensis*. P10 is responsible for inducing lymphoproliferation and contains a major CD4⁺ specific T cell epitope and elicits an IFN- γ -dependent Th1 immune response, which is considered a protective and effective immune response against the infection with fungi of the genus *Paracoccidioides* [49–52]. Immunization with P10 proved to be protective in prophylactic and therapeutic murine infection models when injected with complete Freud's adjuvant [49]. Significantly, the peptide also protected against lethal infection in a model using immunosuppressed mice [53]. Rittner and collaborators (2012), using a gene therapy approach with a pcDNA3 expression vector encoding P10, demonstrated that this therapeutic DNA vaccine, given prior to or after infection, significantly reduced pulmonary fungal burdens in a murine infection model [54]. Also in 2012, Magalhães and collaborators showed that adoptive transfer of dendritic cells (DCs) pulsed with P10, either prior to or after infections, significantly protected mice from *P. brasiliensis* [55]. Building on these DC results, studies were also performed on immunocompromised animals. Bone marrow-derived dendritic cells (BMDCs) pulsed with P10 efficiently reduced the

pulmonary fungal burdens of immunosuppressed mice previously infected with *P. brasiliensis* and also preserved lung tissue by decreasing cellular infiltration into the organ [56]. Further work demonstrated that P10 was able to activate and modulate both BMDCs and monocyte-derived dendritic cells (MoDCs), and MoDCs pulsed with P10 similarly protected against pulmonary infection by *P. brasiliensis*, which is promising as this treatment most closely mirrors what would be administered to a patient with paracoccidioidomycosis [57].

The majority of research on antibodies in paracoccidioidomycosis have focused on their use for serological diagnosis, particularly as high titers of antibodies that are generated by patients with the acute-subacute form of the disease, which is the more aggressive form [101]. However, studies on antibody-mediated immune system modulations in response to an experimental model of cryptococcus infection [102–106] have led to an interest in the generation of protective antibodies for mitigating infections with *Paracoccidioides*. In fact, there are polyclonal [107] and monoclonal [108] antibodies that are protective in paracoccidioidomycosis. However, no vaccine specific for generating a humoral response is established for the treatment of paracoccidioidomycosis. Notably, a recent analysis of extracellular antigens from *Paracoccidioides* species using immunoproteomic approaches combined with immunoprecipitation using B-cells followed by antigen identification by nanoUPLC-MS^E-based proteomics demonstrated a variety of *Paracoccidioides* B-cell epitopes, common or specific to members in the species complex. Using bioinformatic tools, the proteins and the sequence of these epitopes from extracellular antigens were identified; however, these epitopes have not yet been tested. Nevertheless, this work highlights an opportunity for a new approach using synthetic peptides with the potential to stimulate antibody-mediated immune immunity [58].

3.2. Aspergillosis

Among the most feared fungal pathogens that are frequently clinically encountered are *Aspergillus* species, which are responsible for causing invasive aspergillosis as well as chronic bronchopulmonary aspergillosis. *Aspergillus fumigatus* is one of the most common species that notoriously causes infection in immunocompromised hosts, particularly in patients undergoing antineoplastic chemotherapy and those with organ transplants [109–114]. Although several laboratories have investigated the development of a safe and effective vaccine against aspergillosis and some promising results have been obtained in an experimental model using homologous proteins, crude extracts or recombinant allergens from *Aspergillus* [115–118], there is no vaccine against aspergillosis.

In silico assays, such as the use of artificial neural networks and immune epitope databases, facilitate the prediction of B cell epitopes and T cell MHC epitopes [119–121]. Subjecting *A. fumigatus* allergens to such analyses resulted in the identification of five potential allergic proteins (Asp f1, Asp f2, Asp f5, Asp f17, and Asp f34) with common B and T cell epitopes for both mice and humans [59]. Hence, these five proteins with high affinity binding to MHC class I or II epitopes could be used to characterize constituent peptides and develop vaccine candidates for invasive *Aspergillus* infections or therapeutics for allergy immunotherapy for chronic allergic bronchopulmonary aspergillosis. The promise of this approach is supported by prior work demonstrating that peptides from the protein Asp f1 stimulate the production of Th1 cytokines [60]. However, these proteins have not yet been validated as effective vaccine components.

3.3. Candidiasis

Candida species are the most frequently isolated fungal species in blood cultures worldwide and these opportunistic pathogens cause a wide range of infections. Disseminated bloodstream infection has an estimated mortality rate of 40–60% even with the use of antifungal drugs [122–125]. Additionally, *Candida* commonly cause vaginitis, oral thrush, and infections of the skin and nails. Despite the incredibly high overall disease incidence as well as the frequency and severity of invasive infections, there is no vaccine for *Candida* species. However, peptide-based vaccine strategies have

been considered for over two decades for both prevention and protection through active and passive immunization [2,126,127].

Several studies demonstrated that antibodies specific for the peptide Fba or peptide Met6, which were respectively derived from *C. albicans* cell surface fructose biphosphate aldolase (Fab) or β -1,2-mannotriose [β -(Man)3] protein, were induced by a protective glycopeptide vaccine [33,61–63]. More recently, active immunization using DCs pulsed with either Fba peptide (YGKDVKDLFDYAQE) or Met6 peptide (PRIGGQRELKKITE) were protective in both neutropenic and immunocompetent mice [64]. Subsequently, a study of a synthetic 14-mer Fba peptide conjugated to each of the five peptides mimotopes from Met6 (PS2, PS31, PS28, PS55, and PS76) were tested to explore their protective capacity [65]. All five peptides mimotopes induced specific antibody responses, and immunization with three of the peptide conjugate vaccines protected against disseminated candidiasis in mice [65].

Another recent peptide vaccine-based approach was achieved using computational tools to identify immunologically active compounds to combat candidiasis. A screen of 6030 proteins identified in the proteome of *Candida albicans* (sc5314) [66] was undertaken to identify immunodominant HLA class I, HLA class II and linear and discontinuous B-cell epitopes. The screen identified 214 epitopes that were subjected to conservation analysis using 22 *C. albicans* strains with published sequenced genomes, and 18 peptides displayed 100% conservancy. The 18 peptides were then used to construct a multivalent recombinant protein to which they added a synthetic adjuvant called RS09. However, the investigators do not yet describe the efficacy of this polymeric vaccine, and it is notable that this type of protein peptide-based vaccine will only generate responses in patients with specific HLA haplotypes that are able to bind these particular peptide epitopes [67]. Nevertheless, this type of vaccine approach may generate effective immune response using well-defined minimal quantities of antigen, which may minimize unwanted side effects.

There is a growing literature on the production and release of fungal extracellular vesicles (EV), which occurs in both ascomycetes and basidiomycetes [68,69]. These EV contain large quantities of biologically functional compounds that are associated with virulence, including in EV from *Candida* [70]. Recent data highlights how these relatively stable EV can be used as safe source for diverse antigens, including peptides, for vaccine development as administration of *Candida* EV are protective in a murine systemic candidiasis infection model [71].

The vaccine proposal in the most advanced phase of study for combatting a fungal infection is NDV-3A [72]. This vaccine is based on *C. albicans* Als3p, which is a glycoprotein with an agglutinin-like sequence that is associated with virulence through effects on fungal adherence, invasion and biofilm formation [73,74]. The first version of this vaccine, NDV-3, was a His-tagged recombinant Als3 protein N-terminus (rAls3p-N), combined with alum, which was protective in a disseminated candidiasis experimental model [75–78]. In a Phase I clinical trial, NDV-3 was highly immunogenic and well-tolerated [78]. The more recent version of the vaccine, NDV-3A, was prepared with rAls3p-N without the His-tag, and again combined with alum [72]. An exploratory Phase 1b/2a study found that a single intramuscular dose of NDV-3 was safe and immunogenic. In a phase 2 randomized, double-blind, placebo-control trial, NDV-3A was administered to women with recurrent vulvovaginal candidiasis (RVVC) and the vaccine was found to be safe and vaccinated women rapidly developed both T and B cell responses to rAls3p-N. In what the authors describe as “unprecedented”, the vaccine effectively reduced the occurrence and frequency of vulvovaginal candidiasis (VVC) episodes for up to 12 months [72]. Additionally, serum from patients who responded to NDV-3A contained antibodies that prevented fungal adhesion and biofilm formation on plastic as well as fungal invasion of vaginal epithelial cells in vitro [79]. This group recently demonstrated that mice vaccinated with NDV-3A developed high titers of anti-rAls3p-N antibodies and that the presence of these antibodies was sufficient to block *C. albicans* from colonizing jugular vein catheters [80]. The NDV-3A is a highly promising vaccine for treatment of RVVC as well as invasive candidiasis.

NDV-3A has also been tested against *C. auris*, which is an emerging, multi-drug resistant species [81,82] that has marked biological differences from *C. albicans* [83]. Homologs of the *C. albicans*

Als3p, glycoprotein base of the NDV-3A vaccine, are present in isolates of different *C. auris* clades [84]. Preliminary studies have revealed that mice vaccinated with NDV-3A develop antibodies to Als3p and that these antibodies recognize *C. auris* yeast cells in vitro, block their ability to form biofilms, and improve macrophage-mediated fungal killing. In an in vivo murine model, NDV-3A effectively induced cross-reactive humoral and cellular immune responses that protected immunosuppressed mice who received a lethal challenge of *C. auris*. Furthermore, NDV-3A improved the efficacy of sub-therapeutic doses of micafungin [84]. Thus, the advanced development of NDV-3A is extremely promising for use not only against *C. albicans* but also against multidrug-resistant *C. auris*.

3.4. Sporotrichosis

The species that make up the *Sporothrix* complex (*Sporothrix schenckii*, *S. brasiliensis*, *S. globosa* and *S. luriei*) are distributed worldwide [128]. Although cutaneous and lymphocutaneous forms are by far the most common disease manifestations, disseminated disease may occur in immunocompromised patients [129,130] and pulmonary disease may manifest after conidia or propagules of the fungus are inhaled [131]. Significantly, *S. brasiliensis* has an increased tendency to disseminate even in the absence of any immune defect [132]. Several countries have reported an increase in the number of cases of feline zoonotic transmission, and this epidemic is primarily due to *S. brasiliensis* in Brazil [133,134]. Although monoclonal antibodies have experimentally produced positive results [135,136], the search for new and more efficient treatment modalities is ongoing.

Given the increasing incidence and disease severity with *S. brasiliensis*, de Almeida and colleagues undertook a proteomic analysis using the in-silico prediction tools to identify peptides with high affinity to MHC class II. They identified seven peptides that met their criteria, which were synthesized and tested in mice. Three of the peptide vaccines induced proliferation of T cells sensitized by *S. brasiliensis* in vitro. In subsequent in vivo experiments, immunization with each of the three peptides mixed in Freund's incomplete adjuvant reduced the severity of subcutaneous sporotrichosis and one peptide induced the production of high levels of inflammatory cytokines [85]. This work confirmed that a peptide vaccine could effectively induce a protective immune response against *S. brasiliensis*.

Sporothrix glycoprotein Gp70 is a major adhesion factor on the fungal cell surface. The structure of Gp70 was screened by bioinformatic tools, and four peptides were identified and displayed on phage. Of the four, the phage displaying of the peptide KR (kpvqhahltplgldr) protected mice against infection with *S. globosa*, which is the most common species isolated in Northeast China [86]. Furthermore, the mice immunized with phage-KR produced high levels of IFN- γ + Th1 and IL-17 + Th17, indicating that the mechanism for protection by the recombinant phage may occur through the induction of a protective cell-mediated immune responses. However, serum from mice infected with *S. globosa* was also specifically recognized by phage-KR, which suggests that the humoral response may also be stimulated by the KR peptide [87]. Therefore, the phage-KR vaccine may function to enhance both protective cell-mediated and humoral immune responses, and this dual mechanism therapeutic represents a new and potentially safe strategy for the treatment of sporotrichosis.

3.5. Coccidioidomycosis

Coccidioides spp., are environmental pathogens that are responsible for the human respiratory disease Coccidioidomycosis, which occurs mainly in desert soils of the south-western United States and parts of Mexico and Central and South America [137–139]. Between 1998 and 2011, the number of reported cases of coccidioidomycosis increased from 5.3 to 42.6 per 100,000 inhabitants in the endemic region of the United States [140], which correlated with changes in weather and disturbances in soils. Recent studies also show that 17 to 29% of pneumonia cases in endemic regions are due to *Coccidioides* and the endemic regions are expanding [141,142]. Pursuing a vaccine for *Coccidioides* is not a new idea. In fact, a formalin killed spherule vaccine was developed and a clinical trial performed. From 1980–1985, 2876 patients were randomized to receive either three injections of 1.75 mg of FKS or placebo. Unfortunately, no differences in protection were observed [143,144], which may have been due to the

low incidence of coccidioidomycosis during this period or the low dosing, which was required due to toxicity challenges [145].

Following the result with the whole cell vaccine, investigators turned their attentions to specific antigens for study and development. For example, in 2006, Pep1 was shown to be a cell wall dominant antigen that was protective in mice challenged with *C. posadasii*. By applying immunoproteomic and bioinformatic tools, investigators then identified five peptides from Pep1 that were predicted to have high affinity to MHC-II, and these peptides were able to induce IFN- γ by peptide-exposed lymphocytes [88]. Also in 2006, peptides from two additional proteins, Amn1 and Plb, from *C. posadasii* were similarly demonstrated to stimulate IFN- γ production by T-cells [89]. In 2012, an epitope base vaccine (rEBV) [90] consisting of selected immunogenic peptides derived from Pep1, Amn1 and Plb was developed [88,89]. The rEBV significantly reduced fungal burdens, elevated IFN- γ and interleukin (IL)-17 production, as well as prolonged survival in vaccinated mice challenged with a lethal inoculum of *Coccidioides* compared to untreated infected mice [90].

More recently, a recombinant chimeric polypeptide vaccine (rCpa1) [91] was generated using the most immunogenic fragment of Ag2/Pra; the full lengths of Cs-Ag and Pmp1; and promiscuous, immunodominant T-cell epitopes derived from *Coccidioides posadasii*, Pep1, Amn1, and Plb. Administration of rCpa1 with the adjuvant GCP to both C57BL/6 and HLA-DR4 transgenic mice induced high levels of IL-17 in T-cell recall assays, earlier lung infiltration by activated Th1 and Th17, and increased the survival rates of mice lethally infected with *Coccidioides* compared to those that received GCPs alone [91].

Ag2/PRA has also been investigated for utilization in a dendritic cell vaccine for coccidioidomycosis [92–96]. In 2005, Ag2/PRA cDNA transfected into an immortalized dendritic JAWS II cell line was used to immunize mice challenged with *C. posadasii*, and the DC vaccine reduced the fungal burdens in both the lungs and spleens, and increased the amount of IFN- γ in the lung tissues of immunized compared to control mice [92]. Subsequently, Ag2/PRA-cDNA transfected bone marrow-derived dendritic cells administered intranasally were shown to migrate in blood, lung and thymus, and the vaccine induced Ag2/PRA-specific T cell response [94]. Safety testing revealed that intranasal immunization with an Ag2/PRA primary DC vaccine did not cause any illness or detectable injury to the mice. Immunization effectively induced the production of IFN, IL-4 and IL-17 in the lungs and lymph nodes of the vaccinated animals. Additionally, vaccination induced the production of all IgG isotypes [95]. These studies highlight the promising potential of this DC vaccine (Ag2-DC) for coccidioidomycosis as it effectively induced both cellular and humoral immune responses [92,94,95].

3.6. Pan-Fungal Vaccine

Although a vaccine to a single fungal pathogen would be heralded as a remarkable achievement, there is a desire to create a pan-fungal vaccine. The possibility for this is presented by work using monoclonal antibodies to conserved fungal cell surface epitopes to deliver cytotoxic radiation [146–148]. However, peptide-based vaccines also hold great potential. In 2015, transgenic CD4⁺ T cells were used to identify an amino acid determinant within chaperone calnexin that was determined to be conserved across ascomycetes species [97]. The administration of calnexin in glucan particles elicited calnexin-specific CD4⁺ T cells, and vaccinated mice demonstrated resistance to infection by *Blastomyces dermatitidis*, *Histoplasma capsulatum*, *Pseudogymnoascus (Geomyces) destructans*, *Fonsecaea pedrosoi*, and *A. fumigatus* [97]. Similarly, the 13-mer peptide (LVVKNPAAHHAIS), which was generated from the conserved region amino acid determinant within chaperone calnexin, stimulated protective immune responses, and vaccination reduced the severity of infection with *B. dermatitidis* [97].

4. Chimeric Antigen Receptor (CAR) T-Cell Therapy

CAR T-cell therapy has primarily been used in combating diverse cancers [149,150], but there is a growing interest for its use in other diseases, including mycoses. CAR T-cell approaches use a patient's

T-cells to engineer them into chimeric cells that target both a specific antigen and activate other T cells. Currently, the target antigens are glycoproteins and lipids [151,152].

D-CAR is the second-generation of CAR T-cells therapy that targets Dectin-1, which is a C-type lectin receptor specific for β -glucan that is commonly expressed on the surface of diverse fungi [98]. These D-CAR T cells displayed specificity to Dectin-1 that was fused with CD28 and CD3- ζ such that effective T-cell activation signaling was generated. The administration of D-CAR T-cells to immunocompromised mice with invasive aspergillosis resulted in an increase in the levels of IFN- γ and impaired the growth of the *Aspergillus* [99]. Investigators in this area describe how CAR T-cell approaches can be dually impactful by designing the cells to target both the patient's underlying cancer as well as a concomitant invasive fungal infection [100]. The use of engineered CAR T-cells engineering has been remarkably effective in cancer immunotherapy, and it has shown promise as a therapeutic for combating infections by viruses [153–159] and fungi [99], albeit it remains in early phase development for the treatment of mycoses.

5. Conclusions

Fungal diseases are widely neglected [160] and this extends to the development of vaccines to prevent and treat individuals with mycoses. Given that there are over 300 million people suffering from fungal infections annually with over 1.5 million of these dying [16,161,162], it is imperative that vaccine development be accelerated to combat these diseases. As one of the main factors that contribute to the increased frequency and severity of disease are defects cellular and/or humoral immunity [163], vaccine strategies must be safe and effective in hosts with intact and compromised immune systems. Advances in proteomics and system biology have facilitated the advancement of a number of vaccine proposals, particularly as they permit the localization of proteins and the characterization of their modifications, functions and interactions [164]. Predictions of epitope biology permit the rapid selection of peptides with expected immunogenicity that can be injected into hosts for presentation by professional APCs for subsequent recognition by B or T lymphocytes to induce a humoral or cellular immune response, respectively. The advancing studies with peptide vaccines and DC-peptide priming set the stage for future translation of these strategies from the bench to the bedside.

Author Contributions: L.B.R.D.S. wrote the manuscript. J.D.N., C.P.T. revised the manuscript. J.D.N. wrote and revised the manuscript. All authors have read and agreed to the published version of the manuscript.

Funding: This work was supported by grants from the FAPESP, CNPq, and CAPES. CPT is a research fellow of the CNPq. FAPESP numbers: 2016/08730-6, 2018/25171-6 and 2019/20622-2. FAPESP, for the financial support during my stay in NY, which allowed me to have the opportunity to participate in this review.

Conflicts of Interest: The authors declare no conflict of interest. The funders had no role in the design of the study; in the collection, analyses, or interpretation of data; in the writing of the manuscript, or in the decision to publish the results.

References

1. Plotkin, S.A. Vaccines: The fourth century. *Clin. Vaccine Immunol.* **2009**, *16*, 1709–1719. [[CrossRef](#)] [[PubMed](#)]
2. Cutler, J.E.; Deepe, G.S., Jr.; Klein, B.S. Advances in combating fungal diseases: Vaccines on the threshold. *Nat. Rev. Microbiol.* **2007**, *5*, 13–28. [[CrossRef](#)] [[PubMed](#)]
3. Cassone, A. Fungal vaccines: Real progress from real challenges. *Lancet Infect. Dis.* **2008**, *8*, 114–124. [[CrossRef](#)]
4. Spellberg, B. Vaccines for invasive fungal infections. *F1000 Med. Rep.* **2011**, *3*, 13. [[CrossRef](#)]
5. Stevens, D.A.; Clemons, K.V.; Liu, M. Developing a vaccine against aspergillosis. *Med. Mycol.* **2011**, *49* (Suppl. 1), S170–S176. [[CrossRef](#)] [[PubMed](#)]
6. Cassone, A.; Casadevall, A. Recent progress in vaccines against fungal diseases. *Curr. Opin. Microbiol.* **2012**, *15*, 427–433. [[CrossRef](#)]

7. De Amorim, J.; Magalhaes, A.; Munoz, J.E.; Rittner, G.M.; Nosanchuk, J.D.; Travassos, L.R.; Taborda, C.P. DNA vaccine encoding peptide P10 against experimental paracoccidioidomycosis induces long-term protection in presence of regulatory T cells. *Microbes Infect.* **2013**, *15*, 181–191. [\[CrossRef\]](#)
8. Edwards, J.E. Fungal cell wall vaccines: An update. *J. Med. Microbiol.* **2012**, *61*, 895–903. [\[CrossRef\]](#)
9. Boniche, C.; Rossi, S.A.; Kischkel, B.; Barbalho, F.V.; Moura, A.N.D.; Nosanchuk, J.D.; Travassos, L.R.; Taborda, C.P. Immunotherapy against Systemic Fungal Infections Based on Monoclonal Antibodies. *J. Fungi* **2020**, *6*, 31. [\[CrossRef\]](#)
10. Clark, C.; Drummond, R.A. The Hidden Cost of Modern Medical Interventions: How Medical Advances Have Shaped the Prevalence of Human Fungal Disease. *Pathogens* **2019**, *8*, 45. [\[CrossRef\]](#)
11. Papon, N.; Bounoux, M.E.; d’Enfert, C. Tracing the Origin of Invasive Fungal Infections. *Trends Microbiol.* **2020**, *28*, 240–242. [\[CrossRef\]](#) [\[PubMed\]](#)
12. Benedict, K.; Jackson, B.R.; Chiller, T.; Beer, K.D. Estimation of Direct Healthcare Costs of Fungal Diseases in the United States. *Clin. Infect. Dis.* **2019**, *68*, 1791–1797. [\[CrossRef\]](#) [\[PubMed\]](#)
13. Oliveira, G.G.; Belitardo, D.R.; Balarin, M.R.; Freire, R.L.; Camargo, Z.P.; Ono, M.A. Serological survey of paracoccidioidomycosis in cats. *Mycopathologia* **2013**, *176*, 299–302. [\[CrossRef\]](#)
14. Rudkin, F.M.; Raziunaite, I.; Workman, H.; Essono, S.; Belmonte, R.; MacCallum, D.M.; Johnson, E.M.; Silva, L.M.; Palma, A.S.; Feizi, T.; et al. Single human B cell-derived monoclonal anti-Candida antibodies enhance phagocytosis and protect against disseminated candidiasis. *Nat. Commun.* **2018**, *9*, 5288. [\[CrossRef\]](#) [\[PubMed\]](#)
15. Denning, D.W. The ambitious ‘95–95 by 2025’ roadmap for the diagnosis and management of fungal diseases. *Thorax* **2015**, *70*, 613–614. [\[CrossRef\]](#) [\[PubMed\]](#)
16. Brown, G.D.; Denning, D.W.; Gow, N.A.; Levitz, S.M.; Netea, M.G.; White, T.C. Hidden killers: Human fungal infections. *Sci. Transl. Med.* **2012**, *4*, 165rv113. [\[CrossRef\]](#) [\[PubMed\]](#)
17. Rajasingham, R.; Smith, R.M.; Park, B.J.; Jarvis, J.N.; Govender, N.P.; Chiller, T.M.; Denning, D.W.; Loyse, A.; Boulware, D.R. Global burden of disease of HIV-associated cryptococcal meningitis: An updated analysis. *Lancet Infect. Dis.* **2017**, *17*, 873–881. [\[CrossRef\]](#)
18. Rappuoli, R.; Mandl, C.W.; Black, S.; De Gregorio, E. Vaccines for the twenty-first century society. *Nat. Rev. Immunol.* **2011**, *11*, 865–872. [\[CrossRef\]](#)
19. Moxon, E.R.; Siegrist, C.A. The next decade of vaccines: Societal and scientific challenges. *Lancet* **2011**, *378*, 348–359. [\[CrossRef\]](#)
20. Stevens, D.A. Vaccinate against aspergillosis! A call to arms of the immune system. *Clin. Infect. Dis.* **2004**, *38*, 1131–1136. [\[CrossRef\]](#)
21. Sheppard, D.C.; Edwards, J.E., Jr. Development of a vaccine for invasive aspergillosis. *Clin. Infect. Dis.* **2004**, *38*, 1137–1138. [\[CrossRef\]](#) [\[PubMed\]](#)
22. Gow, N.A.R.; Latge, J.P.; Munro, C.A. The Fungal Cell Wall: Structure, Biosynthesis, and Function. *Microbiol. Spectr.* **2017**, *5*. [\[CrossRef\]](#)
23. Alcazar-Fuoli, L.; Mellado, E. Ergosterol biosynthesis in *Aspergillus fumigatus*: Its relevance as an antifungal target and role in antifungal drug resistance. *Front. Microbiol.* **2012**, *3*, 439. [\[CrossRef\]](#) [\[PubMed\]](#)
24. Valiante, V.; Macheleidt, J.; Foge, M.; Brakhage, A.A. The *Aspergillus fumigatus* cell wall integrity signaling pathway: Drug target, compensatory pathways, and virulence. *Front. Microbiol.* **2015**, *6*, 325. [\[CrossRef\]](#)
25. Cassone, A.; Rappuoli, R. Universal vaccines: Shifting to one for many. *mBio* **2010**, *1*. [\[CrossRef\]](#) [\[PubMed\]](#)
26. Hamad, M. Universal fungal vaccines: Could there be light at the end of the tunnel? *Hum. Vaccines Immunother.* **2012**, *8*, 1758–1763. [\[CrossRef\]](#)
27. Bromuro, C.; Romano, M.; Chiani, P.; Berti, F.; Tontini, M.; Proietti, D.; Mori, E.; Torosantucci, A.; Costantino, P.; Rappuoli, R.; et al. Beta-glucan-CRM197 conjugates as candidates antifungal vaccines. *Vaccine* **2010**, *28*, 2615–2623. [\[CrossRef\]](#)
28. Pietrella, D.; Rachini, A.; Torosantucci, A.; Chiani, P.; Brown, A.J.; Bistoni, F.; Costantino, P.; Mosci, P.; d’Enfert, C.; Rappuoli, R.; et al. A beta-glucan-conjugate vaccine and anti-beta-glucan antibodies are effective against murine vaginal candidiasis as assessed by a novel in vivo imaging technique. *Vaccine* **2010**, *28*, 1717–1725. [\[CrossRef\]](#)
29. Torosantucci, A.; Bromuro, C.; Chiani, P.; De Bernardis, F.; Berti, F.; Galli, C.; Norelli, F.; Bellucci, C.; Polonelli, L.; Costantino, P.; et al. A novel glyco-conjugate vaccine against fungal pathogens. *J. Exp. Med.* **2005**, *202*, 597–606. [\[CrossRef\]](#)

30. Romani, L. Immunity to fungal infections. *Nat. Rev. Immunol.* **2011**, *11*, 275–288. [[CrossRef](#)]
31. Rachini, A.; Pietrella, D.; Lupo, P.; Torosantucci, A.; Chiani, P.; Bromuro, C.; Proietti, C.; Bistoni, F.; Cassone, A.; Vecchiarelli, A. An anti-beta-glucan monoclonal antibody inhibits growth and capsule formation of *Cryptococcus neoformans* in vitro and exerts therapeutic, anticytotoxic activity in vivo. *Infect. Immun.* **2007**, *75*, 5085–5094. [[CrossRef](#)] [[PubMed](#)]
32. Torosantucci, A.; Chiani, P.; Bromuro, C.; De Bernardis, F.; Palma, A.S.; Liu, Y.; Mignogna, G.; Maras, B.; Colone, M.; Stringaro, A.; et al. Protection by anti-beta-glucan antibodies is associated with restricted beta-1,3 glucan binding specificity and inhibition of fungal growth and adherence. *PLoS ONE* **2009**, *4*, e5392. [[CrossRef](#)] [[PubMed](#)]
33. Xin, H.; Cutler, J.E. Vaccine and monoclonal antibody that enhance mouse resistance to candidiasis. *Clin. Vaccine Immunol.* **2011**, *18*, 1656–1667. [[CrossRef](#)] [[PubMed](#)]
34. Burnet, M.C.; Zamith-Miranda, D.; Heyman, H.M.; Weitz, K.K.; Bredeweg, E.L.; Nosanchuk, J.D.; Nakayasu, E.S. Remodeling of the *Histoplasma Capsulatum* Membrane Induced by Monoclonal Antibodies. *Vaccines* **2020**, *8*, 69. [[CrossRef](#)] [[PubMed](#)]
35. Coordoba, J.; Otero, M.C.; Lainez, B.; Perez-Tamarit, D.; Molina, J.M.; Calvo, M.A.; Asensi, F.; Gobernado, M. Human immunodeficiency virus and resistance. *Rev. Esp. Quimioter.* **1998**, *11*, 152–156.
36. Matos Baltazar, L.; Nakayasu, E.S.; Sobreira, T.J.; Choi, H.; Casadevall, A.; Nimrichter, L.; Nosanchuk, J.D. Antibody Binding Alters the Characteristics and Contents of Extracellular Vesicles Released by *Histoplasma capsulatum*. *mSphere* **2016**, *1*. [[CrossRef](#)]
37. Jiskoot, W.; Kersten, G.F.A.; Mastrobattista, E.; Slütter, B. Vaccines. In *Pharmaceutical Biotechnology: Fundamentals and Applications*; Crommelin, D.J.A., Sindelar, R.D., Meibohm, B., Eds.; Springer International Publishing: Cham, Switzerland, 2019; pp. 281–304. [[CrossRef](#)]
38. Chai, L.Y.; Netea, M.G.; Vonk, A.G.; Kullberg, B.J. Fungal strategies for overcoming host innate immune response. *Med. Mycol.* **2009**, *47*, 227–236. [[CrossRef](#)]
39. Hernandez-Chavez, M.J.; Perez-Garcia, L.A.; Nino-Vega, G.A.; Mora-Montes, H.M. Fungal Strategies to Evade the Host Immune Recognition. *J. Fungi* **2017**, *3*, 51. [[CrossRef](#)]
40. Thompson, A.L.; Staats, H.F. Cytokines: The future of intranasal vaccine adjuvants. *Clin. Dev. Immunol.* **2011**, *2011*, 289597. [[CrossRef](#)]
41. Petrovsky, N.; Aguilar, J.C. Vaccine adjuvants: Current state and future trends. *Immunol. Cell Biol.* **2004**, *82*, 488–496. [[CrossRef](#)]
42. Sesardic, D. Synthetic peptide vaccines. *J. Med. Microbiol.* **1993**, *39*, 241–242. [[CrossRef](#)] [[PubMed](#)]
43. Anderer, F.A. Preparation and properties of an artificial antigen immunologically related to tobacco mosaic virus. *Biochim. Biophys. Acta* **1963**, *71*, 246–248. [[CrossRef](#)]
44. Anderer, F.A.; Schlumberger, H.D. Properties of Different Artificial Antigens Immunologically Related to Tobacco Mosaic Virus. *Biochim. Biophys. Acta* **1965**, *97*, 503–509. [[CrossRef](#)]
45. Merrifield, R.B. Solid Phase Peptide Synthesis. I. The Synthesis of a Tetrapeptide. *J. Am. Chem. Soc.* **1963**, *85*, 2149–2154. [[CrossRef](#)]
46. Francis, M.J. Peptide vaccines for viral diseases. *Sci. Prog.* **1990**, *74*, 115–130.
47. Aguilar, J.C.; Rodriguez, E.G. Vaccine adjuvants revisited. *Vaccine* **2007**, *25*, 3752–3762. [[CrossRef](#)]
48. Tabor, C.P.; Uran, M.E.; Nosanchuk, J.D.; Travassos, L.R. Paracoccidioidomycosis: Challenges in the Development of a Vaccine against an Endemic Mycosis in the Americas. *Rev. Inst. Med. Trop. Sao Paulo* **2015**, *57* (Suppl. 19), 21–24. [[CrossRef](#)]
49. Tabor, C.P.; Juliano, M.A.; Puccia, R.; Franco, M.; Travassos, L.R. Mapping of the T-cell epitope in the major 43-kilodalton glycoprotein of *Paracoccidioides brasiliensis* which induces a Th-1 response protective against fungal infection in BALB/c mice. *Infect. Immun.* **1998**, *66*, 786–793. [[CrossRef](#)]
50. Travassos, L.R.; Tabor, C.P.; Iwai, L.K.; Cunha-Neto, E.C.; Puccia, R. *The gp43 from Paracoccidioides Brasiliensis: A Major Diagnostic Antigen and Vaccine Candidate*; Springer: Berlin/Heidelberg, Germany, 2004; Volume 12.
51. Souto, J.T.; Figueiredo, F.; Furlanetto, A.; Pfeffer, K.; Rossi, M.A.; Silva, J.S. Interferon-gamma and tumor necrosis factor-alpha determine resistance to *Paracoccidioides brasiliensis* infection in mice. *Am. J. Pathol.* **2000**, *156*, 1811–1820. [[CrossRef](#)]
52. Travassos, L.R.; Tabor, C.P. Linear Epitopes of *Paracoccidioides brasiliensis* and Other Fungal Agents of Human Systemic Mycoses As Vaccine Candidates. *Front. Immunol.* **2017**, *8*, 224. [[CrossRef](#)]

53. Munoz, J.E.; Luft, V.D.; Amorim, J.; Magalhaes, A.; Thomaz, L.; Nosanchuk, J.D.; Travassos, L.R.; Taborda, C.P. Immunization with P10 peptide increases specific immunity and protects immunosuppressed BALB/c mice infected with virulent yeasts of *Paracoccidioides brasiliensis*. *Mycopathologia* **2014**, *178*, 177–188. [[CrossRef](#)] [[PubMed](#)]
54. Rittner, G.M.; Munoz, J.E.; Marques, A.F.; Nosanchuk, J.D.; Taborda, C.P.; Travassos, L.R. Therapeutic DNA vaccine encoding peptide P10 against experimental paracoccidioidomycosis. *PLoS Negl. Trop. Dis.* **2012**, *6*, e1519. [[CrossRef](#)] [[PubMed](#)]
55. Magalhaes, A.; Ferreira, K.S.; Almeida, S.R.; Nosanchuk, J.D.; Travassos, L.R.; Taborda, C.P. Prophylactic and therapeutic vaccination using dendritic cells primed with peptide 10 derived from the 43-kilodalton glycoprotein of *Paracoccidioides brasiliensis*. *Clin. Vaccine Immunol.* **2012**, *19*, 23–29. [[CrossRef](#)] [[PubMed](#)]
56. Silva, L.B.R.; Dias, L.S.; Rittner, G.M.G.; Munoz, J.E.; Souza, A.C.O.; Nosanchuk, J.D.; Travassos, L.R.; Taborda, C.P. Dendritic Cells Primed with *Paracoccidioides brasiliensis* Peptide P10 Are Therapeutic in Immunosuppressed Mice with Paracoccidioidomycosis. *Front. Microbiol.* **2017**, *8*, 1057. [[CrossRef](#)] [[PubMed](#)]
57. Silva, L.B.R.; Taira, C.L.; Dias, L.S.; Souza, A.C.O.; Nosanchuk, J.D.; Travassos, L.R.; Taborda, C.P. Experimental Therapy of Paracoccidioidomycosis Using P10-Primed Monocyte-Derived Dendritic Cells Isolated From Infected Mice. *Front. Microbiol.* **2019**, *10*, 1727. [[CrossRef](#)] [[PubMed](#)]
58. Moreira, A.L.E.; Oliveira, M.A.P.; Silva, L.O.S.; Inacio, M.M.; Bailao, A.M.; Parente-Rocha, J.A.; Cruz-Leite, V.R.M.; Paccez, J.D.; de Almeida Soares, C.M.; Weber, S.S.; et al. Immunoproteomic Approach of Extracellular Antigens From *Paracoccidioides* Species Reveals Exclusive B-Cell Epitopes. *Front. Microbiol.* **2019**, *10*, 2968. [[CrossRef](#)]
59. Thakur, R.; Shankar, J. In silico Identification of Potential Peptides or Allergen Shot Candidates Against *Aspergillus fumigatus*. *Biores Open Access* **2016**, *5*, 330–341. [[CrossRef](#)]
60. Chaudhary, N.; Mahajan, L.; Madan, T.; Kumar, A.; Raghava, G.P.; Katti, S.B.; Haq, W.; Sarma, P.U. Prophylactic and Therapeutic Potential of Asp f1 Epitopes in Naive and Sensitized BALB/c Mice. *Immune Netw.* **2009**, *9*, 179–191. [[CrossRef](#)]
61. Xin, H.; Dziadek, S.; Bundle, D.R.; Cutler, J.E. Synthetic glycopeptide vaccines combining beta-mannan and peptide epitopes induce protection against candidiasis. *Proc. Natl. Acad. Sci. USA* **2008**, *105*, 13526–13531. [[CrossRef](#)]
62. Han, Y.; Kozel, T.R.; Zhang, M.X.; MacGill, R.S.; Carroll, M.C.; Cutler, J.E. Complement is essential for protection by an IgM and an IgG3 monoclonal antibody against experimental, hematogenously disseminated candidiasis. *J. Immunol.* **2001**, *167*, 1550–1557. [[CrossRef](#)]
63. Xin, H.; Cutler, J.E. Hybridoma passage in vitro may result in reduced ability of antimannan antibody to protect against disseminated candidiasis. *Infect. Immun.* **2006**, *74*, 4310–4321. [[CrossRef](#)] [[PubMed](#)]
64. Xin, H. Active immunizations with peptide-DC vaccines and passive transfer with antibodies protect neutropenic mice against disseminated candidiasis. *Vaccine* **2016**, *34*, 245–251. [[CrossRef](#)] [[PubMed](#)]
65. Xin, H.; Glee, P.; Adams, A.; Mohiuddin, F.; Eberle, K. Design of a mimotope-peptide based double epitope vaccine against disseminated candidiasis. *Vaccine* **2019**, *37*, 2430–2438. [[CrossRef](#)] [[PubMed](#)]
66. Sampaio, P.; Santos, M.; Correia, A.; Amaral, F.E.; Chavez-Galarza, J.; Costa-de-Oliveira, S.; Castro, A.G.; Pedrosa, J.; Pais, C. Virulence attenuation of *Candida albicans* genetic variants isolated from a patient with a recurrent bloodstream infection. *PLoS ONE* **2010**, *5*, e10155. [[CrossRef](#)]
67. Tarang, S.; Kesharwani, V.; LaTendresse, B.; Lindgren, L.; Rocha-Sanchez, S.M.; Weston, M.D. In silico Design of a Multivalent Vaccine Against *Candida albicans*. *Sci. Rep.* **2020**, *10*, 1066. [[CrossRef](#)]
68. Albuquerque, P.C.; Nakayasu, E.S.; Rodrigues, M.L.; Frases, S.; Casadevall, A.; Zancoppe-Oliveira, R.M.; Almeida, I.C.; Nosanchuk, J.D. Vesicular transport in *Histoplasma capsulatum*: An effective mechanism for trans-cell wall transfer of proteins and lipids in ascomycetes. *Cell Microbiol.* **2008**, *10*, 1695–1710. [[CrossRef](#)]
69. Rodrigues, M.L.; Nosanchuk, J.D.; Schrank, A.; Vainstein, M.H.; Casadevall, A.; Nimrichter, L. Vesicular transport systems in fungi. *Future Microbiol.* **2011**, *6*, 1371–1381. [[CrossRef](#)]
70. Vargas, G.; Rocha, J.D.; Oliveira, D.L.; Albuquerque, P.C.; Frases, S.; Santos, S.S.; Nosanchuk, J.D.; Gomes, A.M.; Medeiros, L.C.; Miranda, K.; et al. Compositional and immunobiological analyses of extracellular vesicles released by *Candida albicans*. *Cell Microbiol.* **2015**, *17*, 389–407. [[CrossRef](#)]
71. Vargas, G.; Honorato, L.; Guimaraes, A.J.; Rodrigues, M.L.; Reis, F.C.G.; Vale, A.M.; Ray, A.; Nosanchuk, J.D.; Nimrichter, L. Protective effect of fungal extracellular vesicles against murine candidiasis. *Cell Microbiol.* **2020**, e13238. [[CrossRef](#)]

72. Edwards, J.E., Jr.; Schwartz, M.M.; Schmidt, C.S.; Sobel, J.D.; Nyirjesy, P.; Schodel, F.; Marchus, E.; Lizakowski, M.; DeMontigny, E.A.; Hoeg, J.; et al. A Fungal Immunotherapeutic Vaccine (NDV-3A) for Treatment of Recurrent Vulvovaginal Candidiasis—A Phase 2 Randomized, Double-Blind, Placebo-Controlled Trial. *Clin. Infect. Dis.* **2018**, *66*, 1928–1936. [\[CrossRef\]](#)
73. Phan, Q.T.; Myers, C.L.; Fu, Y.; Sheppard, D.C.; Yeaman, M.R.; Welch, W.H.; Ibrahim, A.S.; Edwards, J.E., Jr.; Filler, S.G. Als3 is a *Candida albicans* invasin that binds to cadherins and induces endocytosis by host cells. *PLoS Biol.* **2007**, *5*, e64. [\[CrossRef\]](#) [\[PubMed\]](#)
74. Zhao, X.; Daniels, K.J.; Oh, S.H.; Green, C.B.; Yeater, K.M.; Soll, D.R.; Hoyer, L.L. *Candida albicans* Als3p is required for wild-type biofilm formation on silicone elastomer surfaces. *Microbiology* **2006**, *152*, 2287–2299. [\[CrossRef\]](#) [\[PubMed\]](#)
75. Ibrahim, A.S.; Luo, G.; Gebremariam, T.; Lee, H.; Schmidt, C.S.; Hennessey, J.P., Jr.; French, S.W.; Yeaman, M.R.; Filler, S.G.; Edwards, J.E., Jr. NDV-3 protects mice from vulvovaginal candidiasis through T- and B-cell immune response. *Vaccine* **2013**, *31*, 5549–5556. [\[CrossRef\]](#) [\[PubMed\]](#)
76. Lin, L.; Ibrahim, A.S.; Avanesian, V.; Edwards, J.E., Jr.; Fu, Y.; Baquir, B.; Taub, R.; Spellberg, B. Considerable differences in vaccine immunogenicities and efficacies related to the diluent used for aluminum hydroxide adjuvant. *Clin. Vaccine Immunol.* **2008**, *15*, 582–584. [\[CrossRef\]](#) [\[PubMed\]](#)
77. Spellberg, B.; Ibrahim, A.S.; Lin, L.; Avanesian, V.; Fu, Y.; Lipke, P.; Otoo, H.; Ho, T.; Edwards, J.E., Jr. Antibody titer threshold predicts anti-candidal vaccine efficacy even though the mechanism of protection is induction of cell-mediated immunity. *J. Infect. Dis.* **2008**, *197*, 967–971. [\[CrossRef\]](#)
78. Schmidt, C.S.; White, C.J.; Ibrahim, A.S.; Filler, S.G.; Fu, Y.; Yeaman, M.R.; Edwards, J.E., Jr.; Hennessey, J.P., Jr. NDV-3, a recombinant alum-adjuvanted vaccine for *Candida* and *Staphylococcus aureus*, is safe and immunogenic in healthy adults. *Vaccine* **2012**, *30*, 7594–7600. [\[CrossRef\]](#)
79. Uppuluri, P.; Singh, S.; Alqarihi, A.; Schmidt, C.S.; Hennessey, J.P., Jr.; Yeaman, M.R.; Filler, S.G.; Edwards, J.E.; Ibrahim, A.S. Human Anti-Als3p Antibodies Are Surrogate Markers of NDV-3A Vaccine Efficacy Against Recurrent Vulvovaginal Candidiasis. *Front. Immunol.* **2018**, *9*, 1349. [\[CrossRef\]](#)
80. Alqarihi, A.; Singh, S.; Edwards, J.E., Jr.; Ibrahim, A.S.; Uppuluri, P. NDV-3A vaccination prevents *C. albicans* colonization of jugular vein catheters in mice. *Sci. Rep.* **2019**, *9*, 6194. [\[CrossRef\]](#)
81. Satoh, K.; Makimura, K.; Hasumi, Y.; Nishiyama, Y.; Uchida, K.; Yamaguchi, H. *Candida auris* sp. nov., a novel ascomycetous yeast isolated from the external ear canal of an inpatient in a Japanese hospital. *Microbiol. Immunol.* **2009**, *53*, 41–44. [\[CrossRef\]](#)
82. Lockhart, S.R.; Etienne, K.A.; Vallabhaneni, S.; Farooqi, J.; Chowdhary, A.; Govender, N.P.; Colombo, A.L.; Calvo, B.; Cuomo, C.A.; Desjardins, C.A.; et al. Simultaneous Emergence of Multidrug-Resistant *Candida auris* on 3 Continents Confirmed by Whole-Genome Sequencing and Epidemiological Analyses. *Clin. Infect. Dis.* **2017**, *64*, 134–140. [\[CrossRef\]](#)
83. Zamith-Miranda, D.; Heyman, H.M.; Cleare, L.G.; Couvillion, S.P.; Clair, G.C.; Bredeweg, E.L.; Gacser, A.; Nimrichter, L.; Nakayasu, E.S.; Nosanchuk, J.D. Multi-omics Signature of *Candida auris*, an Emerging and Multidrug-Resistant Pathogen. *mSystems* **2019**, *4*. [\[CrossRef\]](#) [\[PubMed\]](#)
84. Singh, S.; Uppuluri, P.; Mamouei, Z.; Alqarihi, A.; Elhassan, H.; French, S.; Lockhart, S.R.; Chiller, T.; Edwards, J.E., Jr.; Ibrahim, A.S. The NDV-3A vaccine protects mice from multidrug resistant *Candida auris* infection. *PLoS Pathog.* **2019**, *15*, e1007460. [\[CrossRef\]](#)
85. De Almeida, J.R.F.; Jannuzzi, G.P.; Kaihami, G.H.; Breda, L.C.D.; Ferreira, K.S.; de Almeida, S.R. An immunoproteomic approach revealing peptides from *Sporothrix brasiliensis* that induce a cellular immune response in subcutaneous sporotrichosis. *Sci. Rep.* **2018**, *8*, 4192. [\[CrossRef\]](#) [\[PubMed\]](#)
86. Yu, X.; Wan, Z.; Zhang, Z.; Li, F.; Li, R.; Liu, X. Phenotypic and molecular identification of *Sporothrix* isolates of clinical origin in Northeast China. *Mycopathologia* **2013**, *176*, 67–74. [\[CrossRef\]](#) [\[PubMed\]](#)
87. Chen, F.; Jiang, R.; Wang, Y.; Zhu, M.; Zhang, X.; Dong, S.; Shi, H.; Wang, L. Recombinant Phage Elicits Protective Immune Response against Systemic *S. globosa* Infection in Mouse Model. *Sci. Rep.* **2017**, *7*, 42024. [\[CrossRef\]](#) [\[PubMed\]](#)
88. Tarcha, E.J.; Basrur, V.; Hung, C.Y.; Gardner, M.J.; Cole, G.T. A recombinant aspartyl protease of *Coccidioides posadasii* induces protection against pulmonary coccidioidomycosis in mice. *Infect. Immun.* **2006**, *74*, 516–527. [\[CrossRef\]](#)
89. Tarcha, E.J.; Basrur, V.; Hung, C.Y.; Gardner, M.J.; Cole, G.T. Multivalent recombinant protein vaccine against coccidioidomycosis. *Infect. Immun.* **2006**, *74*, 5802–5813. [\[CrossRef\]](#)

90. Hurtgen, B.J.; Hung, C.Y.; Ostroff, G.R.; Levitz, S.M.; Cole, G.T. Construction and evaluation of a novel recombinant T cell epitope-based vaccine against Coccidioidomycosis. *Infect. Immun.* **2012**, *80*, 3960–3974. [\[CrossRef\]](#)
91. Hung, C.Y.; Zhang, H.; Castro-Lopez, N.; Ostroff, G.R.; Khoshlenar, P.; Abraham, A.; Cole, G.T.; Negron, A.; Forsthuber, T.; Peng, T.; et al. Glucan-Chitin Particles Enhance Th17 Response and Improve Protective Efficacy of a Multivalent Antigen (rCpa1) against Pulmonary Coccidioides posadasii Infection. *Infect. Immun.* **2018**, *86*. [\[CrossRef\]](#)
92. Awasthi, S.; Awasthi, V.; Magee, D.M.; Coalson, J.J. Efficacy of antigen 2/proline-rich antigen cDNA-transfected dendritic cells in immunization of mice against Coccidioides posadasii. *J. Immunol.* **2005**, *175*, 3900–3906. [\[CrossRef\]](#)
93. Awasthi, S. Dendritic cell-based vaccine against coccidioides infection. *Ann. N. Y. Acad. Sci.* **2007**, *1111*, 269–274. [\[CrossRef\]](#) [\[PubMed\]](#)
94. Vilekar, P.; Awasthi, V.; Lagisetty, P.; King, C.; Shankar, N.; Awasthi, S. In vivo trafficking and immunostimulatory potential of an intranasally-administered primary dendritic cell-based vaccine. *BMC Immunol.* **2010**, *11*, 60. [\[CrossRef\]](#) [\[PubMed\]](#)
95. Awasthi, S.; Vilekar, P.; Conkleton, A.; Rahman, N. Dendritic cell-based immunization induces Coccidioides Ag2/PRA-specific immune response. *Vaccine* **2019**, *37*, 1685–1691. [\[CrossRef\]](#) [\[PubMed\]](#)
96. Awasthi, S. Intranasal Antifungal Vaccination Using DNA-Transfected Dendritic Cells. *Methods Mol. Biol.* **2017**, *1625*, 75–83. [\[CrossRef\]](#)
97. Wuthrich, M.; Brandhorst, T.T.; Sullivan, T.D.; Filutowicz, H.; Sterkel, A.; Stewart, D.; Li, M.; Lerksuthirat, T.; LeBert, V.; Shen, Z.T.; et al. Calnexin induces expansion of antigen-specific CD4(+) T cells that confer immunity to fungal ascomycetes via conserved epitopes. *Cell Host Microbe* **2015**, *17*, 452–465. [\[CrossRef\]](#)
98. Bowman, S.M.; Free, S.J. The structure and synthesis of the fungal cell wall. *Bioessays* **2006**, *28*, 799–808. [\[CrossRef\]](#)
99. Kumaresan, P.R.; Manuri, P.R.; Albert, N.D.; Maiti, S.; Singh, H.; Mi, T.; Roszik, J.; Rabinovich, B.; Olivares, S.; Krishnamurthy, J.; et al. Bioengineering T cells to target carbohydrate to treat opportunistic fungal infection. *Proc. Natl. Acad. Sci. USA* **2014**, *111*, 10660–10665. [\[CrossRef\]](#)
100. Kumaresan, P.R.; da Silva, T.A.; Kontoyiannis, D.P. Methods of Controlling Invasive Fungal Infections Using CD8(+) T Cells. *Front. Immunol.* **2017**, *8*, 1939. [\[CrossRef\]](#)
101. Shikanai-Yasuda, M.A.; Mendes, R.P.; Colombo, A.L.; Telles, F.Q.; Kono, A.; Paniago, A.M.M.; Nathan, A.; Valle, A.; Bagagli, E.; Benard, G.; et al. Brazilian guidelines for the clinical management of paracoccidioidomycosis. *Epidemiol. Serv. Saude* **2018**, *27*, e0500001. [\[CrossRef\]](#)
102. Fries, B.C.; Taborda, C.P.; Serfass, E.; Casadevall, A. Phenotypic switching of Cryptococcus neoformans occurs in vivo and influences the outcome of infection. *J. Clin. Investig.* **2001**, *108*, 1639–1648. [\[CrossRef\]](#)
103. Taborda, C.P.; Rivera, J.; Zaragoza, O.; Casadevall, A. More is not necessarily better: Prozone-like effects in passive immunization with IgG. *J. Immunol.* **2003**, *170*, 3621–3630. [\[CrossRef\]](#) [\[PubMed\]](#)
104. Taborda, C.P.; Casadevall, A. Immunoglobulin M efficacy against Cryptococcus neoformans: Mechanism, dose dependence, and prozone-like effects in passive protection experiments. *J. Immunol.* **2001**, *166*, 2100–2107. [\[CrossRef\]](#)
105. Casadevall, A.; Pirofski, L.A. A new synthesis for antibody-mediated immunity. *Nat. Immunol.* **2011**, *13*, 21–28. [\[CrossRef\]](#)
106. Albuquerque, P.; Casadevall, A. Quorum sensing in fungi—A review. *Med. Mycol.* **2012**, *50*, 337–345. [\[CrossRef\]](#) [\[PubMed\]](#)
107. Bueno, R.A.; Thomaz, L.; Munoz, J.E.; da Silva, C.J.; Nosanchuk, J.D.; Pinto, M.R.; Travassos, L.R.; Taborda, C.P. Antibodies Against Glycolipids Enhance Antifungal Activity of Macrophages and Reduce Fungal Burden After Infection with Paracoccidioides brasiliensis. *Front. Microbiol.* **2016**, *7*, 74. [\[CrossRef\]](#) [\[PubMed\]](#)
108. Thomaz, L.; Nosanchuk, J.D.; Rossi, D.C.; Travassos, L.R.; Taborda, C.P. Monoclonal antibodies to heat shock protein 60 induce a protective immune response against experimental Paracoccidioides lutzii. *Microbes Infect.* **2014**, *16*, 788–795. [\[CrossRef\]](#)
109. Sable, C.A.; Donowitz, G.R. Infections in bone marrow transplant recipients. *Clin. Infect. Dis.* **1994**, *18*, 273–281; quiz 282–284. [\[CrossRef\]](#)
110. Wingard, J.R. Fungal infections after bone marrow transplant. *Biol. Blood Marrow. Transplant.* **1999**, *5*, 55–68. [\[CrossRef\]](#)

111. Denning, D.W. Invasive aspergillosis. *Clin. Infect. Dis.* **1998**, *26*, 781–803; quiz 804–805. [[CrossRef](#)]
112. Thakur, R.; Anand, R.; Tiwari, S.; Singh, A.P.; Tiwary, B.N.; Shankar, J. Cytokines induce effector T-helper cells during invasive aspergillosis; what we have learned about T-helper cells? *Front. Microbiol.* **2015**, *6*, 429. [[CrossRef](#)]
113. Chaudhary, N.; Marr, K.A. Impact of *Aspergillus fumigatus* in allergic airway diseases. *Clin. Transl. Allergy* **2011**, *1*, 4. [[CrossRef](#)] [[PubMed](#)]
114. Shah, A.; Panjabi, C. Allergic Bronchopulmonary Aspergillosis: A Perplexing Clinical Entity. *Allergy Asthma Immunol. Res.* **2016**, *8*, 282–297. [[CrossRef](#)] [[PubMed](#)]
115. Cenci, E.; Mencacci, A.; Bacci, A.; Bistoni, F.; Kurup, V.P.; Romani, L. T cell vaccination in mice with invasive pulmonary aspergillosis. *J. Immunol.* **2000**, *165*, 381–388. [[CrossRef](#)] [[PubMed](#)]
116. Ito, J.I.; Lyons, J.M.; Hong, T.B.; Tamae, D.; Liu, Y.K.; Wilczynski, S.P.; Kalkum, M. Vaccinations with recombinant variants of *Aspergillus fumigatus* allergen Asp f 3 protect mice against invasive aspergillosis. *Infect. Immun.* **2006**, *74*, 5075–5084. [[CrossRef](#)]
117. Liu, M.; Capilla, J.; Johansen, M.E.; Alvarado, D.; Martinez, M.; Chen, V.; Clemons, K.V.; Stevens, D.A. *Saccharomyces* as a vaccine against systemic aspergillosis: ‘the friend of man’ a friend again? *J. Med. Microbiol.* **2011**, *60*, 1423–1432. [[CrossRef](#)]
118. Diaz-Arevalo, D.; Bagramyan, K.; Hong, T.B.; Ito, J.I.; Kalkum, M. CD4+ T cells mediate the protective effect of the recombinant Asp f3-based anti-aspergillosis vaccine. *Infect. Immun.* **2011**, *79*, 2257–2266. [[CrossRef](#)]
119. Rana, A.; Rub, A.; Akhter, Y. Proteome-wide B and T cell epitope repertoires in outer membrane proteins of *Mycobacterium avium* subsp. *paratuberculosis* have vaccine and diagnostic relevance: A holistic approach. *J. Mol. Recognit.* **2015**, *28*, 506–520. [[CrossRef](#)]
120. Vita, R.; Overton, J.A.; Greenbaum, J.A.; Ponomarenko, J.; Clark, J.D.; Cantrell, J.R.; Wheeler, D.K.; Gabbard, J.L.; Hix, D.; Sette, A.; et al. The immune epitope database (IEDB) 3.0. *Nucleic Acids Res.* **2015**, *43*, D405–D412. [[CrossRef](#)]
121. Wang, P.; Sidney, J.; Dow, C.; Mothe, B.; Sette, A.; Peters, B. A systematic assessment of MHC class II peptide binding predictions and evaluation of a consensus approach. *PLoS Comput. Biol.* **2008**, *4*, e1000048. [[CrossRef](#)]
122. Marc, G.; Araniciu, C.; Oniga, S.D.; Vlase, L.; Pirnau, A.; Duma, M.; Marutescu, L.; Chifiriuc, M.C.; Oniga, O. New N-(oxazolylmethyl)-thiazolidinedione Active against *Candida albicans* Biofilm: Potential Als Proteins Inhibitors. *Molecules* **2018**, *23*, 2522. [[CrossRef](#)]
123. Azzazy, E.A.; El-Gammal, N.E.; Metwally, W.S.; Attia, M.H. Role of *Candida Albicans* Germ Tube Antibody in Diagnosis of Invasive Candidiasis in End Stage Liver Disease Patients. *Egypt. J. Immunol.* **2018**, *25*, 161–170. [[PubMed](#)]
124. Lee, J.H.; Kim, Y.G.; Gupta, V.K.; Manoharan, R.K.; Lee, J. Suppression of Fluconazole Resistant *Candida albicans* Biofilm Formation and Filamentation by Methylindole Derivatives. *Front. Microbiol.* **2018**, *9*, 2641. [[CrossRef](#)] [[PubMed](#)]
125. Li, Y.; Sun, L.; Lu, C.; Gong, Y.; Li, M.; Sun, S. Promising Antifungal Targets Against *Candida albicans* Based on Ion Homeostasis. *Front. Cell Infect. Microbiol.* **2018**, *8*, 286. [[CrossRef](#)] [[PubMed](#)]
126. Mochon, A.B.; Cutler, J.E. Is a vaccine needed against *Candida albicans*? *Med. Mycol.* **2005**, *43*, 97–115. [[CrossRef](#)]
127. Cassone, A.; De Bernardis, F.; Torososantucci, A. An outline of the role of anti-*Candida* antibodies within the context of passive immunization and protection from candidiasis. *Curr. Mol. Med.* **2005**, *5*, 377–382. [[CrossRef](#)]
128. De Beer, Z.W.; Duong, T.A.; Wingfield, M.J. The divorce of *Sporothrix* and *Ophiostoma*: Solution to a problematic relationship. *Stud. Mycol.* **2016**, *83*, 165–191. [[CrossRef](#)]
129. Moreira, J.A.; Freitas, D.F.; Lamas, C.C. The impact of sporotrichosis in HIV-infected patients: A systematic review. *Infection* **2015**, *43*, 267–276. [[CrossRef](#)]
130. Barros, M.B.; de Almeida Paes, R.; Schubach, A.O. *Sporothrix schenckii* and Sporotrichosis. *Clin. Microbiol. Rev.* **2011**, *24*, 633–654. [[CrossRef](#)]
131. Aung, A.K.; Teh, B.M.; McGrath, C.; Thompson, P.J. Pulmonary sporotrichosis: Case series and systematic analysis of literature on clinico-radiological patterns and management outcomes. *Med. Mycol.* **2013**, *51*, 534–544. [[CrossRef](#)]

132. Almeida-Paes, R.; de Oliveira, M.M.; Freitas, D.F.; do Valle, A.C.; Zancoppe-Oliveira, R.M.; Gutierrez-Galhardo, M.C. Sporotrichosis in Rio de Janeiro, Brazil: *Sporothrix brasiliensis* is associated with atypical clinical presentations. *PLoS Negl. Trop. Dis.* **2014**, *8*, e3094. [[CrossRef](#)]
133. Gremiao, I.D.; Miranda, L.H.; Reis, E.G.; Rodrigues, A.M.; Pereira, S.A. Zoonotic Epidemic of Sporotrichosis: Cat to Human Transmission. *PLoS Pathog.* **2017**, *13*, e1006077. [[CrossRef](#)]
134. Moussa, T.A.A.; Kadasa, N.M.S.; Al Zahrani, H.S.; Ahmed, S.A.; Feng, P.; Gerrits van den Ende, A.H.G.; Zhang, Y.; Kano, R.; Li, F.; Li, S.; et al. Origin and distribution of *Sporothrix globosa* causing sapronoses in Asia. *J. Med. Microbiol.* **2017**, *66*, 560–569. [[CrossRef](#)] [[PubMed](#)]
135. De Almeida, J.R.; Kaihama, G.H.; Jannuzzi, G.P.; de Almeida, S.R. Therapeutic vaccine using a monoclonal antibody against a 70-kDa glycoprotein in mice infected with highly virulent *Sporothrix schenckii* and *Sporothrix brasiliensis*. *Med. Mycol.* **2015**, *53*, 42–50. [[CrossRef](#)] [[PubMed](#)]
136. Nascimento, R.C.; Espindola, N.M.; Castro, R.A.; Teixeira, P.A.; Loureiro y Penha, C.V.; Lopes-Bezerra, L.M.; Almeida, S.R. Passive immunization with monoclonal antibody against a 70-kDa putative adhesin of *Sporothrix schenckii* induces protection in murine sporotrichosis. *Eur. J. Immunol.* **2008**, *38*, 3080–3089. [[CrossRef](#)] [[PubMed](#)]
137. Cole, G.T.; Xue, J.M.; Okeke, C.N.; Tarcha, E.J.; Basrur, V.; Schaller, R.A.; Herr, R.A.; Yu, J.J.; Hung, C.Y. A vaccine against coccidioidomycosis is justified and attainable. *Med. Mycol.* **2004**, *42*, 189–216. [[CrossRef](#)]
138. Galgiani, J.N.; Ampel, N.M.; Blair, J.E.; Catanzaro, A.; Geertsma, F.; Hoover, S.E.; Johnson, R.H.; Kusne, S.; Lisse, J.; MacDonald, J.D.; et al. Infectious Diseases Society of America (IDSA) Clinical Practice Guideline for the Treatment of Coccidioidomycosis. *Clin. Infect. Dis.* **2016**, *63*, e112–e146. [[CrossRef](#)]
139. Freedman, M.; Jackson, B.R.; McCotter, O.; Benedict, K. Coccidioidomycosis Outbreaks, United States and Worldwide, 1940–2015. *Emerg. Infect. Dis.* **2018**, *24*, 417–423. [[CrossRef](#)]
140. Tsang, C.A.; Tabnak, F.; Vugia, D.J.; Benedict, K.; Chiller, T.; Park, B.J.; Centers for Disease Control and Prevention (CDC). Increase in reported coccidioidomycosis—United States, 1998–2011. *MMWR Morb. Mortal. Wkly. Rep.* **2013**, *62*, 217–221.
141. Thompson, G.R., 3rd. Pulmonary coccidioidomycosis. *Semin. Respir. Crit. Care Med.* **2011**, *32*, 754–763. [[CrossRef](#)]
142. Litvintseva, A.P.; Marsden-Haug, N.; Hurst, S.; Hill, H.; Gade, L.; Driebe, E.M.; Ralston, C.; Roe, C.; Barker, B.M.; Goldoft, M.; et al. Valley fever: Finding new places for an old disease: *Coccidioides immitis* found in Washington State soil associated with recent human infection. *Clin. Infect. Dis.* **2015**, *60*, e1–e3. [[CrossRef](#)]
143. Pappagianis, D. Evaluation of the protective efficacy of the killed *Coccidioides immitis* spherule vaccine in humans. The Valley Fever Vaccine Study Group. *Am. Rev. Respir. Dis.* **1993**, *148*, 656–660. [[CrossRef](#)] [[PubMed](#)]
144. Williams, P.L.; Sable, D.L.; Sorgen, S.P.; Pappagianis, D.; Levine, H.B.; Brodine, S.K.; Brown, B.W.; Grumet, F.C.; Stevens, D.A. Immunologic responsiveness and safety associated with the *Coccidioides immitis* spherule vaccine in volunteers of white, black, and Filipino ancestry. *Am. J. Epidemiol.* **1984**, *119*, 591–602. [[CrossRef](#)] [[PubMed](#)]
145. Pappagianis, D.; Levine, H.B. The present status of vaccination against coccidioidomycosis in man. *Am. J. Epidemiol.* **1975**, *102*, 30–41. [[CrossRef](#)] [[PubMed](#)]
146. Helal, M.; Allen, K.J.H.; van Dijk, B.; Nosanchuk, J.D.; Snead, E.; Dadachova, E. Radioimmunotherapy of Blastomycosis in a Mouse Model with a (1→3)-beta-Glucans Targeting Antibody. *Front. Microbiol.* **2020**, *11*, 147. [[CrossRef](#)] [[PubMed](#)]
147. Nosanchuk, J.D.; Dadachova, E. Radioimmunotherapy of fungal diseases: The therapeutic potential of cytotoxic radiation delivered by antibody targeting fungal cell surface antigens. *Front. Microbiol.* **2011**, *2*, 283. [[CrossRef](#)]
148. Bryan, R.A.; Guimaraes, A.J.; Hopcraft, S.; Jiang, Z.; Bonilla, K.; Morgenstern, A.; Bruchertseifer, F.; Del Poeta, M.; Torosantucci, A.; Cassone, A.; et al. Toward developing a universal treatment for fungal disease using radioimmunotherapy targeting common fungal antigens. *Mycopathologia* **2012**, *173*, 463–471. [[CrossRef](#)] [[PubMed](#)]
149. Baxevanis, C.N.; Perez, S.A.; Papamichail, M. Cancer immunotherapy. *Crit. Rev. Clin. Lab. Sci.* **2009**, *46*, 167–189. [[CrossRef](#)] [[PubMed](#)]

150. Seif, M.; Einsele, H.; Löffler, J. CAR T Cells Beyond Cancer: Hope for Immunomodulatory Therapy of Infectious Diseases. *Front. Immunol.* **2019**, *10*, 2711. [[CrossRef](#)] [[PubMed](#)]
151. Dotti, G.; Gottschalk, S.; Savoldo, B.; Brenner, M.K. Design and development of therapies using chimeric antigen receptor-expressing T cells. *Immunol. Rev.* **2014**, *257*, 107–126. [[CrossRef](#)]
152. Johnson, L.A.; June, C.H. Driving gene-engineered T cell immunotherapy of cancer. *Cell Res.* **2017**, *27*, 38–58. [[CrossRef](#)]
153. Zhen, A.; Peterson, C.W.; Carrillo, M.A.; Reddy, S.S.; Youn, C.S.; Lam, B.B.; Chang, N.Y.; Martin, H.A.; Rick, J.W.; Kim, J.; et al. Long-term persistence and function of hematopoietic stem cell-derived chimeric antigen receptor T cells in a nonhuman primate model of HIV/AIDS. *PLoS Pathog.* **2017**, *13*, e1006753. [[CrossRef](#)] [[PubMed](#)]
154. Leibman, R.S.; Richardson, M.W.; Ellebrecht, C.T.; Maldini, C.R.; Glover, J.A.; Secreto, A.J.; Kulikovskaya, I.; Lacey, S.F.; Akkina, S.R.; Yi, Y.; et al. Supraphysiologic control over HIV-1 replication mediated by CD8 T cells expressing a re-engineered CD4-based chimeric antigen receptor. *PLoS Pathog.* **2017**, *13*, e1006613. [[CrossRef](#)] [[PubMed](#)]
155. Ghanem, M.H.; Bolivar-Wagers, S.; Dey, B.; Hajducski, A.; Vargas-Inchaustegui, D.A.; Danielson, D.T.; Bundoc, V.; Liu, L.; Berger, E.A. Bispecific chimeric antigen receptors targeting the CD4 binding site and high-mannose Glycans of gp120 optimized for anti-human immunodeficiency virus potency and breadth with minimal immunogenicity. *Cytotherapy* **2018**, *20*, 407–419. [[CrossRef](#)] [[PubMed](#)]
156. Hale, M.; Mesojednik, T.; Romano Ibarra, G.S.; Sahni, J.; Bernard, A.; Sommer, K.; Scharenberg, A.M.; Rawlings, D.J.; Wagner, T.A. Engineering HIV-Resistant, Anti-HIV Chimeric Antigen Receptor T Cells. *Mol. Ther.* **2017**, *25*, 570–579. [[CrossRef](#)] [[PubMed](#)]
157. Anthony-Gonda, K.; Bardhi, A.; Ray, A.; Flerin, N.; Li, M.; Chen, W.; Ochsenbauer, C.; Kappes, J.C.; Krueger, W.; Worden, A.; et al. Multispecific anti-HIV duoCAR-T cells display broad in vitro antiviral activity and potent in vivo elimination of HIV-infected cells in a humanized mouse model. *Sci. Transl. Med.* **2019**, *11*. [[CrossRef](#)] [[PubMed](#)]
158. Festag, M.M.; Festag, J.; Frassle, S.P.; Asen, T.; Sacherl, J.; Schreiber, S.; Muck-Hausl, M.A.; Busch, D.H.; Wisskirchen, K.; Protzer, U. Evaluation of a Fully Human, Hepatitis B Virus-Specific Chimeric Antigen Receptor in an Immunocompetent Mouse Model. *Mol. Ther.* **2019**, *27*, 947–959. [[CrossRef](#)]
159. Proff, J.; Brey, C.U.; Ensser, A.; Holter, W.; Lehner, M. Turning the tables on cytomegalovirus: Targeting viral Fc receptors by CARs containing mutated CH2-CH3 IgG spacer domains. *J. Transl. Med.* **2018**, *16*, 26. [[CrossRef](#)]
160. Rodrigues, M.L.; Nosanchuk, J.D. Fungal diseases as neglected pathogens: A wake-up call to public health officials. *PLoS Negl. Trop. Dis.* **2020**, *14*, e0007964. [[CrossRef](#)]
161. Bongomin, F.; Gago, S.; Oladele, R.O.; Denning, D.W. Global and Multi-National Prevalence of Fungal Diseases-Estimate Precision. *J. Fungi* **2017**, *3*, 57. [[CrossRef](#)]
162. Rodrigues, M.L.; Albuquerque, P.C. Searching for a change: The need for increased support for public health and research on fungal diseases. *PLoS Negl. Trop. Dis.* **2018**, *12*, e0006479. [[CrossRef](#)]
163. Casadevall, A. Fungal Diseases in the 21st Century: The Near and Far Horizons. *Pathog. Immun.* **2018**, *3*, 183–196. [[CrossRef](#)] [[PubMed](#)]
164. Yates, J.R.; Ruse, C.I.; Nakorchevsky, A. Proteomics by mass spectrometry: Approaches, advances, and applications. *Annu. Rev. Biomed. Eng.* **2009**, *11*, 49–79. [[CrossRef](#)] [[PubMed](#)]

