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Systematic Review of Genus *Cerradomys* Weksler, Percequillo and Voss, 2006 (Rodentia: Cricetidae: Sigmodontinae: Oryzomyini), with Description of Two New Species from Eastern Brazil

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Systematic Review of Genus *Cerradomys* Weksler, Percequillo and Voss, 2006 (Rodentia: Cricetidae: Sigmodontinae: Oryzomyini), with Description of Two New Species from Eastern Brazil

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ABSTRACT

*Cerradomys* is a monophyletic genus that includes four known species, *Cerradomys subflavus*, *C. maracajuensis*, *C. marinhus*, and *C. scotti*, distributed throughout the open vegetation belt across South America, from northeastern Brazil to southeastern Bolivia, and from eastern to northwestern Paraguay. We revised the status of the species currently assigned to this genus by analyzing skins, skulls, karyotypes, and cytochrome b DNA sequences. We also described two novel species, one distributed in the Brazilian states of Minas Gerais, Bahia, and Sergipe, and the other in the states of Paraíba, Pernambuco, Piauí, Ceará, and Maranhão. Molecular analysis suggested the following phylogenetic arrangement: (((*C. subflavus*—*C. sp.n.2*) *C. sp.n.1*) *C. scotti*)((*C. marinhus*—*C. maracajuensis*)). Apparently, both novel species inhabit the Catinga domain and penetrated the coastal Atlantic rainforest, differing from the remaining congeneric species that are typical open-area inhabitants.

RESUMO

O gênero *Cerradomys* é uma entidade monofilética que reúne quatro espécies, *Cerradomys subflavus*, *C. maracajuensis*, *C. marinhus*, e *C. scotti*, distribuídas através do cinturão de vegetação

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seca que divide a América do Sul do nordeste do Brasil ao sudeste da Bolívia e noroeste do Paraguai. Baseados no estudo de peles, crânios, cariótipos e sequências de nucleotídeos de ADN, nós revisamos as espécies atualmente atribuídas ao gênero e reconhecemos duas espécies novas, cujas distribuições estão atualmente limitadas aos estados brasileiros de Minas Gerais, Bahia e Sergipe, e aos estados de Paraíba, Pernambuco, Piauí, Ceará e Maranhão, respectivamente. Análises moleculares empregando algoritmos de parcimônia sugerem o seguinte padrão de adjacência entre seis táxons terminais: (((C. subflavus–C. sp.n. 2) C. sp.n.1) C. scotti(C. marinhus–C. maracajuensis)). Aparentemente, as novas espécies são habitantes típicas da Caatinga, que penetram marginalmente na Floresta Atlântica costeira do nordeste do Brasil, diferindo das demais espécies do grupo que são típicas formas de áreas abertas.

INTRODUCTION

Oryzomyini (sensu Weksler, 2006; Weksler et al., 2006) is a diverse and ubiquitous tribe, with 26 genera in South America, ranging from northern Colombia to northern Argentina, inhabiting cis-Andean and trans-Andean lowland and highland forests, montane forests, dry forests, savannas, grasslands, scrubs, and coastal marshes (Musser and Carleton, 2005; Weksler et al., 2006).

Several reports on the systematic of this tribe, with emphasis on the traditional conception of the polyphyletic genus Oryzomys (Musser and Carleton, 2005), resulted in more accurate definitions of genera, species group, and species in the past decade (Weksler, 1996; Musser et al., 1998; Percequillo, 1998, 2003; Bonvicino and Moreira, 2001; Langguth and Bonvicino, 2002; Musser and Carleton, 2005; Weksler, 2003, 2006). These studies led to the recent division of Oryzomys species groups in 10 new oryzomyine genera: Aegialomys, Cerradomys, Ereomeryomys, Euryzomys, Hyllaemys, Mindomys, Nepelomys, Oreyzomys, Sooretamys and Transandinomys (Weksler et al., 2006). However, the species group taxa in several of these genera still remain taxonomically unresolved, with undescribed forms and poorly diagnosed species.

Recently, the genus Cerradomys (formerly Oryzomys subflavus group) has been extensively studied by morphologic, karyologic, and molecular approaches (Bonvicino et al., 1999; Bonvicino and Moreira, 2001; Langguth and Bonvicino, 2002; Bonvicino, 2003; Brooks et al., 2004). These studies allowed for the recognition and description of four new species, considerably altering the species diversity in a group known to include only C. subflavus until 2002.

Currently, the genus Cerradomys comprises four species, namely: C. maracajuensis Langguth and Bonvicino, 2002; C. marinhus Bonvicino, 2003; C. scotti Langguth and Bonvicino, 2002 (including Cerradomys andersoni Brooks et al. 2004; see Emmons et al., 2006); and C. subflavus (Wagner, 1842). The monophyly of this group has been demonstrated in several molecular studies (Bonvicino and Moreira, 2001; Bonvicino, 2003; Brooks et al., 2004), which included cytochrome b DNA haplotypes of all known taxa.

Cerradomys species are distributed throughout an open-vegetation belt, also known as the dry diagonal corridor of South America (Costa, 2003; Bonvicino, 2003). This open-vegetation area crosses South America from northeastern Brazil to southeastern Bolivia and northwestern Paraguay, including the Caatinga, Cerrado, and Chaco domains (Costa, 2003).

According to morphologic (Percequillo, 1998), karyologic (Maiá and Hulak, 1981; Almeida and Yonenaga-Yassuda, 1985; Svartman and Almeida, 1992; Bonvicino et al., 1999; Andrades-Miranda et al, 2002), and molecular evidence (Bonvicino and Moreira, 2001, Bonvicino, 2003), Cerradomys was considered very diverse, still presenting undescribed forms (see Bonvicino and Moreira, 2001; Bonvicino, 2003). Here we review the current recognized species of genus Cerradomys and describe two new species from eastern Brazil, based on morphologic, morphometric, and karyologic evidence. Moreover, we also show the phylogenetic relationship within Cerradomys based on cytochrome b DNA sequence data.

MATERIAL AND METHODS

SPECIMENS: We studied skins, skulls, skeletons, and fluid-preserved specimens deposited in the following collections:
AMNH American Museum of Natural History, New York
APC uncatalogued specimens collected by Ana Paula Carmignotto (MZUSP)
ARP uncatalogued specimens collected by Alexandre Reis Percequillo (MZUSP)
BMNH The Natural History Museum, London
CBF Coleccion Boliviana da Fauna, La Paz
CRB uncatalogued specimens collected by Cibele R. Bonvicino (MN)
FMNH The Field Museum, Chicago
GD uncatalogued specimens collected by Guilhermo D’Elia (MHNM)
JD uncatalogued specimens collected by Julio Dalponte (UFMT)
LPC uncatalogued specimens collected by Leonora Pires Costa (UFMT)
LV Laboratório de Vertebrados, UFRJ, Rio de Janeiro
MHNM Museo de Historia Natural de Montevideo, Montevideo
MLP Museu La Plata, La Plata
MN Museu Nacional da Universidade Federal do Rio de Janeiro, Rio de Janeiro
MVZ Museum of Vertebrate Zoology, Berkeley
MW uncatalogued specimens collected by Marcelo Weksler (MN)
MZUSP Museu de Zoologia da Universidade de São Paulo, São Paulo
RM uncatalogued specimens collected by Raquel Moura (UFMG)
RP uncatalogued specimens collected by Renata Pardini (MZUSP)
UFMG Departamento de Zoologia da Universidade Federal de Minas Gerais, Belo Horizonte
UFMT Universidade Federal de Mato Grosso, Cuiabá
UFPB Departamento de Sistemática e Ecologia, Universidade Federal da Paraíba, João Pessoa
UMMZ University of Michigan, Museum of Zoology, Ann Arbor
UnB Departamento de Zoologia, Universidade de Brasília, Distrito Federal
USNM National Museum of Natural History, Washington, DC
YL uncatalogued specimens collected by Yuri Leite (UFMG)

A gazetteer of collecting localities is furnished in appendix 1.

Lists of specimens examined are provided in the species account, along with species descriptions with synonymy, type locality, geographic distribution, diagnosis, karyology, and natural history. Uppercase letters preceding museum number refer to sex of voucher specimens, M for males, F for females and I for nonsexed specimens. Lists of examined specimens are arranged in alphabetical order, by country, state or province, and locality of collection.

**EXTERNAL AND CRANIAL MEASUREMENTS:**
All measurements are expressed in millimeters (mm) except weight, which is expressed in grams (g). The following external measurements were obtained from specimen tags or from wild-caught specimens during fieldwork:

- **TL** Total length
- **LT** Length of the tail
- **Ear** Pinnae length
- **HF** Length of hind foot
- **Wt** Weight

Head-and-body length (HBL) was obtained by subtracting length of tail from total length.

Fifteen cranial measurements (based on Langguth, 1963; Voss, 1988; Brandt and Pessôa, 1994; Musser et al., 1998) were obtained with digital calipers, to the nearest 0.01 mm:

- **CIL** Condylo-incisive length, measured from the greater curvature of one upper incisor to the articular surface of the occipital condyle on the same side
- **LD** Length of diastema, from the crown of the first upper molar to the lesser curvature of the upper incisor on the same side
- **LM** Length of molars, crown length from M1 to M3
- **LIF** Length of incisive foramen, greatest anterior-posterior dimension of one incisive foramen
- **PB** Palatal breadth, measured at the labial margin of maxillary bone across the third molars
- **BR** Breadth of rostrum, greatest dimension measured across the external border of the nasolacrimal capsules
- **LN** Length of nasals, greatest anterior-posterior dimension of one nasal bone
- **LPB** Length of palatal bridge, measured from the posterior border of the
incisive foramen to the anterior border of the mesopterygoid fossa

HB Height of braincase, greatest height of braincase, measured from the sphenoccipital suture to the frontoparietal suture

LIB Least interorbital breadth, least distance across the frontal bones

ZB Zygomatic breadth, greatest dimension across the squamosal root of zygomatic arches

CZL Condylo-zygomatic length, measured from the occipital condyle to the posteriormost edge of the zygomatic notch

OFL Orbital fossa length, greatest length of the orbital fossa between the squamosal and maxillary roots of the zygomatic arch

BB Bullar breadth, greatest breadth from the petrosal-basioccipital suture to the dorsal process of the ectotympanic

**AGE CRITERIA AND ANATOMY:** We followed the terminology and definitions employed by Musser et al. (1998) for age classes, and those defined by Carleton (1973), Reig (1977), Voss (1988, 1993), Voss and Carleton (1993), Steppan (1995), Voss et al. (2001), and Weksler (2006) for the external and cranial measurements, anatomical traits, and molar nomenclature.

**STATISTICAL ANALYSES:** We quantitatively compared adults from all species assigned to *Cerradomys*. Univariate comparisons among the *C. subflavus* and the two new species were performed through one-way analysis of variance (one-way ANOVA) of body and skull measurements. Principal components analysis was also performed, using the 15 log-transformed cranial measurements and the covariance matrix. We explored morphometric differentiation among the three species through a discriminant analysis of the log-transformed cranial measurements. Statistical procedures are detailed in Neff and Marcus (1980) and Sokal and Rohlf (1995). All the reported results were obtained using SAS v.8.02.

**CYTOGENETICS:** Chromosome preparations were obtained from short-term cell cultures. Bone marrow was directly cultured for two hours (around 37°C) in Falcon tubes containing sterile medium (80% RPMI, 20% fetal calf serum, 5 μg/ml of ethidium bromide and 10^-6 M colchicine). G-banding was carried out for identifying chromosomal homologies as described by Seabright (1971). Chromosomes were ordered according to morphology and decreasing size.

**PHYLOGENETIC ANALYSIS:** Phylogenetic reconstructions were carried out with DNA-sequence data of the mitochondrial gene cytochrome *b*. We analyze the two new species herein described (GenBank accession number AF181275, museum specimens MN 35898 and LV–FC 148; and AF181276, museum specimen MN 96786 [AL 3655]), plus *C. scotti* (AF181277—comprising specimens MN50379 and MN61677 sharing the same haplotype), *C. maracajuensis* (AF181278 – MN44178), *C. marinhus* (AF181278 – MN63824) and *C. subflavus* (AF181274 – CEG42), *Nectomys squamipes* (AF181283 – MN42685), *Nectomys garleppii* (U03539 – MVZ147667), and *Sooretamys angouya* (AF181281 – CRB1273). *Neotoma albigula* (AF108704 – MVZ 147667) and *Scotinomys teguina* (AF108705 – UMMZ 3373) were used as outgroups. *Cerradomys andersoni* could not be included in the present analyses due to lack of available nucleotide sequence data in GenBank.

**MEGA 3.1** (Kumar et al., 1993) was used to estimate *p* distances between haplotypes. Parsimony analysis was carried out by branch-and-bound search, with all characters equally weighted. Bootstrap values were estimated on the basis of 1,000 replicates (PAUP* 4.0, Swofford, 2003).

**TAXONOMIC ACCOUNTS**

*Cerradomys* Weksler,
Percequillo and Voss, 2006

**TYPE SPECIES:** *Hesperomys subflavus* Wagner, 1842.

**CONTENTS:** *Cerradomys maracajuensis* Langguth and Bonvicino, 2002; *Cerradomys marinhus* Bonvicino, 2003; *Cerradomys scotti* Langguth and Bonvicino, 2002 (including *C. andersoni*, Brooks et al., 2004); *Cerradomys subflavus* (Wagner, 1842); and two new species below described (see Comments).

**GEOGRAPHIC DISTRIBUTION:** Species attributed to *Cerradomys* are usually associated to the diagonal belt of open and drier vegetation formed mainly by the Chaco,
Cerrado, Pantanal, and Caatinga biomes, from northeastern Brazil to eastern Paraguay and central Bolivia (figs. 1, 2). In northeastern Brazil, from Bahia to Paraíba states, there are also samples associated with coastal Atlantic Forest.

**Diagnosis:** *Cerradomys* species exhibit a long, dense, lax, and coarsely grizzled dorsal pelage (wool hairs: 6–12 mm; cover hairs: 9–17 mm; guard hairs: 13–22 mm); tail longer than head and body length combined (110%–123% of head and body length); incisive foramina very long, with lateral margins wider medially and anteroposterior margins sharp; interorbital region strongly convergent anteriorly, with well-developed supraorbital crests; complex posterolateral palatal pits recessed at very deep and wide fossae (except in *Cerradomys maracajuensis* and *C. marinhus*); long and wide sphenopalatine vacuities present (except in *Cerradomys maracajuensis* and *C. marinhus*); stapedial foramen and posterior opening of alisphenoid canal absent or small, squamosal-alisphenoid groove and sphenofrontal foramen absent, and secondary branch crosses dorsal surface of pterygoid plate (pattern 3 of Voss, 1988); capsular process of lower incisor present; phallus with reduced cartilaginous baculum, two-digitated, with central digit absent; cartilaginous baculum situated outside the glans penis body; bony baculum extremely elongated; length of cartilaginous baculum about 1/8 of the length of osseous baculum (for more diagnostic traits, see also Weksler et al., 2006).

**Morphological Description:** Large body size (HBL range, 119–185 mm; n = 305) and tail much longer than head and body length (TL range, 131–227 mm; n = 297); hind-feet robust, presenting a considerable span of size (HFL range, 20–43 mm in length; n = 314). Pinnae small (ear range, 15–27 mm; n = 321). Dorsal pelage variably short to long, and dense; wool hairs thin, long and wavy (range: 5–12 mm); cover hairs longer and thicker on distal half (range, 9–17 mm); guard hairs much longer, stiffer, and thicker on its distal third (range, 13–22 mm). Dorsal body pelage uniform from head to rump, coarsely grizzled, varying from buffy yellow grizzled with dark brown to buffy orange-red grizzled with black; in some species, head distinctly colored from body (head grayish and body orange to reddish, in *C. subflavus* and the two new species). Ventral pelage short composed by wool, cover and guard hairs (similar, but shorter to dorsal hairs). Ventral pelage ranges from white to buff grizzled with gray (ventral hairs always gray-based). Body flanks buffy yellow to reddish orange. Mystacial vibsvisae very dense and long, barely surpassing the ears when laid back; dorsal mystacial vibrissae dark brown with golden tips and ventral entirely white. Tail covered by short hairs (apparently almost naked) or long hairs (hirsute); tail uniformly colored (*C. marinhus* and *C. maracajuensis*), weakly bicolored (*C. subflavus*) or distinctly bicolored (*C. scotti*). Dorsal surface of hindfoot densely covered by short hairs; ungual tufts dense, not concealing the claws. Pinnae small, densely covered by short hairs, both internally and externally; pinnae hairs brown, with buffy or orange tips. Four pairs of mammae present, in inguinal, abdominal, postaxial, and pectoral positions.

Skull large and robust (figs. 3–6; CIL range, 28.4–36.6 mm; n = 313). Rostrum long and broad, tapering, flanked by shallow to deep and wide zygomatic notch; rostral fossae moderately to deeply excavated; premaxillae short, not produced anteriorly (not forming with nasals a rostral tube); nasals long, tapering posteriorly, not extending posteriorly beyond lacrimals; interorbital region strongly converging anteriorly, with well-developed supraorbital crests, forming a distinctly projecting shelf; braincase oblong, with temporal crest developed; interparietal broad, nearly equaling caudal border of parietals; lambdoidal crests well developed and sharp, and occipital region with developed occipital crests. Zygomatic plate (in lateral view) with anterior margin straight or slightly concave; zygomatic spine present in some individuals of some species (e.g., *C. maracajuensis*), but predominantly rounded in shape; lambdoidal crests well developed and sharp, and occipital region with developed occipital crests. Zygomatic plate ranging from narrow to broad (BZP range, 2.8–4.4 mm; n = 344). Zygomatic arches strong and slightly divergent posteriorly; jugal present (separating the maxillary and squamosal ramus of zygomatic arch). Incisive foramina very long (LIF range, 6.1–8.6 mm; n = 344; occupying ca. 75% of the length of diastema in all species), with lateral margins...
Fig. 1. Known collection localities of the six species of *Cerradomys* in South America. The area delimited by a square is detailed in figure 2. See gazetteer (appendix 1), where numbers are associated with collection localities. Brazilian states acronyms: AL, Alagoas; BA, Bahia; CE, Ceará; DF, Distrito Federal; Go, Goiás; MA, Maranhão; MT, Mato Grosso; MS, Mato Grosso do Sul; MG, Minas Gerais; PB, Paraíba; PE, Pernambuco; PI, Piauí; SP, São Paulo; SE, Sergipe; TO, Tocantins.
Fig. 2. Detail of map showing known collection localities for five species of *Cerradomys*.
Fig. 3. Dorsal and ventral cranial views of three species of *Cerradomys*. Left: *C. langguthi* from Corredor São João—Fazenda Pacatuba, Paraíba (MN 69786; CIL, 32.1 mm). Middle: *C. vivoi* from Fazenda Canoas, Juramento, Minas Gerais (MN 61663; CIL, 32.92 mm). Right: *C. subflavus* from Parque Nacional da Serra do Cipó, Minas Gerais (MN 31393; CIL, 32.41 mm).
Fig. 4. Dorsal and ventral cranial views of three species of *Cerradomys*. Left: *C. marinhus* from Fazenda Sertão do Formoso, Jaborandi (MN 63834; CIL, 33.39 mm). Middle: *C. maracajuensis* from Maracaju, Mato Grosso do Sul (MN 4376; CIL, 30.83 mm). Right: *C. scotti* from Estação Ecológica Santa Bárbara, São Paulo (MZUSP APC 1157; CIL, 31.38 mm).
Fig. 5. Lateral cranial and mandible views of three species of *Cerradomys*. Top: *C. langguthi* from Corredor São João—Fazenda Pacatuba, Paraíba (MN 69786; CIL, 32.1 mm). Middle: *C. vivoi* from Fazenda Canoas, Juramento, Minas Gerais (MN 61663; CIL, 32.92 mm). Bottom: *C. subflavus* from Parque Nacional da Serra do Cipó, Minas Gerais (MN 31393; CIL, 32.41 mm).
Fig. 6. Lateral cranial and mandible views of three species of *Cerradomys*. Top: *C. marinhus* from Fazenda Sertão do Formoso, Jaborandi (MN 63834; CIL, 33.39 mm). Middle: *C. maracajuensis* from Maracaju, Mato Grosso do Sul (MN 4376; CIL, 30.83 mm). Bottom: *C. scotti* from Estação Ecológica Santa Bárbara, São Paulo (MZUSP APC 1157; CIL, 31.38 mm).
wider medially and anteroposterior margins round to acute; posterior margins of incisive foramina extending to or between M1 alveoli (not extending in some individuals of some species; e.g., C. scotti). Palate with complex posterolateral palatal pits located on deep (in C. maracajuensis and C. marinus) to very deep and wide palatal notches; palatal excrescences variably present. Mesopterygoid fossa narrow to wide; anterior border of mesopterygoid fossa reach the alveolus of M3 in younger individuals (but this condition persists in some adults of some species); bony roof of fossa completely ossified or perforated by small to very large sphenopalatine vacuities present. Parapterygoid plates wide and moderately excavated, without vacuities or fontanelles. Alisphenoid strut variably present (buccinator-masticatory and accessory oval foramina confluent [in C. scotti] or not). Carotid arterial circulation derived, with stapedial foramen and posterior opening of alisphenoid canal absent or small, squamosalalisphenoid groove and sphenofrontal foramina absent, and secondary branch crosses dorsal surface of pterygoid plate (pattern 3 of Voss, 1988). Posglenoid foramen large and rounded to small and narrow; subsquamosal fenestra variably small, vestigial, or absent; hamular process of squamosal wide (indistinct in specimens with obliterated subsquamosal fenestra). Tegmen tympani short, overlapping squamosal or not; posterior suspensory process of squamosal absent. Auditory bulla globose; eustachian tube short to medium, with or without a distinct medial bony lamina dorsal to carotid canal; bony process dorsal to stapedial process of ectotympanic present, overlapped or not to squamosal.

Mandible robust (figs. 5, 6); coronoid process developed, falciform to triangular, nearly equal in height to condyloid process; superior notch shallow; angular process short, variably surpassing the condyloid process posteriorly; inferior notch shallow; capsular projection of lower incisor is present (in adult specimens).

Incisors opisthodont; anterior enamel surface orange. Molars pentalophodont, low-crowned; main cusps arranged in opposite pairs; labial and lingual flexi overlap at median molar plane. M1 with paracone connected medially to protocone (defining a long and obliquely oriented parafossetus); mesoloph long and narrow; metacone posteromedially connected to posteroloph; posteroloph long and narrow. M2 similar to M1; paracone connected medially to protocone; metacone connected medially and posteromedially to posteroloph; mesoloph long and narrow in most species or reduced/absent in other (C. scotti and few individuals of C. maracajuensis); posteroloph long and narrow. M3 small, with metacone-hypocone pair reduced; mesoloph fused to metacone; deep and well-defined hypoflexus. Lower m1 with narrow and undivided anteroconid; protolophid and mesolophid narrow (fusing to anterolingual conulid, entoconid with slight wear, respectively). m2 similar to m1; anterolabial cingulum well developed; protolophid deep; mesolophid long and narrow in most species, reduced or absent in others (C. scotti and C. maracajuensis). m3 with anterolabial cingulum weakly developed; protolophid shallow. M1 with four roots, one anterior, one posterior and two accessory rootlets, in labial and lingual positions; M2 and M3 with three roots. m1 with two roots and two accessory rootlets; m2 and m3 with two roots.

Postcranial axial skeleton formed by 7 cervical, 12 toraxic, 7 lumbar, 3–4 sacral, and 33–37 caudal vertebrae (modal number 36); fifth lumbar vertebrae variably present anapophysis, small or large; hemal arches with conspicuous posterior process variably on vertebrae 1/2, 2/3, 3/4, and 4/5.

Stomach unilocular and hemiglandular (sensu Carleton, 1973), without extension of glandular epithelium into corpus. Phallus elongate and narrow (fig. 7); distal cartilaginous baculum extremely reduced, central digit reduced or absent (two-digitated); cartilaginous baculum situated outside the glans penis body; bony baculum extremely elongated (length of cartilaginous baculum about 1/8 of the length of osseous baculum). Urethral ventral flaps absent; urethral processes without subapical lobules; dorsal papilla spineless; small spines densely ornament the phallus epidermis.

Comparisons: For detailed comparisons with other Oryzomyini genera, see Weksler et al. (2006).

Comments: In addition to the four nominal taxa presently assigned to Cerradomys,
the existence of new species of this genus has been informally mentioned in literature. This was the case of Cerradomys sp.n.2 (= Oryzomys sp.n.2; Bonvicino and Moreira, 2001), which is the same taxon denominated Oryzomys subflavus variant 3 (Bonvicino et al., 1999) and Oryzomys subflavus 2 (Bonvicino, 2003: 82, table 1). And similarly, of Cerradomys sp.n.1 (= Oryzomys sp.n.1; Bonvicino and Moreira, 2001), a synonym of Oryzomys subflavus variant 1 (Bonvicino et al., 1999) and Oryzomys subflavus 3 (Bonvicino, 2003: 82, table 1).

We compared the voucher specimens of Bonvicino et al. (1999), Langguth and Bonvicino (2002), and Bonvicino (2003) to other museum samples. Samples from northeastern Brazil, from the left bank of Rio Sao Francisco, in the states of Pernambuco and Paraiba, Ceará, and Maranhão are herein assigned as a new species, Cerradomys langguthi n. sp., based on the morphological congruence between the specimen karyotyped by Bonvicino and Moreira (2001; Oryzomys sp.n.1; MN 69786, field number AL 3655) and other examined specimens. Specimens from Sergipe, Bahia, and northern Minas Gerais, from the right bank of Rio Sao Francisco, share several morphologic similarities with specimen LV–FC 148 (named Oryzomys sp.n.2) analyzed by Bonvicino and Moreira (2001) and with the new species herein described, Cerradomys vivoi n. sp.

Cerradomys langguthi, new species

Figures 1, 3, 5, 8; tables 1 and 2

Holotype: MN 69786, an adult specimen of unknown sex collected by Alfredo Langguth (original field number AL3655), on December 2, 1995. The holotype consists of a skull with an incomplete right zygomatic arch and a partial postcranial skeleton. A bone marrow suspension of cells in Carnoy’s fixative (methanol:acetic acid) and a liver tissue sample preserved in ethanol are housed at Laboratório de Biologia e Parasitologia de Mamíferos Reservatórios Silvestres, Instituto Oswaldo Cruz–FIOCRUZ, under the original field number AL 3655. Cytochrome b DNA data were deposited in GenBank with the accession number AF181276.

Selected skull dimensions of holotype are: CIL, 32.1; LD, 9.6; LM, 4.95; BM1, 1.45; LIF, 7.15; PB, 5.7; BR, 5.65; LN, 13.55; LPB, 5.2; HB, 10.4; LIB, 5.35; CZL, 24.3; OL, 11.8.

Paratypes: All specimens listed under “Specimens Examined” are herein assigned as paratypes of Cerradomys langguthi.

Type Locality: The holotype of Cerradomys langguthi was collected at Corredor São João–Fazenda Pacatuba, Sapê, State of Paraíba, Brazil, at ca. 07°02’S, 35°09’W. Fazenda São João and Fazenda Pacatuba are Atlantic Forest remnants, which are connected by a corridor of native vegetation.

Distribution: The known collection localities of C. langguthi are distributed on the left bank of Rio São Francisco, throughout the Brazilian states of Pernambuco, Paraíba, Ceará, and Maranhão. In Pernambuco, Paraíba, and Ceará, distributional records extend from coastal lowlands to inland highlands and mountain ranges. Collection records in Maranhão are associated with lowlands of the central portion of the state (fig. 1).

Although the specimen MN 69786 is lacking its skin, it was selected as the holotype, because it is the only available specimen with skull, karyotype, and tissue sample, together the most informative traits for recognition of this species.
Fig. 8. Dorsal, ventral, and lateral cranial views of holotype of *Cerradomys langguthi* (MN 69786; CIL, 32.1 mm).
TABLE 1
Diagnostic Traits and Morphologic Comparisons of the Six Known Species of *Cerradomys*
Abbreviations: w, wool hairs; c, cover hairs; g, guard hairs.

<table>
<thead>
<tr>
<th>Trait</th>
<th><em>langguthi</em></th>
<th><em>maracajuensis</em></th>
<th><em>marinus</em></th>
<th><em>scotti</em></th>
<th><em>subflavus</em></th>
<th><em>vivoi</em></th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Dorsal fur length</strong></td>
<td>w: 5–8 mm; c: 9–12 mm; g: 13–16 mm</td>
<td>w: 11–12 mm; c: 16–17 mm; g: 19–22 mm</td>
<td>w: 11–12 mm; c: 16–17 mm; g: 19–22 mm</td>
<td>w: 9–10 mm; c: 12–15 mm; g: 17–20 mm</td>
<td>w: 9–11 mm; c: 12–14 mm; g: 18–20 mm</td>
<td>w: 6–11 mm; c: 9–15 mm; g: 14–20 mm</td>
</tr>
<tr>
<td><strong>Dorsal body color</strong></td>
<td>orange grizzled with brown</td>
<td>yellow grizzled with brown</td>
<td>yellow grizzled with brown</td>
<td>yellow/buffy intensely grizzled with brown</td>
<td>orange grizzled with brown</td>
<td>orange grizzled with brown</td>
</tr>
<tr>
<td><strong>Head color</strong></td>
<td>grayish, white, grayish white with yellow tones</td>
<td>same as dorsum intense yellow, buffy or grayish buff</td>
<td>same as dorsum intense yellow, buffy or grayish buff</td>
<td>same as dorsum white, grayish white or grayish white with yellow tones</td>
<td>grayish white, grayish white with yellow tones</td>
<td>grayish white, grayish white with yellow tones</td>
</tr>
<tr>
<td><strong>Ventral body color</strong></td>
<td>white, grayish white or grayish white with yellow tones</td>
<td>intense yellow, buffy or grayish buff</td>
<td>intense yellow, buffy or grayish buff</td>
<td>intense yellow, buffy or grayish buff</td>
<td>intense yellow, buffy or grayish buff</td>
<td>intense yellow, buffy or grayish buff</td>
</tr>
<tr>
<td><strong>Tail fur</strong></td>
<td>moderately hirsute bicolored dorsoventrally</td>
<td>moderately hirsute bicolored</td>
<td>moderately hirsute bicolored</td>
<td>moderately hirsute bicolored</td>
<td>densely hirsute strongly bicolored</td>
<td>moderately hirsute bicolored dorsoventrally</td>
</tr>
<tr>
<td><strong>Tail color</strong></td>
<td>bicolored</td>
<td>slightly bicolored</td>
<td>slightly bicolored</td>
<td>slightly bicolored</td>
<td>uniformly bicolored</td>
<td>uniformly bicolored</td>
</tr>
<tr>
<td><strong>Spheno-palatine vacuities</strong></td>
<td>Short, narrow, restricted to presphenoid, exposing orbitosphenoid</td>
<td>very short, narrow, restricted to presphenoid or absent</td>
<td>very short, narrow, restricted to presphenoid or absent</td>
<td>long, wide, extending on pre- and basisphenoid, barely exposing orbitosphenoid</td>
<td>long, wide, extending on pre- and basisphenoid, exposing orbitosphenoid</td>
<td>Long, wide, extending on pre- and basisphenoid, exposing orbitosphenoid</td>
</tr>
<tr>
<td><strong>Alisphenoid strut</strong></td>
<td>absent</td>
<td>absent</td>
<td>absent</td>
<td>present</td>
<td>present</td>
<td>absent</td>
</tr>
<tr>
<td><strong>Mesoloph/id</strong></td>
<td>both developed</td>
<td>developed/rarely reduced</td>
<td>developed/reduced or absent</td>
<td>reduced/reduced or absent</td>
<td>both developed</td>
<td>both developed</td>
</tr>
<tr>
<td><strong>Palatal fossae</strong></td>
<td>very deep</td>
<td>shallow</td>
<td>shallow</td>
<td>deep</td>
<td>deep</td>
<td>deep</td>
</tr>
<tr>
<td><strong>Projection of eustachian tube</strong></td>
<td>present in most specimens</td>
<td>absent</td>
<td>absent</td>
<td>absent</td>
<td>present in most specimens</td>
<td>present in few specimens</td>
</tr>
</tbody>
</table>
### ETYMOLOGY
This species is named after Dr. Alfredo Ricardo Langguth Bonino for his long-term dedication and commitment to the development of Brazilian mammalogy.

### DIAGNOSIS
*Cerradomys langguthi* is characterized by small body size, short and dense dorsal pelage, dorsal body color orange grizzled with brown, head color grayish, ventral body color grayish or slightly yellowish, short and narrow sphenopalatine vacuities, restricted to presphenoid, exposing partially orbitosphenoid, alisphenoid strut absent, deep palatal fossae (complex posterolateral palatal pits), and a unique chromosomal formula (2n = 48–50, FN = 56).

### MORPHOLOGICAL DESCRIPTION
Head and body size small (table 2); tail length longer than head and body (102%–144% of head and body length); hindfeet moderately narrow and long (21%–24% of head and body length), with large and fleshy interdigital, thenar, and hypothenar pads; pinnae rounded and small (11.3%–17% of head and body length). Dorsal pelage short and dense (table 1), consisting of short, dense underfur (wool hairs; thin, wavy, short) and longer and lax overfur (cover and guard hairs; thick, long). Dorsal body color buffy orange densely grizzled with black; wool hairs (range: 5–8 mm) with basal part grayish and distal part (1/10 of total length) orange or brown; cover hairs long (range: 9–12 mm), with distal 1/4 dark brown with a subterminal orange band; guard hairs sparse and long (range: 13–16 mm), with distal half entirely black or dark brown. Anterior half of head (until eyes) covered with gray-based and white- or buffy-tipped hairs, clearly distinct from color of posterior half of head and dorsal body fur. Ventral pelage composed of wool underfur and cover and guard hairs, with individual hairs grayish-based and tipped with white, buffy or yellowish; general ventral color grayish, buffy, or yellowish, slightly grizzled, and distinctively lighter than dorsal pelage. Flanks bright orange; banded cover hairs and dark guard hairs rare. Mystacial vibrissae long, reaching but not surpassing pinnae when laid back. Tail slightly bicolored to bicolorod, covered with short, sparse brown hairs and scales on dorsal surface and unpigmented hairs and scales on ventral surface. Dorsal surface of hind foot white, covered with short, entirely white hairs (only young individuals present hairs with 3/4 distal portion white and

<table>
<thead>
<tr>
<th>Measure</th>
<th><em>C. subflavus</em></th>
<th><em>C. vivoi</em></th>
<th><em>C. langguthi</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>HBL</td>
<td>151.38 ± 11.46 (125–179) 52</td>
<td>143.31 ± 8.65 (127–170) 36</td>
<td>136.30 ± 7.76 (119–153) 73</td>
</tr>
<tr>
<td>LT</td>
<td>179.87 ± 13.44 (150–210) 47</td>
<td>177.36 ± 11.48 (153–200) 39</td>
<td>169.61 ± 10.93 (144–195) 69</td>
</tr>
<tr>
<td>HF</td>
<td>34.02 ± 2.07 (30–38) 51</td>
<td>33.12 ± 1.53 (30–36) 39</td>
<td>32.58 ± 1.35 (30–35) 77</td>
</tr>
<tr>
<td>Ear</td>
<td>21.35 ± 2.04 (17–25) 52</td>
<td>21.94 ± 1.70 (18–25) 40</td>
<td>20.54 ± 0.90 (19–22) 59</td>
</tr>
<tr>
<td>Wt</td>
<td>96.44 ± 16.97 (69–129) 27</td>
<td>77.56 ± 12.49 (54–110) 38</td>
<td>62.57 ± 10.86 (37.5–85) 70</td>
</tr>
<tr>
<td>CIL</td>
<td>34.36 ± 1.40 (30.67–36.52) 52</td>
<td>32.93 ± 1.13 (30.76–35.41) 43</td>
<td>31.51 ± 1.00 (29.23–33.45) 65</td>
</tr>
<tr>
<td>LD</td>
<td>10.32 ± 0.68 (8.79–11.77) 56</td>
<td>10.07 ± 0.58 (8.89–11.24) 42</td>
<td>9.40 ± 0.45 (8.32–10.45) 65</td>
</tr>
<tr>
<td>LM</td>
<td>5.17 ± 0.18 (4.77–5.52) 56</td>
<td>4.91 ± 0.17 (4.58–5.29) 40</td>
<td>4.90 ± 0.15 (4.52–5.19) 59</td>
</tr>
<tr>
<td>BMI</td>
<td>1.49 ± 0.06 (1.35–1.62) 60</td>
<td>1.41 ± 0.07 (1.26–1.58) 43</td>
<td>1.42 ± 0.06 (1.28–1.55) 77</td>
</tr>
<tr>
<td>LIF</td>
<td>7.69 ± 0.48 (6.53–8.63) 60</td>
<td>7.51 ± 0.37 (6.82–8.38) 43</td>
<td>6.97 ± 0.30 (6.28–7.5) 79</td>
</tr>
<tr>
<td>PB</td>
<td>6.13 ± 0.24 (5.56–6.77) 59</td>
<td>5.71 ± 0.26 (5.17–6.23) 43</td>
<td>5.71 ± 0.22 (5.18–6.14) 79</td>
</tr>
<tr>
<td>BR</td>
<td>9.65 ± 0.56 (8.51–10.84) 53</td>
<td>9.34 ± 0.59 (7.89–10.61) 41</td>
<td>9.66 ± 0.47 (8.57-10.78) 69</td>
</tr>
<tr>
<td>LN</td>
<td>14.17 ± 0.86 (11.98–16.00) 60</td>
<td>13.73 ± 0.66 (12.30–15.01) 41</td>
<td>13.54 ± 0.57 (12.22–14.97) 72</td>
</tr>
<tr>
<td>LPB</td>
<td>5.94 ± 0.22 (5.39–6.50) 57</td>
<td>5.89 ± 0.28 (5.37–6.60) 42</td>
<td>5.62 ± 0.31 (4.91–6.31) 79</td>
</tr>
<tr>
<td>HB</td>
<td>11.04 ± 0.37 (10.39–11.97) 54</td>
<td>10.73 ± 0.42 (9.78–11.69) 40</td>
<td>10.32 ± 0.37 (9.58–11.13) 62</td>
</tr>
<tr>
<td>LIB</td>
<td>6.33 ± 0.33 (5.73–7.06) 60</td>
<td>5.83 ± 0.26 (5.19–6.39) 41</td>
<td>5.67 ± 0.30 (5.14–6.4) 79</td>
</tr>
<tr>
<td>ZB</td>
<td>18.97 ± 0.72 (17.27–20.56) 47</td>
<td>17.89 ± 0.62 (16.63–19.09) 40</td>
<td>17.52 ± 0.66 (16.17–19.2) 66</td>
</tr>
<tr>
<td>CZL</td>
<td>26.39 ± 1.01 (24.04–28.31) 52</td>
<td>25.20 ± 0.82 (23.72–26.98) 43</td>
<td>24.15 ± 0.87 (21.92–26.5) 69</td>
</tr>
<tr>
<td>OFL</td>
<td>13.08 ± 0.56 (11.56–14.01) 60</td>
<td>12.38 ± 0.45 (11.43–13.46) 42</td>
<td>11.81 ± 0.41 (10.81–12.83) 78</td>
</tr>
<tr>
<td>BB</td>
<td>5.34 ± 0.15 (5.04–5.71) 54</td>
<td>5.03 ± 0.18 (4.64–5.39) 41</td>
<td>4.85 ± 0.22 (4.34–5.34) 68</td>
</tr>
</tbody>
</table>

### TABLE 2
Descriptive Statistics of External and Skull Measurements for the Three Species of *Cerradomys* from Eastern Brazil

Mean ± standard deviation, (minimum-maximum), N.

- **HBL**: Head and body length
- **Ear**: Ear length
- **Wt**: Body weight
- **CIL**: Ciliature length
- **LD**: Lateral dental series length
- **LM**: Maxillary length
- **BMI**: Maxillary breadth
- **LIF**: Lateral incisor length
- **PB**: Premaxillary breadth
- **BR**: Body ring length
- **LN**: Lower incisor length
- **LPB**: Lower premaxillary breadth
- **HB**: Head breadth
- **LIB**: Lower incisor breadth
- **ZB**: Zygomatic breadth
- **CZL**: Crowns and zygomatic breadth
- **OFL**: Occipital breadth
- **BB**: Braincase breadth

**C. subflavus**

**C. vivoi**

**C. langguthi**
basal 1/4 grayish or washed brown); ungual tufts sparse, shorter than claws especially on digit I; ventral surface naked, unpigmented, with four interdigital pads and two tarsal pads (thenar and hypothenar). Pinnae covered internally with short orange hairs and externally with orange, brown-tipped hairs.

Skull size small (tables 1, 2; figs. 3, 5, 8). Rostrum long and broad, tapering, with inflated capsular projection of nasolacrimal foramen, and flanked by deeply excavated zygomatic notches; interorbital region long and narrow (table 2), converging anteriorly, with dorsolateral margins with sharp and well-developed supraorbital crests; braincase oblong, with prominent temporal crests. Zygomatic plate (in lateral view) projected forward, with dorsal free margin rounded and anterior margin straight or slightly concave, and zygomatic spine absent. Incisive foramina long (averaging about 74.4% of length of diastema), with lateral margins concave and diverging posteriorly, and wider posteriorly; posterior margins extending or not between the alveolus of upper first molars. Palate long and wide (sensu Hershkovitz, 1962); postero-lateral palatal pits numerous and complex, recessed in very deep palatal fossae; palatal excrescences rarely present. Mesopterygoid fossa narrow, with anterior margin rounded or slightly acute, not reaching the alveolus of M3; bony roof of mesopterygoid fossa perforated by short, narrow to wide sphenopalatine vacuities, restricted to presphenoid, partially or totally exposing the orbitosphenoid. Alisphenoid strut absent (buccinator-masticatory foramen and ovale foramen confluent). Postglenoid foramen large, falciform or triangular, nearly equal to condyloid process; superior notch shallow; angular process short, not surpassing the condyloid process posteriorly; inferior notch shallow; capsular process of lower incisor well developed.

Incisors, upper and lower molars as for the genus (no mesoloph/mesolophid reduction was observed in C. langguthi). Mammary counts and soft anatomy (stomach and glans penis) as described for the genus.

**Karyology:** The *Cerradomys langguthi* holotype shows a karyotype with 2n = 50 and FN = 56 (table 7). The autosomal complement of specimens with 2n = 50 comprises four biarmed pairs (1 large, 3 medium to small pairs) and 20acrocentric pairs (3 large and 17 medium to small pairs). The X chromosome is a medium-sized acrocentric and the Y chromosome a small-sized acrocentric. Variation in diploid number is due to centric fusion affecting two acrocentric pairs.

Maia and Hulak (1981) reported a variation on the chromosomal diploid number in Pernambuco samples (Tupanatinga, Buíque, Bom Conselho, Capoeiras, Correntes, Panels, Caruaru, São Lourenço, and Exu), with 2n = 48, 49, and 50; all karyotypes present the same fundamental number FN = 56. The karyotype of the holotype is coincident with the 2n = 50, FN = 56 karyotype presented by Maia and Hulak (1981). Moreover, when examining specimens from Buíque, one of the localities sampled by Maia and Hulak (1981), we found them to be conspecific with *C. langguthi*.

The 2n = 48-49-50 polymorphism reported by Maia and Hulak (1981) results from multiple centric fusions. The karyotype observed in the Exu sample, 2n = 46, FN = 56, differs from the basic complement by the presence of two large submetacentric pairs formed by centric fusions. Bonvicino (2003) recognizes this karyotype as a distinct karyomorph, although we do not have compelling morphologic or molecular evidence to consider it a separate form.

**Natural History:** *Cerradomys langguthi* inhabits several vegetation types: the coastal lowland humid Atlantic Forest (locally called “Zona da Mata”); the open and relatively dry forests (locally called as “Agreste”) in the zone between the more humid and dense coastal forest and the more open and drier Caatinga;
the arbustive and arboreal Caatinga; and the forests restricted to humid slopes of mountain ranges in areas of Caatinga ("Brejo").

Paiva (1973) associated this species with sugar-cane plantations, near humid and mesic areas, in Ceará. This habitat is also pointed by Mares et al. (1981) and Streilein (1982) as typical for *C. langguthi* in Pernambuco. Karimi et al. (1976) also obtained this species in natural and cultivated fields, reporting nests in more humid grass patches. We trapped specimens of *C. langguthi* at secondary forests and shade coffee plantations on the slopes of Serra de Baturité (a typical "Brejo") in Ceará.

**Material Examined:**

*Cerradomys maracajuensis* (Langguth and Bonvicino, 2002)

*Cerradomys maracajuensis* Langguth and Bonvicino, 2002: 292; type locality: “Brazil, Mato Grosso do Sul: Municipality of Maracaju (approx. 21°38′S, 55°09′W) Fazenda da Mata”.

**Typical Habitat:** Weksler et al., 2006: 8.

**Type Locality:** “Brazil, Mato Grosso do Sul: Municipality of Maracaju (approx. 21°38′S, 55°09′W) Fazenda da Mata”.

**Geographic Distribution:** The known collection localities of *C. maracajuensis* (figs. 1, 2) are distributed across the Brazilian Cerrado on Minas Gerais, Mato Grosso do Sul, and Mato Grosso to Paraguayan open areas of Amambay, Caaguazu, Canendiyu, Paraguari, La Cordillera, San Pedro, and Concepción. To west, *C. maracajuensis* penetrates the open vegetation lowlands of Bolivia, at Beni, Santa Cruz, and La Paz, and Peru, at Puno. Most records are from lowlands although there are also records of specimens from highlands and dissected highlands of central South America; the known altitudinal range varies from 102 m (Tacuati, Paraguay; USNM 293154) to 1750 m (Pitiguaya, Bolivia; e.g., AMNH 72641).

**Diagnosis:** *C. maracajuensis* is characterized by large body and tail size (HBL range: 140–185 mm; TL range: 171–227 mm; see table 3), long and more robust feet (HF range: 34–43 mm), dorsal body color coarsely grizzled, buffy brown to orange brown, ventral body color grayish to buffy to yellow gray, skull (figs. 4, 6) with shallow rostral fossa, mesopterygoid fossa with small and narrow sphenopalatine vacuities or fully ossified, shallow palatal fossae (simple and large table 3), long and more robust feet (HF range: 34–43 mm), dorsal body color coarsely grizzled, buffy brown to orange brown, ventral body color grayish to buffy to yellow gray, skull (figs. 4, 6) with shallow rostral fossa, mesopterygoid fossa with small and narrow sphenopalatine vacuities or fully ossified, shallow palatal fossae (simple and large postero-lateral palatal pits), developed palatal excrescences, mesolophid developed (narrow and reduced in few individuals from Bolivia), central cartilaginous digit of distal baculum absent, and a unique chromosomal formula (2n = 56, FN = 58).

**Karyology:** Diploid number of 56 chromosomes and low fundamental number of 58 autosomes are diagnostic for this species when compared with other congeneric species (see Bonvicino et al., 1999; Langguth and Bonvicino, 2002: fig. 3).

**Natural History:** Available data on museum specimens relating to the natural history of *C. maracajuensis* report habitat preference: Near the type locality, Maracaju, specimens were collected in some distinct habitats like “woods”, “bush and grass”, “Brush pile in
grass at edge of rice field”, “Brush-Swampy”, “Forest”, and “Brush and grass” (R.M. Gilmore, in museum tags at AMNH and MN), but also in gallery forest (Langguth and Bonvicino, 2002). In Bolivia, specimens were collected in dense shrub-grass field (clear forest), dense forest adjacent (G. Schmitt, AMNH tags). Pregnant females were observed in Bolivia during June and September, with two and four embryos (Anderson, 1997).

**Specimens Examined: BOLIVIA: BENI:**

### TABLE 3
Descriptive Statistics of External and Skull Measurements for the Three Species of *Cerradomy*
Mean ± standard deviation, (minimum-maximum), N.

<table>
<thead>
<tr>
<th></th>
<th><em>C. maracajuensis</em></th>
<th><em>C. marinhus</em></th>
<th><em>C. scotti</em></th>
</tr>
</thead>
<tbody>
<tr>
<td>HBL</td>
<td>151.48 ± 13.17 (123–185)</td>
<td>168.00 ± 11.07 (153–179)</td>
<td>148.23 ± 13.02 (125–181)</td>
</tr>
<tr>
<td>HF</td>
<td>35.16 ± 2.30 (30–40)</td>
<td>40.33 ± 1.75 (38–43)</td>
<td>31.65 ± 2.11 (27–37)</td>
</tr>
<tr>
<td>Ear</td>
<td>20.49 ± 0.97 (19–22)</td>
<td>22.58 ± 1.07 (21.50–24)</td>
<td>22.18 ± 1.92 (18–26)</td>
</tr>
<tr>
<td>Wt</td>
<td>91.10 ± 16.78 (54–125)</td>
<td>130 ± 18.03 (105–150)</td>
<td>91.95 ± 18.10 (55–133)</td>
</tr>
<tr>
<td>CIL</td>
<td>32.60 ± 1.14 (29.68–35.39)</td>
<td>36.05 ± 1.38 (34.32–38.25)</td>
<td>33.08 ± 1.41 (30.27–36.07)</td>
</tr>
<tr>
<td>LD</td>
<td>9.50 ± 0.49 (8.50–10.82)</td>
<td>10.38 ± 0.20 (10.18–10.63)</td>
<td>9.66 ± 0.60 (8.59–10.95)</td>
</tr>
<tr>
<td>LM</td>
<td>5.26 ± 0.19 (4.79–5.67)</td>
<td>5.74 ± 0.11 (5.58–5.86)</td>
<td>5.05 ± 0.16 (4.71–5.39)</td>
</tr>
<tr>
<td>BM1</td>
<td>1.54 ± 0.08 (1.37–1.74)</td>
<td>1.72 ± 0.04 (1.66–1.77)</td>
<td>1.52 ± 0.04 (1.44–1.60)</td>
</tr>
<tr>
<td>LIF</td>
<td>7.07 ± 0.40 (6.10–7.96)</td>
<td>7.47 ± 0.40 (7.09–8.15)</td>
<td>6.83 ± 0.32 (6.14–7.55)</td>
</tr>
<tr>
<td>PB</td>
<td>6.22 ± 0.26 (5.60–6.80)</td>
<td>6.41 ± 0.12 (6.25–6.57)</td>
<td>6.22 ± 0.31 (5.55–6.92)</td>
</tr>
<tr>
<td>BR</td>
<td>9.80 ± 0.75 (7.65–11.48)</td>
<td>9.92 ± 0.50 (9.62–10.67)</td>
<td>9.89 ± 0.66 (8.55–11.49)</td>
</tr>
<tr>
<td>LN</td>
<td>14.02 ± 0.74 (12.3015.88)</td>
<td>16.16 ± 0.59 (15.14–16.70)</td>
<td>14.54 ± 0.99 (12.08–16.93)</td>
</tr>
<tr>
<td>LPB</td>
<td>6.37 ± 0.39 (5.62–7.32)</td>
<td>7.53 ± 0.59 (6.84–8.46)</td>
<td>6.45 ± 0.42 (5.66–7.39)</td>
</tr>
<tr>
<td>HB</td>
<td>11.06 ± 0.34 (10.26–11.82)</td>
<td>11.74 ± 0.33 (11.22–11.99)</td>
<td>11.24 ± 0.46 (10.34–12.26)</td>
</tr>
<tr>
<td>LIB</td>
<td>5.92 ± 0.42 (5.16–6.84)</td>
<td>7.08 ± 0.34 (6.73–7.56)</td>
<td>5.83 ± 0.40 (4.89–6.72)</td>
</tr>
<tr>
<td>ZB</td>
<td>18.88 ± 0.74 (17.12–20.69)</td>
<td>19.85 ± 0.77 (18.79–20.66)</td>
<td>18.82 ± 0.90 (17.12–20.89)</td>
</tr>
<tr>
<td>CZL</td>
<td>25.63 ± 0.99 (23.29–27.72)</td>
<td>27.47 ± 0.52 (26.71–28.01)</td>
<td>25.76 ± 1.11 (23.46–27.91)</td>
</tr>
<tr>
<td>OFL</td>
<td>12.80 ± 0.40 (11.93–13.64)</td>
<td>13.58 ± 0.45 (13.09–14.20)</td>
<td>13.32 ± 0.53 (12.32–14.44)</td>
</tr>
<tr>
<td>BB</td>
<td>5.26 ± 0.19 (4.85–5.73)</td>
<td>5.42 ± 0.08 (5.34–5.55)</td>
<td>5.35 ± 0.21 (4.79–5.70)</td>
</tr>
</tbody>
</table>


_Cerradomys marinus_ (Bonvicino, 2003)

_Oryzomys marinus_ Bonvicino, 2003: 84; type locality: “Fazenda Sertão do Formoso (known before as Fazenda Jucurutu, 14°40′20″S 45°49′71″S [sic]W, altitude around 775 m), Jaborandi municipality, state of Goiás, Brazil.”

[ _Cerradomys_ ] _marinus_: Weksler et al., 2006: 8.

TYPE LOCALITY: “Fazenda Sertão do Formoso (known before as Fazenda Jucurutu, 14°40′20″S 45°49′71″S [sic]W, altitude around 775 m), Jaborandi municipality, state of Goiás, Brazil.” However, Bonvicino (2003: 79) previously reported that Fazenda Sertão do Formoso was “located in Jaborandi and Cocos municipalities, Bahia state” (see also IBGE, 1972, for the location of these municipalities). Moreover, the geographical coordinates originally presented by Bonvicino did not refer to the collecting locality of the type series. More precisely, all these specimens were captured in one specific habitat type, “veredas”, whose geographical coordinates were also furnished by the author (Bonvicino, 2003: 79). Thus, the correct type locality for this species is herein corrected and restricted to: Fazenda Sertão do Formoso, (formerly known as Fazenda Jucurutu, 14°48′8″S, 45°57′W, altitude around 775 m), Jaborandi municipality, state of Bahia, Brazil.

**Geographic Distribution:** Besides the type locality, _C. marinus_ is known from one collection locality in northwestern Minas Gerais state (figs. 1, 2).

**Diagnosis:** _C. marinus_ is characterized by large body and tail size (HBL range, 153–179 mm; TL range, 198–212 mm; table 3), and robust feet (HF range, 38–43 mm), dorsal body color coarsely grizzled, buffy brown to orange brown, ventral body color grayish to buffy to yellowish gray, skull with shallow rostral fossa (figs. 4, 6), mesopterygoid fossa with small and narrow sphenopalatine vacuities or fully ossified, shallow palatal fossae (simple and large posterolateral palatal pits), m3 with reduced or absent mesolophid, and a unique chromosomal formula (2n = 56, FN = 54).

**Karyology:** The karyotype of _C. marinus_ presents 2n = 56 and FN = 54; the autosomal complement comprises 27 acrocentric pairs, from large to small. Sexual chromosomes differ in size, with the X chromosome being a large acrocentric and the Y a small-size acrocentric (Bonvicino et al., 1999; Bonvicino, 2003).

**Natural History:** Specimens of _O. marinus_ from Fazenda Sertão do Formoso were captured in a particular Cerrado habitat, called “vereda”. “Vereda” is a periodically flooded grassland habitat with scattered palm species of genera _Mauritia_ and _Mauritiella_, generally in Cerrado stream headwaters (see detailed description in Bonvicino, 2003: 79, 87). Specimens from Parque Nacional Grande Sertão Veredas were captured at seasonally flooded semideciduous forests (A.P. Carmignotto field notes).

Reproductive data suggested that females breed throughout the year because pregnant females were captured both in dry and rainy seasons, with embryo numbers ranging from 2–4, and with a modal number of 4 embryos

2These specimens were employed by D’Elia et al. (submitted) as vouchers for the first record of _C. maracajaensis_ for Paraguay.
Cerradomys marinhus is infested by ectoparasites like mites and ticks of the order Mesostigmata (family Ixodidae); fleas Polygenis [Polygenis] tripus; and flies of the family Hippoboscidae (Bonvicino, 2003: 88).


**Cerradomys scotti**

*(Langguth and Bonvicino, 2003)*

*Oryzomys scotti* Langguth and Bonvicino, 2003: 290; type locality: “Brazil, Goiás: municipality of Corumbá de Goiás (approx. 15° 54’S, 48° 48’W), Morro dos Cabeludos”.

*Oryzomys andersoni* Brooks, Baker, Vargas, Tarifa, Aranibar and Rojas, 2004: 3; type locality: “Pozo Mario, Estancia Las Conchas, Santa Cruz, Bolivia; 1735–46.9’S, 5930–20.5°W”.

**Type Locality:** “Brazil, Goiás: municipality of Corumbá de Goiás (approx. 15°54’S, 48°48’W), Morro dos Cabeludos”.

**Geographic Distribution:** *Cerradomys scotti* presents a large distribution area in central South America (figs. 1, 2), with limits similar to those of the biome Cerrado. Collecting records were found along a north-south transect, from the Brazilian states of Piauí, Maranhão, and Tocantins to the south-central state of Paraguari in Paraguay. Similarly, collecting records of *C. scotti* were found across an east-west transect, from the eastern Brazilian state of Minas Gerais to the eastern Departamento de Santa Cruz in Bolivia.

This species is geographically restricted to the central Brazilian highlands and several adjacent areas. Altitudinally, *C. scotti* is distributed from 250 m in Santa Rosa de la Roca, Santa Cruz, Bolivia, to 1180 m in Alto Paraiso, Goiás, Brazil.

**Taxonomic Comments:** Based on molecular evidence, Emmons et al. (2006) suggest that *Cerradomys andersoni* is conspecific with, and consequently a junior synonym of *C. scotti*. Brooks et al. (2004) also highlighted the close relationship of cytochrome *b* haplotype of *C. andersoni* holotype (CBF 6151) and the specimen deposited in GenBank under the accession number AF181277 (voucher specimens: MN50379, MN61677), herein identified as *C. scotti*.

The only known specimen of *C. andersoni* shared similarities with specimens of *C. scotti* (based on photographs of the holotype provided by Dr. Julieta Vargas): bicolored tail, presence of alisphenoid strut, and reduction of mesoloph on M2. Additionally, it was nearly identical with the *C. scotti* holotype, with some other Bolivian specimens from Santa Cruz (AMNH 263872), and with some Paraguayan specimens from Sapucay (USNM 121389–91, 121393–5), Tobati (UMMZ 126005, MVZ 145148), Altos (MVZ 145147), and Concepcion (MVZ 141890).

The karyotype of *C. andersoni* is unknown, precluding comparisons with our samples of *C. scotti*. However, our data support the taxonomic decision held by Emmons et al. (2006), and we also consider *C. andersoni* a junior synonym to *C. scotti*.

**Diagnosis:** *C. scotti* is characterized by medium body and tail size (HBL range, 125–181 mm; TL range, 141–198 mm; table 3), small feet (HF range, 24–37 mm), dorsal body color coarsely grizzled, buffy brown, ventral body color grayish, tail bicolored and hirsute, skull (fig. 4, 6) with deep rostral fossa, mesopygoid fossa with large and wide sphenopalatine vacuities (exposing orbitosphenoid), alisphenoid strut present, deep palatal fossae (complex posterolateral palatal pits), M2 with reduced mesoloph, m1 and m2 with reduced or absent mesolophid, central cartilaginous digit of distal baculum absent, and a unique chromosomal formula (2n = 58, FN = 70–72).

**Karyology:** The *Cerradomys scotti* karyotype exhibits a 2n = 58 and FN = 70–72 (table 7). The autosomal complement comprises seven biarmed pairs and 20 acrocentric pairs from large to small. The X chromosome is a large submetacentric and the Y a medium submetacentric (Bonvicino et al., 1999; Langguth and Bonvicino, 2002).

**Natural History:** Distinctly from other congenic species (more associated with forest habitats, as gallery forest and “cerrado”), *Cerradomys scotti* is most frequently captured on the open habitats of Cerrado and on the ecotone between forested and open...
areas. Thus, based on tags, journal notes, and collector information this species is frequently observed on “campo sujo”, “campo cerrado”, and “cerrado sensu stricto”. Langguth and Bonvicino (2002) stated that this species is also found on “veredas” and gallery forests, but less frequently than in more open vegetational habitats of the Cerrado. *Cerradomys scotti* is known to present scansorial ability (Alho and Villela, 1984).

**Specimens Examined:** **BOLÍVIA:** SANTA CRUZ: El Refugio Pampa, NE from camp: F: USNM 584583. Pozo Mario, Estancia Las Conchas: M: CBF 6151 (photography). Santa Rosa de La Roca: M: UFMG 263872.


**Cerradomys subflavus** (Wagner, 1842)

*Mus vulpinus* Lund, 1840: 279; type locality: “Lagoa Santa, Minas Gerais, Brazil”. Pre-occupied by *Mus vulpinus* Brants 1827, a name associated with genus *Holochilus*.


*Mus vulpinoides* Schinz, 1845: 193; renaming of *Mus vulpinus* Lund, 1840.

*Calomys laticeps*: Winge, 1888: 51 (erroneous name combination; see Musser et al., 1998: 263, 298).

*Oryzomys subflavus* Thomas, 1901: 528.

*Cerradomys* subflavus*:* Weksler et al., 2006: 8.

**Type Locality:** Lagoa Santa, Minas Gerais, Brazil; 19°37’S, 43°53’W; 760 m.

**Geographic Distribution:** Populations currently assigned to *Cerradomys subflavus* (figs. 1, 2) are distributed throughout the Brazilian states of Goiás, Minas Gerais, and São Paulo, usually associated with the interior highlands in Minas Gerais and São Paulo, and with the Central Brazilian Highland in Goiás.

**Diagnosis:** *C. subflavus* is characterized by medium to large body and tail size (HBL range, 120–179 mm; TL range, 150–210 mm; table 3), small feet (HF range, 31–34 mm),
dorsal body color coarsely grizzled, buffy to orange brown, head color grayish, ventral body color grayish, skull (figs. 3, 5) with deep rostral fossa, mesopterygoid fossa with large and wide sphenopalatine vacuities (exposing orbitosphenoid), deep, narrow, and long palatal fossae (complex posterolateral palatal pits), eustachian tube short with a distinct medial bony lamina dorsal to carotid canal, central cartilaginous digit of distal baculum reduced, and a unique chromosomal formula (2n = 54, FN = 62).

**Karyotype:** The Cerradomys subflavus karyotype shows 2n = 54 and FN = 62 (table 7, fig. 10). The autosome complement comprises five biarmed pairs (three large and two small) and 21 small acrocentric pairs. The X chromosome is a medium acrocentric and the Y, a small acrocentric (Bonvicino et al., 1999; Langguth and Bonvicino, 2002).

**Natural History:** The Brazilian populations of Cerradomys subflavus are known to occur at the mesic habitats of Cerrado, in gallery forest and “cerradão”, as well as the patches of semideciduous forest of Brazilian highlands, and the coastal Atlantic Forest. On the other hand, museum tags of specimens from the Brazilian locality of Anápolis, Goiás, describe C. subflavus habitats as: “high dry grass, edge woods and farm”; “Brush and high grass-dry-hill-edge of woods-farm”; “edge of cane fields and woods-dry”; “edge of corn field and woods”; “heavy grass, open woods, small stream”; “Edge of high grass, woods, and farms”; “Dry, high grass on hill-50 y[ar]ds. from woods”; “Capim seco [= Dry grass], near woods + farm, farm” (R.M. Gilmore, in museum tags at AMNH and MN).

Cerradomys subflavus is predominantly a terrestrial species (Stallings, 1989), although it might eventually be captured in trees (Fonseca and Kierulff, 1989).

Cerradomys vivoi, new species

Figures 1–3, 5, 7, 9–11; tables 1 and 2

Holotype: MN 35898, an adult male collected by E. Hingst-Zaher and M. Lara (original field number EDH 60), on 29 December, 1992. The holotype consists of an undamaged skin, skull with fractured left zygomatic arch and right auditory bulla missing, and postcranial skeleton. The cell suspension of bone marrow in Carnoy’s fixative (methanol:acetic acid) and the liver-tissue aliquot preserved in ethanol are housed at Laboratório de Biologia e Parasitologia de Mamíferos Reservatórios Silvestres, Instituto Oswaldo Cruz–FIOCRUZ, under original field number EDH 60. Cytochrome b DNA data are available in GenBank under the accession number AF181275.

External and selected skull dimensions of holotype are: HBL, 152; LT, 180; HF, 34 (including claw); Ear, 21; Wt, 82.15; CIL, 32.23; LD, 9.62; LM, 4.88; BM1, 1.40; LIF, 6.82; PB, 5.45; BR, 6.56; LN, 13.79; LPB, 5.89; HB, 10.36; LIB, 5.19; CZL, 25.10; OL, 12.62.

Paratypes: We assign all specimens herein examined (see below) as paratypes of C. vivoi.

Type Locality: The Cerradomys vivoi holotype was captured in coastal Atlantic Forest, near Itabuna in the Brazilian state of Bahia. The type locality is situated at Parque Zoobotânico da Comissão Executiva do Plano da Lavoura Cacaueira (CEPLAC), situated 6 km E of Itabuna by road, state of Bahia, Brazil, at 14°48' S, 39°16' W, on sea level (figs. 1, 2).

Distribution: Known collection localities of C. vivoi are distributed in the Brazilian states of Minas Gerais, Bahia, and Sergipe. In the state of Minas Gerais, distribution records are restricted to the northern portion of this state, around Rio Jequitinhonha, and Rio São Francisco and its right bank tributaries. In Bahia, sampling localities of C. vivoi extend from the coastal region to the eastern bank of Rio São Francisco. In Sergipe, the only recorded site of collection is in the coastal region, near the mouth of Rio São Francisco (figs. 1, 2).

Etymology: This species is named after Dr. Mario de Vivo, mammal curator of Museu de Zoologia da Universidade de São Paulo, for his outstanding contribution to the development of mammalogy in Brazil.

Diagnosis: Cerradomys vivoi is characterized by intermediate body size, short and dense dorsal pelage, dorsal body color orange grizzled with brown, head color grayish, ventral body color grayish or slightly yellowish, skull with long and wide sphenopalatine vacuities, alisphenoid strut absent, deep palatal fossae (complex posterolateral palatal pits), and a unique chromosomal formula (2n = 50, FN = 62–64).

Morphological Description: Head and body medium sized (table 2); tail length longer than head and body (127%–150% of head and body length); hind feet moderately narrow and long (21%–24% of head and body length), with large and fleshy interdigital, thenar, and hypothenar pads; pinnae rounded and small (12.5%–16% of head and body length). Dorsal pelage short and dense (table 1), consisting of short and dense underfur (wool hairs; thin, wavy, short) and longer and lax overfur (guard and cover hairs; thick, long). Dorsal body color buffy orange densely grizzled with black; wool hairs (range: 6–11 mm) with basal part grayish and distal part (1/10 of total length) orange or brown; cover hairs long (range: 9–15 mm), with distal 1/4 dark brown with a subterminal orange band; guard hairs sparse and long (range: 14–20 mm), with distal half entirely black or dark brown. Anterior half of head (until eyes) covered with gray-based and white- or buffy-tipped hairs, clearly distinct from color of posterior half of head and dorsal body fur. Ventral pelage composed of wool, cover and guard hairs, with individual hairs gray at the base and tipped with white, buff, or yellow; general ventral color grayish, buffy, or yellowish, slightly grizzled, and distinctively lighter than dorsal pelage. Flanks bright orange; banded cover hairs and dark guard hairs rare. Mystacial vibrissae long, reaching but not surpassing pinnae when laid back. Tail slightly bicolored, covered with short, sparse brown hairs and scales on dorsal surface and

The specimen MN 35898 was selected as holotype, because it is the only available specimen with karyotype and tissue sample.
Fig. 9. Dorsal, ventral, and lateral cranial views of the holotype of *Cerradomys vivoi* (MN 35898; CIL, 32.2 mm).
unpigmented hairs and scales on ventral surface. Dorsal surface of hind foot predominantly white, covered with short hairs, with 3/4 distal portion white and basal 1/4 grayish or washed brown; ungual tufts sparse, shorter than claws especially on digit I; ventral surface naked, unpigmented, with four interdigital pads and two tarsal pads (thenar and hypothenar). Pinnae covered internally with short orange hairs and externally with orange, brown-tipped hairs.

Skull size intermediate (tables 1, 2; figs. 3, 5). Rostrum long, broad, tapering, with inflated capsular projection of nasolacrimal foramen, and flanked by deeply excavated zygomatic notches; interorbital region long and narrow (table 2), converging anteriorly, with dorsolateral margins with sharp and well-developed supraorbital crests; braincase oblong, with prominent temporal crests. Zygomatic plate (in lateral view) projected forward, with dorsal free margin rounded and anterior margin straight or slightly concave, and zygomatic spine absent. Incisive foramina long (averaging about 74% of length of diastema), with lateral margins concave and
diverging posteriorly, wider posteriorly; posterior margins extending or not between the alveolus of upper first molars. Palate long and wide (sensu Hershkovitz, 1962); posterolateral palatal pits numerous and complex, recessed in moderately deep palatal fossae; palatal excrescences rarely present. Mesopterygoid fossa narrow, with anterior margin rounded or slightly acute, not reaching the alveolus of M3; bony roof of mesopterygoid fossa perforated by long and wide sphenopalatine vacuities, exposing the orbitosphenoid. Alisphenoid strut absent (buccinator-masticatory foramen and ovale foramen confluent). Postglenoid foramen large and nearly semicircular in shape separated from small subsquamosal fenestra (absent in specimens with moderate to heavy tooth wear), by a wide hamular process of squamosal. Tegmen tympani weakly overlapping squamosal; posterior suspensory process of squamosal absent. Ectotympanic bullae globose; eustachian tube short, with distinct medial laminae in a few specimens; stapedial process short and wide, overlapping squamosal; bony process dorsal to stapedial process present, overlapping squamosal.

Mandible long and deep (fig. 9); coronoid process large, falciform or triangular, nearly equal to condyloid process; superior notch shallow; angular process short, not surpassing the condyloid process posteriorly; inferior notch shallow; capsular process of lower incisor well developed.

Incisors, upper and lower molars as for the genus (no mesoloph/mesolophid reduction was observed in *C. vivoi*). Mammary counts and soft anatomy (stomach and glans penis) as described for the genus.

**Karyology:** The holotype of *Cerradomys vivoi* (MN 35898), as well as seven specimens collected in Caetité, Bahia, and Juramento, Minas Gerais, showed a karyotype with 2n = 50 and FN = 62–63 (table 7, figs. 10, 11). The autosomal complement comprises seven biarmed pairs (1 large, 4 medium, and 2 small pairs) and 17 acrocentric pairs (1 large and 16 small pairs). The X chromosome is a medium-sized acrocentric and the Y, small-sized. Variation in autosome fundamental number is due to pericentric inversion affecting a medium acrocentric. The G-banding pattern of *C. vivoi* allowed the unequivocal identification of homologues in autosome pairs and sex chromosomes (fig. 11). One male specimen (LV-FC22) and one female specimen (MN 61661) showed 2n = 50, FN = 63, due to a pericentric inversion involving one chromosome of a medium autosomal pair.

**Natural History:** In the Cerrado-Caatinga transitional areas, *Cerradomys vivoi* inhabits secondary semideciduous and gallery forests, as well as arboreal Caatinga (all information on the natural history of this species is based on Hingst et al., 1997, except where noted). In the same region this species was trapped along with *Gracilarmania agilis*, *Marmosops incanus*, *Monodelphis domestica*, *Calomys expulsus*, *Oligoryzomys fornesi*, *Oligoryzomys nigripes*, *Oligoryzomys stramineus*, *Nectomys ratus*, *Rhipidomys sp.*, *Wiedomys pyrrhorhinos*, and *Thrichomys aperiodes*. The diet of *C. vivoi* consists mainly of vegetal material and arthropods. The only ectoparasite species found in *C. vivoi* is the mite *Gigantolaelaps vitzhumi* (Acari: Laelapidae).

In the Atlantic Forest, at Una Biological Reserve located in southern Bahia (Pardini, 2004), *Cerradomys vivoi* was captured by R. Pardini (unpubl.; in litt., A.R. Percequillo archives), who provided the following account on this species abundance and habitat preference:

I had performed 36,288 trap nights employing Sherman traps and 10,368 pitfall nights using 35 liters buckets, an effort equally divided through six habitats (“cabruca” [disturbed canopy forests, mixed with cacao plantations: the understorey is removed and replaced by cacao plants], “capoeiras em estadio inicial a medio-inicial de regeneracao” [disturbed forests in initial and inicial/advanced stage of regeneration], and interior and edge of mature forest with more than 1,000 ha and interior and edge of mature forest less than 100 ha). I captured 9 *Oryzomys subflavus* [= *Cerradomys vivoi*], 1 in “cabruca” and 8 in “capoeiras”. Moreover, in preliminary un-standardized sampling, . . . I captured few individuals. . . of *subflavus* [= *C. vivoi*] in open abandoned pasture areas.... Considering the smaller effort of this sampling, the number of
individuals captured (2 or 3) was comparatively larger. Considering these numbers, I believe that \textit{subflavus} (= \textit{C. vivoi}) is absent or very rare in mature forests, including the edges, occurs in disturbed forests in initial stages of regeneration or other disturbed forests (as cabrucas) and that is probably more common in open areas. It is noteworthy that even in “capoeiras” or “cabrucas” they are relatively rare, since only 8 individuals were captured in 350 captures in “capoeiras” and only 1 specimen was captured in 299 captures in “cabrucas”.


**Species Limits and Comparisons**

**Morphologic Variation:** \textit{Cerradomys vivoi} differs from \textit{C. maracajuensis} and \textit{C. marinhus} by its shorter, rich orange-brown dorsal pelage, when compared to the longer and denser yellow-brown dorsal pelage of the latter two taxa (table 1). Additionally, the anterior half of dorsal head pelage is distinctively grayish to yellow gray in \textit{C. vivoi}, whereas in \textit{C. maracajuensis} and \textit{C. marinhus} its color is the same as the dorsal body pelage. The ventral pelage in \textit{C. vivoi} varies from grayish white to grayish yellow; conversely, in both \textit{C. maracajuensis} and \textit{C. marinhus} this region exhibits an intense yellow or buffy color. \textit{Cerradomys vivoi} also differs from \textit{C. maracajuensis} and \textit{C. marinhus} by the presence of posterolateral palatal pits recessed in deeply excavated fossae, whereas in the latter species pits are recessed in shallow palatal fossae. Another cranial feature that discriminates \textit{C. vivoi} from \textit{C. marinhus} and \textit{C. maracajuensis} is the roof of mesopterygoid fossa, which is perforated by long and wide sphenopalatine vacuities, largely exposing the orbitosphenoid region in \textit{C. vivoi}. On the other hand, both \textit{C. maracajuensis} and \textit{C. marinhus} are characterized by shorter vacuities restricted to plesphenoid (in all 92 Bolivian specimens of \textit{C. maracajuensis} surveyed for this trait) or by a completely ossified roof of mesopterygoid fossa (in 77% of 31 Paraguayan and Brazilian specimens of \textit{C. maracajuensis}).

\textit{Cerradomys vivoi} can be easily distinguished from \textit{C. scotti} and from the holotype of \textit{C. andersoni} (CBF 6151) by dorsal body color, which is buffy yellow with brown in the latter two species. \textit{Cerradomys vivoi} can also be recognized from \textit{C. scotti} and \textit{C. andersoni} in some tail traits (table 1): \textit{C. vivoi} exhibits a moderately hirsute tail, ranging from uniform to bicolored dorsoventrally, whereas both \textit{C. andersoni} and \textit{C. scotti} present strongly hirsute and sharply bicolored tails. Cranial differences are also conspicuous: \textit{C. scotti}, as well the holotype of \textit{C. andersoni}, are characterized by the presence of alisphenoid strut in 84% of specimens (n = 92), while this trait is
consistently absent in *C. vivoi*. Palatal excrescences are also present in *C. scotti* and absent in *C. vivoi* (small in the young specimen of *C. andersonii*). Dental morphological traits also are distinctive regarding *C. vivoi* and *C. scotti*: the mesolophid is reduced in M2 (22% of 78 examined specimens) and the mesolophid is consistently absent from m1 and m2 (in all surveyed specimens for this trait) in *C. scotti* (also in *C. andersonii*); conversely, both mesoloph and mesolophid are present in all specimens of *C. vivoi*.

The cranium of *C. vivoi* is readily distinguished from *C. subflavus* and *C. langguthi* by its long and wide sphenopalatine vacuities, largely exposing the orbitosphenoid, while *C. subflavus* shows long and narrow vacuities, barely exposing the orbitosphenoid and *C. langguthi* exhibits short and narrow sphenopalatine vacuities.

**Morphometric Variation: Cerradomys scotti, C. marinus, and C. maracajuensis** can be readily and unambiguously distinguished from *C. vivoi* by discrete qualitative traits (see also table 3 for external and cranial measurements). In contrast, *C. vivoi* is most similar to *C. subflavus* and *C. langguthi*, with more subtle cranial differences. Therefore, all our subsequent analyses (morphometric, karyologic, and molecular) were restricted to these three forms in order to emphasize the differences among them.

Sexual dimorphism in species of genus *Cerradomys* is not an important component of variation in cranial morphometrics (Brandt and Pêso, 1994; Percequillo, 1998), a pattern recurrent in oryzomyine (Goldman, 1918; Musser and Williams, 1985) and other sigmodontine rodents (Voss, 1988: 362; Carleton and Musser, 1989; Voss and Marcus, 1992). Consequently, we combined males and females for all subsequent univariate and multivariate statistical analyses herein performed.

Among external measurements, *C. vivoi* and *C. langguthi* differ significantly from *C. subflavus* in body length (one-way ANOVA, df = 160, F = 40.05, p <0.001) and weight (df = 134, F = 71.16, p <0.001). With respect to cranial measurements, 7 of 15 variables significantly discriminated the three species: CIL (one-way ANOVA, df = 159, F = 84.73, p <0.001), HB (df = 155, F = 51.03, p <0.001), LIB (df = 179, F = 84.08, p <0.001), ZB (df = 152, F = 65.87, p <0.001), CZL (df = 163, F = 91.87, p <0.001), OFL (df = 179, F = 123.31, p <0.001), and BB (df = 162, F = 104.18, p <0.001). All cranial measurements exhibited differences among these three species, with *C. subflavus* significantly larger, and the two other species of similar size (table 2). For those variables significantly different among the three species, *C. vivoi* is of intermediate size, while *C. subflavus* is consistently the most robust, and *C. langguthi* is always the smallest (table 2).

Principal component analysis confirmed that size was an important characteristic separating the three species. The first principal component is responsible for more than 50% of the observed variance (table 4). Moreover, all eigenvectors were positively related to the greatest variation axis. Some overlapping can be observed along the first axis (fig. 12), especially between the two smaller species, *C. vivoi* and *C. langguthi*. Discriminant analysis using log-transformed cranial measurements (fig. 13) separated the three species along the first and second canonical axes. OL, BB, and CIL were

![Table 4](https://example.com/table4.png)
the variables that mostly contributed to separation on first and second axis, respectively (table 5). Mahalanobis $D^2$ distances between species were all significant (table 6).

**Karyologic Variation:** Karyologic data reinforced the uniqueness of *Cerradomys vivoi* with respect to other species of the genus (*table 7*). When compared to *C. subflavus*, karyotypes were similar, since both species shared the same fundamental number. However, this resemblance is only apparent because they differed in diploid number and chromosome morphology. Complex rearrangements were responsible for these differences (fig. 10): *C. subflavus* has three large and two small biarmed chromosome pairs, while *C. vivoi* has two extra medium-sized biarmed chromosome pairs. Furthermore, *C. vivoi* exhibits a large acrocentric pair, the largest of its autosomal complement, without a recognizable counterpart in the *C. subflavus* complement, composed only of small-sized acrocentric pairs.

A similar $2n = 50$ and FN = 64 karyotype was previously reported for specimens collected within the distribution range of *Cerradomys vivoi*: specimens from localities from central Bahia, namely Jacobina, Andaraí, Morro do Chapéu, and Mucugê (Percequillo, 1998) and from Sergipe (Andrades-Miranda et al., 2002). Moreover, a $2n = 50$ and FN = 62 karyotype was found in specimens from northern Minas Gerais, Mocambinho (J. A. Oliveira, personal commun.). Quantitative and qualitative anal-

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**Fig. 12.** Individual specimen scores based on log-transformed values of 15 cranial measurements, projected onto the first and second principal components extracted from analysis of adult specimens of three species of genus *Cerradomys*: *C. subflavus*, *C. vivoi*, and *C. langguthi*. Results of principal components analysis are in table 4.
yses of voucher specimens from Mucugê (MZUSP 27403) and Mocambinho (MN 34432–4, 34436) and non-karyotyped specimens from Andaraí (UFPB 2376) allowed us to identify them as *C. vivoi*.

Some karyologic similarities were observed between *C. vivoi* and *C. langguthi*, which are characterized by a $2n = 48–50$ and FN = 56. This intrapopulational polymorphism overlaps with the known diploid number of *C. vivoi* (2n = 50), but not with its fundamental autosome number (FN = 64). However, *C. vivoi* was not variable in diploid number in samples from Bahia and Minas Gerais, south of Rio São Francisco (there is no available information of samples from Sergipe). Moreover, the morphologic distinctiveness of the autosomal complement between the samples from north of Rio São Francisco (herein *C. langguthi*) and *C. vivoi* allows for the recognition of both forms as separate species.

**Molecular Variation:** Genetic distance estimates between *C. vivoi* and *C. subflavus* were smaller (1.7%–2.0%) than between other species pairs of *Cerradomys* (table 8). Maximum-parsimony analyses placed Scotinomys and Neotoma apart from the oryzomyine species (fig. 14). The oryzomyines formed a trichotomy consisting of Nectomys genus, Sooretamys angouya, and a well-supported monophyletic group corresponding to *Cerradomys* species.

Within *Cerradomys*, the clade formed by *C. maracajuensis* and *C. marinhus* is the sister group to the remaining species of the genus: a
clade joining C. scotti and a clade grouping Cerradomys langguthi, Cerradomys vivoi, and C. subflavus. The topology of this latter clade is well resolved, with Cerradomys langguthi emerging as a sister taxon to a monophyletic group formed by C. subflavus and Cerradomys vivoi.

**SUMMARY, COMPARISONS, AND DISCUSSION**

Unlike other Brazilian species of Cerradomys (C. scotti, C. maracajuensis, and C. marinhus), C. vivoi, C. subflavus, and C. langguthi do not exhibit discrete autapomorphic diagnostic traits. On the contrary, as several sigmodontine rodents (see Patton et al., 2000; Bonvicino, 2003; D’Elia and Pardin, 2004; Gonçalves et al., 2005; Emmons and Patton, 2005; Percequillo et al., 2005) these species are diagnosed by a unique combination of character states, together with karyologic, morphometric, and molecular evidence.

*Cerradomys* species are characterized by a remarkable karyologic divergence (see Langguth and Bonvicino, 2002; Bonvicino, 2003); current recognized species, including C. vivoi and C. langguthi are clearly diagnosed by diploid and/or fundamental numbers, and by the morphology of autosomes and the sex chromosomes.

The combination of uni- and multivariate analyses confirms the existence of differences in the size and shape of cranial measurements among the valid species of *Cerradomys*: C. maracajuensis, C. marinhus, C. scotti, C. subflavus, C. vivoi, and Cerradomys langguthi. Regarding *Cerradomys subflavus*, C. vivoi, and C. langguthi, skull dimensions contributing significantly to this differentiation were related to the orbital region, auditory bulla, and cranial length and height. External measurements also separated the three species and, combined with skull dimensions, indicated a pattern in which *C. subflavus* is always larger and more robust than *C. vivoi*, whereas *C. langguthi* is consistently smaller.

Maximum-parsimony analysis clearly supported the monophyly of *Cerradomys*. This analysis also demonstrated the close relationship between *C. maracajuensis* and *C. marinhus*, and *C. scotti* as a sister branch with respect to the clade formed by *C. subflavus*, *C. vivoi*, and *C. langguthi*. This analysis also suggested that the ancestor of *C. subflavus* and *Cerradomys vivoi* was more recent than their common ancestor with *C. langguthi*.

Species of the genus *Cerradomys* inhabit open and drier biomes in South America, namely the Caatinga, Cerrado, and Chaco. Nevertheless, species of this genus are predominantly associated with the more mesic habitats of these biomes, such as gallery forests; humid forests formed by orographic rainfall, observed in hills and mountain ranges in the biome Caatinga (Mares et al., 1981).
### TABLE 7

Summary of Karyotypic Variation from Samples of the Six Species of *Cerradomys*

<table>
<thead>
<tr>
<th>Current taxonomy</th>
<th>2n</th>
<th>FN</th>
<th>Voucher specimens</th>
<th>Locality</th>
<th>Source</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>C. langguthi</em></td>
<td>46, 48–50</td>
<td>56</td>
<td>MN 69786</td>
<td>Tupanatinga, Bom Conselho, Capoeiras, Correntes, Panelas, Caruaru, São Lourenço, Exu, PE</td>
<td>Maia and Hulak, 1981; Bonvicino et al., 1999; this study</td>
</tr>
<tr>
<td><em>C. maracajuensis</em></td>
<td>56</td>
<td>58</td>
<td>MN 44178</td>
<td>Maracaju, MS</td>
<td>Bonvicino et al., 1999; Langguth and Bonvicino, 2002</td>
</tr>
<tr>
<td><em>C. marinhus</em></td>
<td>56</td>
<td>54</td>
<td>MN 63810–11, MN 63819, MN 66024, MN 63826, MN 63832, MN 63834</td>
<td>Jaborandi, BA</td>
<td>Bonvicino, 2003</td>
</tr>
<tr>
<td><em>C. scotti</em></td>
<td>58</td>
<td>70–72</td>
<td>MN 44176, MN 50380, MN 61667, MN 61672, MN 61674–80, MN 61683–87</td>
<td>Brasilia, DF; Cavalcanti, Alto Paraíso, Corumbá de Goiás, GO</td>
<td>Bonvicino et al., 1999</td>
</tr>
<tr>
<td><em>C. subflavus</em></td>
<td>54</td>
<td>62</td>
<td>CEG 42</td>
<td>P.N. do Rio Doce, MG</td>
<td>Bonvicino and Moreira, 2001</td>
</tr>
<tr>
<td><em>C. subflavus</em></td>
<td>54–56</td>
<td>62–63</td>
<td>–</td>
<td>Itapetininga, Paulinia and Santa Maria da Serra, SP</td>
<td>Almeida and Yonenaga-Yassuda, 1985</td>
</tr>
<tr>
<td><em>C. subflavus</em></td>
<td>54</td>
<td>62</td>
<td>UFPB 1926, MW13</td>
<td>Lagoa Santa, MG; Nova Viçosa, BA</td>
<td>Langguth and Bonvicino, 2002</td>
</tr>
<tr>
<td><em>C. vivoi</em></td>
<td>50</td>
<td>64</td>
<td>–</td>
<td>Brejo Grande, SE; Valença, Fazenda Unacau, BA</td>
<td>Andrades-Miranda et al., 2002</td>
</tr>
<tr>
<td><em>C. vivoi</em></td>
<td>50</td>
<td>62–63</td>
<td>MN 35898, MN 61661, MN 61662-05, LV–FC 22, LV–FC 148, MN 63377, MN 63381</td>
<td>Itabuna, Caetité, BA; Juramento, MG</td>
<td>this study</td>
</tr>
</tbody>
</table>
locally known as *brejos de altitude*; patches of more humid, closed canopy forest to drier, open-canopy woodland, locally known as *cerradão* (Eiten, 1972, 1992); and semideciduous forest. Exceptions to this pattern are the species that inhabit eastern Brazil: *C. subflavus, C. vivoi,* and *C. langguthi.* *C. vivoi* is a Caatinga-dwelling species that penetrates on Atlantic coastal rainforest in southern Bahia. The occurrence of *C. vivoi* in the latter habitat could be interpreted as a response to long-term, anthropic alteration of the habitat in this region (for example, extensive cacao plantations), since this species is found in low densities only in secondary and more open forest fragments, that occur marginally to more preserved fragments of Atlantic Forest (R. Pardini, unpublished data). A similar situation might be postulated for explaining the presence of *Cerradomys langguthi* (in this case, probably due to sugar-cane cultivation) and *C. subflavus* (probably due to extensive *Eucalyptus* plantations) in coastal Atlantic Forest in northeastern Brazil.

As the core species of the group in terms of geographic distribution (figs. 1, 2), *C. scotti* is

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### TABLE 8

Genetic Distance Estimates between Species of *Cerradomys*

<table>
<thead>
<tr>
<th>Taxon</th>
<th>Number</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
</tr>
</thead>
<tbody>
<tr>
<td><em>Cerradomys subflavus</em></td>
<td>LV–CEG 42</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>C. vivoi</em></td>
<td>LV–FC 148</td>
<td>0.020</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>C. vivoi</em></td>
<td>MN 35898</td>
<td>0.017</td>
<td>0.003</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>C. langguthi</em></td>
<td>MN 69786</td>
<td>0.046</td>
<td>0.057</td>
<td>0.057</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>C. scotti</em></td>
<td>MN 50379</td>
<td>0.085</td>
<td>0.084</td>
<td>0.085</td>
<td>0.087</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>C. scotti</em></td>
<td>MN 61677</td>
<td>0.084</td>
<td>0.087</td>
<td>0.084</td>
<td>0.086</td>
<td>0.001</td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>C. maracajuensis</em></td>
<td>MN 44178</td>
<td>0.122</td>
<td>0.122</td>
<td>0.119</td>
<td>0.127</td>
<td>0.112</td>
<td>0.111</td>
<td></td>
</tr>
<tr>
<td><em>C. marinhus</em></td>
<td>MN 63824</td>
<td>0.119</td>
<td>0.113</td>
<td>0.110</td>
<td>0.122</td>
<td>0.107</td>
<td>0.106</td>
<td>0.092</td>
</tr>
</tbody>
</table>

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Fig. 14. Maximum-parsimony topology of the phylogenetic analysis of all known species of genus *Cerradomys,* employing DNA sequence data of the mitochondrial gene cytochrome *b.* Numbers near nodes represent bootstrap values obtained on the basis of 1,000 replicates.
sympatric with almost all described species of the group: *C. maracajuensis* in Mato Grosso and Mato Grosso do Sul, Brazil, Sapucay, Paraguay, and Santa Cruz, Bolivia; *C. marinhus*, at the type locality of this species in Bahia; and samples of *C. subflavus* in Minas Gerais and São Paulo. However, there are some important differences among these species regarding habitat preferences in the Cerrado: *C. scotti* is associated more with the open vegetation of the Cerrado biome, namely the open and closed scrubs, both with scattered trees, locally known as campo cerrado and cerrado sensu stricto, respectively (Eiten, 1992), whereas *C. maracajuensis* and *C. subflavus* are associated more often with gallery forest and *C. marinhus* with grass marshes with buriti palms, called veredas (Eiten and Goodland, 1979), and flooded forests. At present, there is no evidence of sympatry between *C. vivoi*, *C. langguthi*, and other congeneric forms: the known distributional records of the first species are restricted to the habitats of Caatinga and Atlantic Forest biomes south and east of the Rio São Francisco; the available collecting localities for the second species are distributed on the Caatinga and Atlantic Forest habitats north to the Rio São Francisco.

On the basis of the evidence herein presented, the evolutionary differentiation of the genus *Cerradomys* can be somehow inferred. Species from eastern Brazil, *C. subflavus*, *C. vivoi* and *C. langguthi*, exhibit a clear trend of clinal variation, observed both in quantitative morphologic and karyologic traits. Samples of *C. subflavus*, the species with the southernmost distribution, present the largest dimensions in most external and cranial measurements, while samples of northernmost *Cerradomys langguthi* are characterized by the smallest values (table 2). Although some overlap is observed, average body and cranial dimensions diminish from south to north: geographic samples of *C. subflavus* are larger than those of *C. vivoi*, which are larger that those of *C. langguthi*. The same pattern of variation is observed regarding the diploid number (table 7), which decreases from south to north. In addition *Cerradomys subflavus*, *C. vivoi*, and *C. langguthi* also share a common ancestor. If this were accepted as indicative of sequence events since divergence, this clade is the most recent evolutionary branch of this group, a likely explanation for the striking morphologic resemblance among these species.

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REFERENCES


**APPENDIX 1**

**Gazetteer of *Cerradomys* Localities**

A list of the sampled localities of genus *Cerradomys* and their respective geographical coordinates are given below. Note that the numbers preceding collecting localities are the same presented in figures 1 and 2. For each locality we also provide the altitude, source for geographical coordinates, collector, collecting date, museum[s] or collection[s], and species obtained in each locality.

The gazetteer is arranged in alphabetical order, by country, state or province, and collecting locality.

LA PAZ


SANTA CRUZ
17. 6 km N of Buen Retiro, 17°13’S, 63°38’W; 300 m (collector). S. Anderson (col.), J.A. Cook (col.), September, 1984. AMNH. C. maracajuensis.
18. 6 km by road, W of Ascencion, 15°43’S, 63°04’W; 240 m (collector). S. Anderson (col.). AMNH. C. maracajuensis.
22. Ayacucho, 17°00’S, 63°55’W; 250 m (collector) [includes Ibanez, Ayacucho]. S. Anderson (col.), October, 1987. AMNH. USNM. C. maracajuensis.
24. Cordillera, Basilio, not located; the geographical coordinates of Basilio are 18°08’S, 63°19’W; 703 m. USNM. C. maracajuensis.


**BRAZIL**

**BAHIA**


38. Fazenda Santa Rita, 8 km E Andarai, Andarai, 12°48'06'S, 41°05'41"W; 399m (collector). L.P. Costa (col.), may, 1998. UFMG. C. vivoi.


**CEARÁ**


**DISTrito Federal (DF)**


55. Parque Nacional de Brasília, 15°40’S, 48°00’W (CDF; IBG, 1969) [includes Brasília]. A. C. Santos (col.), June–August, 1970. Antônio and
C. scotti


GOIÁS


59. 5 km N Alto Paraíso, Alto Paraíso, 14°05’S, 47°31’W (collector). C.R. Bonvicino (col.). MN. C. scotti.


MARAHÃO

66. Alto Parnayba (= Alto Paranaiba), 400–600m, 09°06’S, 45°57’W (USBGN, 1963); 400–600 m. E. Snethlage (col.), August, 1925. FMNH. C. langguthi.


68. Fazenda Lagoa Nova, Município Bacabal, not located. Here are employed the geographical coordinates of Bacabal, 04°14’S, 44°47’W (USBGN, 1963). MPEG. C. langguthi.

MATO GROSSO

69. 264 km N Xavantina, Serra do Roncador, 400 m, 12°49’S, 51°46’W (Musser et al., 1998). I.R. Bishop (col.), August, October, 1968 BMNH. C. maracajuensis, C. scotti.

70. Escola Evangélica Buriti, Chapada dos Guimarães, not located. Accordingly to the curator of UFMT mammal collection (M. Shiraiwa), this is an Evangelic School situated near the town of Chapada dos Guimarães, located at 15°26’S, 55°45’W (USBGN, 1963). J. Dalponte (col.). UFMT. C. scotti.


MATO GROSSO DO SUL

75. Fazenda da Mata, Maracaju, 21°38’S, 55°09’W (Langguth and Bonvicino, 2002); type locality of C. maracajuensis. C. R. Bonvicino and E.D.
Hingst (cols.), July, 1992. MN. *C. maracajuensis*.

76. Fazenda Primavera, Bataiporã, not located. Here are employed the geographical coordinates of Bataiporã, 22°20'S, 53°17'W (USBGN, 1963). L. E. Pereira (col.), fevereiro de 1993. MZUSP. *C. maracajuensis*.

77. Maracaju, 21°38'S, 55°09'W (USBGN, 1963); 500 m (collector). R.M. Gilmore (col.), M.E.S.-Serviço de Estudos e Pesquisas sobre a Febre Amarela (SEPFA)/The Rockefeller Foundation–International Division Health; March–July, 1937. AMNH, MN. *C. maracajuensis*.

**MINAS GERAIS**


79. [Área 23.] Usina Hidroelétrica de Miranda, Uberlândia, 19°05'S, 47°56'W; 710 m (collector). L.P. Costa (col.), August, 1998. UFMG. *C. scotti*.


85. Cerrado Fazenda Boa, BR 452, km 169, margem esquerda do Rio Araguari, not located. Here are employed the geographical coordinates of Indianópolis, 19°02'S, 47°55'W (USBGN database). G. Hermann (col.), October, 1987. UFMG. *C. scotti*.

86. Cerrado João Alonso, Perdizes, not located. Here are employed the geographical coordinates of Perdizes, 19°21'S, 47°17'W (USBGN, 1963) [includes Mata de galeria João Alonso, Perdizes]. UFMG. *C. scotti*.

87. COPASA–Área de Proteção Ambiental, Serra Azul, Mateus Leme, not located. Here are employed the geographical coordinates of Serra Azul, 20°04'S, 44°26'W (USBGN, 1963). April–May, 1986. UFMG. *C. scotti*, *C. subflavus*.


91. Fazenda Baroneza, Santa Luzia, not located. Here are employed the geographical coordinates of Santa Luzia, 19°47'S, 43°52'W (USBGN, 1963). Severino (col.), setembro de 1978. UFMG. *C. subflavus*.


93. Fazenda Canoas, 36 km NE, 12 km W Montes Claros (by road), Juramento, not located. Here are employed the geographical coordinates of Juramento, 16°50'S, 43°35'W (USBGN, 1963). Labvert (col.). MN. *C. vivoi*.

94. Fazenda Capão Grande, Santa Juliana, not located. Here are employed the geographical coordinates of Santa Juliana, 19°19'S, 47°32'W
C. scotti


97. Fazenda Esmeralda, Rio Casca, not located. Here are employed the geographical coordinates of Rio Casca, a municipality of Minas Gerais [and not the Rio Casca, a right bank tributary of Rio Doce], 20° 13′S, 42° 39′W (USBGN, 1963). December, 1985. UFMG. C. subflavus.


103. Lagoa Santa, 19° 37′S, 43° 53′W; 760m (USBGN, 1963) [includes Sitio no bairro Quebra, Lagoa Santa]; type locality of C. subflavus. A. Langguth (col.). UFPB. J.T. Reinhardt (col.), June–July, 1851. BMNH. C. subflavus. A. Langguth (col.).

104. Mata do Edésio, 8 km NW Nova Ponte, Nova Ponte, 19° 07′50″S, 47° 44′22″W; 854m (collector). L.P. Costa (col.), June, 1998. UFMG. C. subflavus.


106. Mata do João Lindolfo, 8 km NW Nova Ponte, Nova Ponte, 19° 07′10″S, 47° 43′53″W; 706 m (collector). L.P. Costa (col.), June, 1998. UFMG. C. subflavus.


110. Parque Estadual do Rio Doce, 13 km E Marliéria, 19° 34′S, 42° 39′W; 300m (collector) [includes Parque Estadual do Rio Doce, Marliéria]. L. Paulo (col.), December, 1986. C. subflavus.


114. Reserva do Jacob, Nova Ponte, not located; here are employed the geographical coordinates of Nova Ponte, 19° 08′S, 47° 40′W (USBGN, 1963). R. Moura and E. Sabato (cols.), March, 1996. C. maracajuensis.


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120. Vargem do Retiro, Ribeirão Mascates, Parque Nacional da Serra do Cipó, 800m; not located. Here are employed the geographical coordinates of Ribeirão Mascates, 19°20'S, 43°36'W (USBGN, 1963). MN. C. subflavus.

PARAÍBA


PERNAMBUCO


C. maracajuensis
ca.
C. scotti
C. langguthi
C. subflavus
S

PARAGUAY

Amambay


Caaguazu


Canendiyu


151. Villa Igatimi [also spelled Ygatimi or Icatimi], 24°05'S, 55°30'W, 151 m (USBGN, 1957). J. Contreras (col.). MLP. C. maracajuensis.

Concepcion


Cordillera


Paraguari

don near growth of *Bromelia balansae*” (collector). C. Wharton (col.), April, 1950. USNM. *C. maracajuensis*.

**SAN PEDRO**


**PERU**

**PUNO**
